

35 frying, and others (Obidiegwu 2020; Singh and Twinkle 2022). The yam therefore
36 constitutes an essential source of food, employment, income and a vital component of food
37 security in the region (Zangoma et al 2025).

38 The yam tuber contains within it, a functional dynamism responsible for its evolution from
39 initiation to the next dormancy break, passing successively through stages of growth, maturity,
40 and dormancy. During the growth phase, the histological study conducted by Degbeu et al.
41 2019 showed the existence of a longitudinal growth gradient whose point of growth is the apex
42 and the sense of maturity of the distal part at the proximal end. The cambium generates cells
43 at the lower end of the apex. These very vacuolated cells differentiate and will form the median
44 and proximal parts of the tuber. This cambium generating activity would cease when the tuber
45 become matures. The existence of this gradient has an impact on the technological suitability
46 of the different parts of the yam tuber. Thus, the suitability for cooking decreases from the
47 proximal part to the distal end. Firmness and dry matter decrease from the proximal part to
48 the distal end of the tuber (Brunnschweiler, 2004).

49 At maturity, the tuber enters a period of dormancy, defined as the period during which
50 endogenous metabolic activity is reduced and there is no intrinsic bud growth, although it
51 retains the potential for future growth (Ile et al 2006, Nwogha et al 2022). The duration of
52 dormancy depends on the species, storage conditions, weather conditions and the degree of
53 maturity of the tuber or the time of tuber harvest (Hamadina 2011; Nwogha 2022; Zangoma
54 et al 2025). In general, *D. alata* varieties have a long dormancy period (three to four months)
55 compared to those of *D. cayenensis-rotundata* (one to one and a half months) (Craufurd et
56 al 2001; Wickham 2019). Early varieties harvested in August begin to break dormancy in
57 January. This is four to five months after harvest. Once dormancy is broken, the shoot
58 develops, producing leaves and branches, leading to the plant's volubility. At the same time,
59 the underground roots develop and the mother tuber that gave rise to the plant begins to
60 degenerate (Trousnot 1985).

61 The initiation of tuberization is identified by the appearance of "suber bursting" at the base of
62 the stem in plants propagated by tuber fragments. In yam plants grown from seed, the initiation
63 of tuberization is manifested by a bulge located between the starting point of the main root
64 and the base of the first scale leaf of the main axis (Trousnot 1985). Micro-plant studies have
65 shown that the Microtuberization is affected by plant growth regulators such as Naphthalene
66 acetic acid, gibberellic acid and jasmonic acid (Balogun 2006; Hamadina et al 2010).

67 Since dormancy breaking is the first stage in the natural regeneration process of the tuber, its
68 variability could impact the second stage, which is the initiation of tuberization. This is the
69 research hypothesis of the present study, the objective of which is to understand the origin of
70 the precocity of the Kponan yam variety. In this study, the morphological aspect of tuberization
71 of two yam varieties (Kponan and Kangba) of the species *Dioscorea cayenensis-rotundata*
72 was analyzed. The Kponan variety is early maturing and the Kangba variety is late maturing.
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74 2. MATERIAL AND METHODS

75 2.1 Materiel

76 The kangba and kponan varieties of yam, belonging to the species *Dioscorea cayenensis-*
77 *rotundata*, were used as plant material. The fragmentation process produced pieces of yam
78 with an average mass of 137.3 ± 35.6 g for the kponan variety and 93.5 ± 28.9 g for the kangba
79 variety. For this study, the head of the tuber was avoided as much as possible to ensure
80 uniformity of emergence. The seedlings are at the end of dormancy, i.e., at the beginning of
81 germination. Most of them have a bud characteristic of dormancy emergence. Three samples
82 with shoots measuring 2.5 cm, 3 cm, and 12 cm, respectively, were observed for the kponan
83 variety
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85 2.2 Methods

86 2.2.1 Setting up the plots

87 The field was created at Nangui Abrogoua University on an area of 50 m². The land is fallow
 88 soil. During periods of low or no rainfall, the plants were watered once or twice a week as
 89 needed. The bags were filled with forest soil enriched with pig manure. The stakes used were
 90 Chinese bamboo placed between the rows of bags. The bags were used to ensure the even
 91 distribution of soil nutrients and to allow each plant to have the same nutrients. The seeds
 92 were planted in 20 cm x 20 cm nursery bags and arranged in rows 1 m apart, with 0.5 m
 93 between bags. After the yams were bagged, the field was visited weekly to monitor “field
 94 emergence,” i.e., the appearance of the bud above the mound (Trouslot, 1985), in this case
 95 the bag. Once the bud has appeared above the bag, the date is recorded and will serve as a
 96 reference point for monitoring tuberization.

97 98 **2.2.2 Sampling technique**

99 The tubers were harvested at different times by cutting them with a stainless-steel knife. This
 100 allows the tuber to be separated from the pre-tuber. The tubers were harvested at the
 101 beginning of tuberization at the stage when the tuber bursts, which is the first visible sign of
 102 tuberization (day 0; day 7; day 15; day 30 and day 40). Monitoring then took place from the
 103 16th week after the seeds were bagged until the 26th week, i.e., weeks 16, 18, 20, 22, 24, and
 104 26. Week 26 corresponds to the onset of leaf yellowing. At each harvest period, the size of
 105 the tuber was measured using a Mitutoyo caliper (Braive Instrument, Belgium) and the mass
 106 was determined using a Scaltec scale (Heiligenstadt, Germany).

107 108 **3. RESULTS**

109 **3.1 Sprouting period of yam cutting after planting and tuber initiation**

110 Three cutting samples of Kponan had buds measuring 2.5 cm, 3 cm, and 13.3 cm,
 111 respectively, or 1, 3, and 6 scale leaves on the day of planting on 20 plants of the Kponan
 112 variety. For these three samples, growth resumed 16, 9, and 6 days after planting,
 113 respectively. The emergence period for these three plants appears to be scattered. This
 114 represents an average of 10 days \pm 5. Most emergence ranges from 22 days \pm 2 to 35 days
 115 \pm 5. Late emergence 56 days after planting was observed (**table 1**). For the Kangba variety,
 116 cutting without buds, on the day of planting, show emergence 34 days \pm 3 to 45 days \pm 2 after
 117 being placed in bags. Early emergence was observed at 27 days in two plants.

118 From the date of emergence in the field, the first visible sign of tuberization in the Kponan
 119 variety appears between 48 \pm 4 and 57 days \pm 3. Unlike the emergence period in the field, the
 120 bursting stage is not dispersed. The average is around 53 days \pm 5. Plants with late emergence
 121 initiate tuberization earlier, at 32 days after emergence in the field. For the kangba variety, this
 122 period extends from 48 \pm 2 days to 71 days \pm 4 after emergence in the field. This period is
 123 longer compared to observations made on the kponan variety ($p < 0.05$).

124 In general, after planting on April 14, 2007, tuber bursting appears from 71 days \pm 5, or 10
 125 weeks to 86 days \pm 3 (12 weeks) in the kponan variety. In the kangba variety, this period is
 126 around 95 days \pm 4 after planting, or 13 weeks.

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128 **Table 1:** Sprouting period of yam cutting and tuber initiation

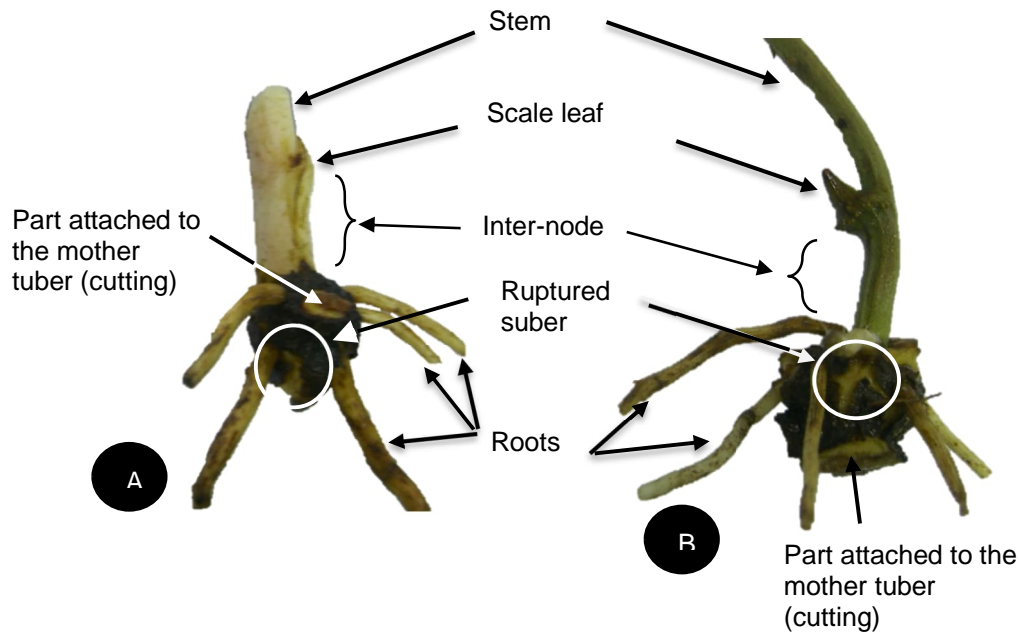
varieties	Appearance of sprouting (number of days)	Tuber initiation	Planting date-tuber initiation
Kponan	10 days \pm 5 (3 budding plants)	57 days \pm 3	71 days \pm 5 to 86 days \pm 3
	28 days \pm 1 (2 ungerminated plants)	55 days	
	22 days \pm 2 (6 ungerminated plants)	53 days \pm 7	
	35 days \pm 5 (5 ungerminated plants)	48 days \pm 4	
	56 days (1 ungerminated plants)	32 days	
Kangba	27 days (2 ungerminated plants)	71 days \pm 4	95 days \pm 4
	34 days \pm 2 (7 ungerminated plants)	62 days \pm 5	
	43 days \pm 2 (7 ungerminated plants)	50 days \pm 3	

45 days ± 2 (4 ungerminated plants) 48 days ± 2

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3.2 Development of the emerging tuber

The first visible sign of tuberization is the rupture of the suber covering the base of the stem (Fig. 1A and 1B). Over the following days, the bursting spreads as the proliferation of the underlying tissue intensifies. The suber disintegrates and disappears under the effect of this proliferation at the apical end of the tuber (Fig. 1C and 1D). At the beginning of tuberization, new roots appear. These roots have a white membrane that is not suberized, unlike the first roots. From the 7th day onwards, a tuber bud is visible (Fig. 1C and 1D). The base of the stem begins to develop into a new tuber or neo-tuber, while the mother tuber (cutting) shrinks (Fig. 1A) and eventually degenerates (Fig.1B). It will form the pre-tuber at harvest. In the Kponan variety, the onset of tuberization is accompanied by total degeneration of the yam cutting. This is not the case with the Kangba variety, where the rest of the seed is still visible. The yam cutting of the Kangba variety has simply shriveled. In addition, the arrangement of the new roots differs between the two varieties. In Kangba, the new roots appear on the new tuber (Fig. 1C). This is not the case with the Kponan variety (Fig. 1E). Seven days after the start of tuberization, the new tuber of the latter has a bulbous shape characteristic of a tuber. In the Kangba variety, however, the arrangement of the new roots on the new tuber is characteristic of a woody mass. The apical end of the developing tuber is light yellow in both the Kponan and Kangba varieties. It is covered with a cap. From the seventh day onwards, the organ thus formed hypertrophies longitudinally and radially to produce a more rounded tuber on the 21st day (Figs. 2A and 2B). Three weeks after the start of tuberization, the mother tuber disappears completely. The protrusion at the base of the stem is more intense at the apical end and is responsible for the tuber's grown significantly in length. This results in a cylindrical-conical tuber with a rounded and slightly tapered apical end, reflecting the predominance of apical meristem activity. As the tuber's growth in length intensifies, suberization of the outer layer or skin of the yam can be observed at the proximal end. The intensity of this suberization decreases from the proximal part toward the distal end. Suberization is more pronounced in the Kponan variety at the proximal part of the tuber at 18 weeks after planting (Fig. 2C). It extends to the entire tuber at 22 weeks after planting (Fig. 2E). In the Kangba variety, suberization begins in the proximal part at 18 weeks after planting (Fig. 2D). However, this phenomenon is more visible at 22 weeks after planting (Fig. 2F). The distal end remains unsuberized.



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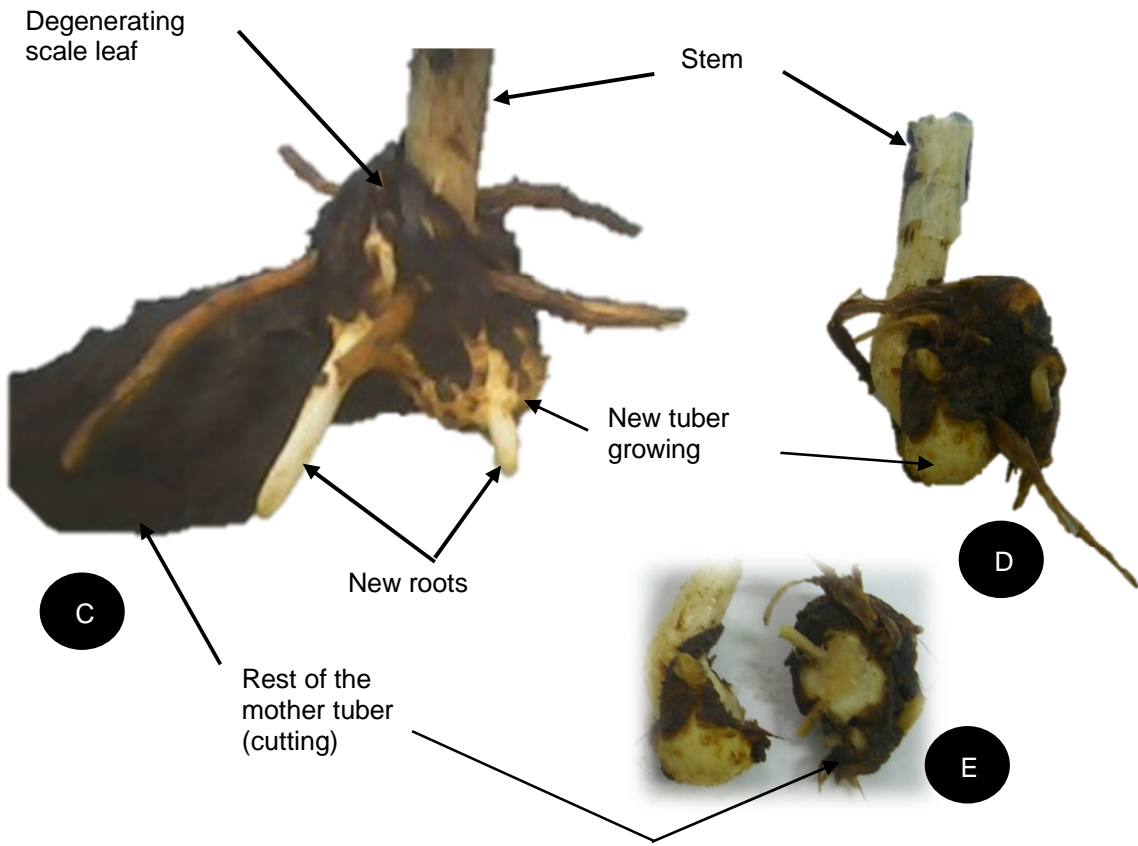
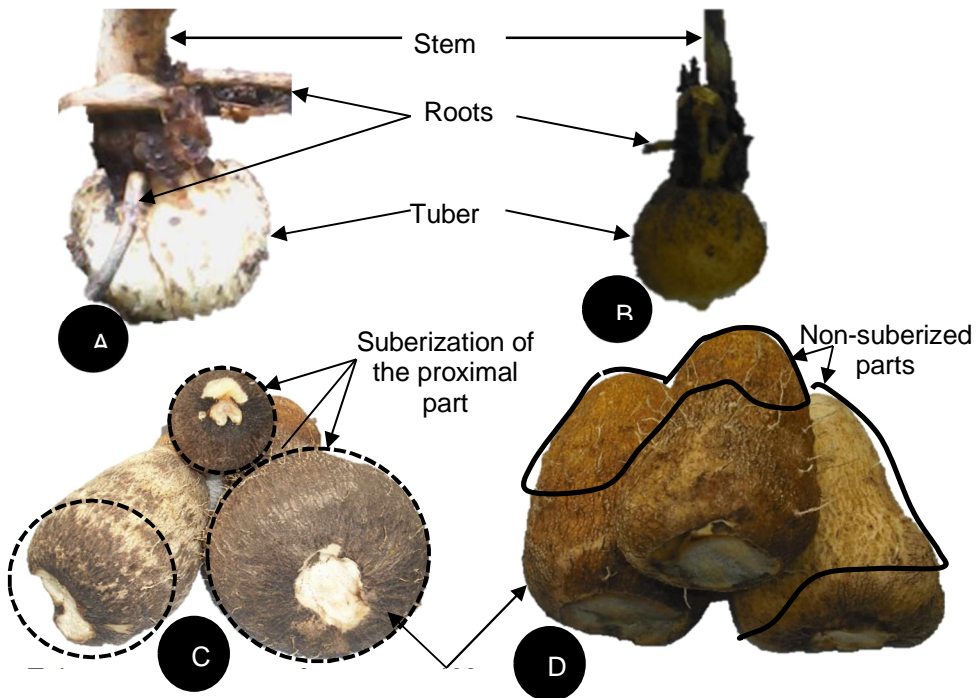
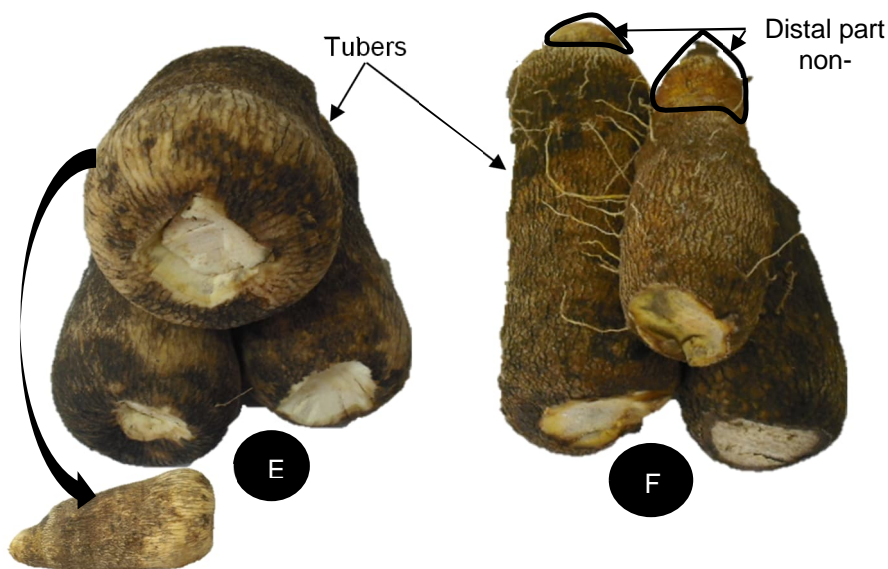


Fig. 1. Earlier stage of tuberization. The tuber initiation is materialized by the rupture of the suber (A, B). The following days, the protuberance of meristem cells lead to the tuber formation and growing observable seven days after rupture of the suber (C, D). The cutting is observable for var. kangba (C), not for var. kponan (E). Scale bars: 1 cm



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Fig. 2: Yam tubers at different stages of maturity. A, B are respectively tubers of. var. Kponan and Kangba at twenty-one days after the rupture of the suber. C and E are tubers of var. Kponan at respectively eighteen and twenty-two weeks after the cutting were bagged. D and F are tubers of var. Kponan at respectively eighteen and twenty-two weeks after the cutting were bagged.

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3.3 Evolution of the size and mass of the emerging tuber

From the seventh day onwards, the mass of the tuber formed is around 1.56 ± 1.2 g and 1.35 ± 0.1 cm in length (Kponan variety) (Table 2). The Kangba variety has approximately the same mass and size (1.47 ± 1.1 g A; 1.2 ± 0.3 cm). Two weeks after the start of tuberization, the mass of the Kponan variety tubers is higher than that of the Kangba variety. From the 21st day after the start of tuber initiation, the mass of the tubers was greater in the Kponan variety (15.3 ± 4 g to 36.25 ± 5.3 g). The size remained the same.

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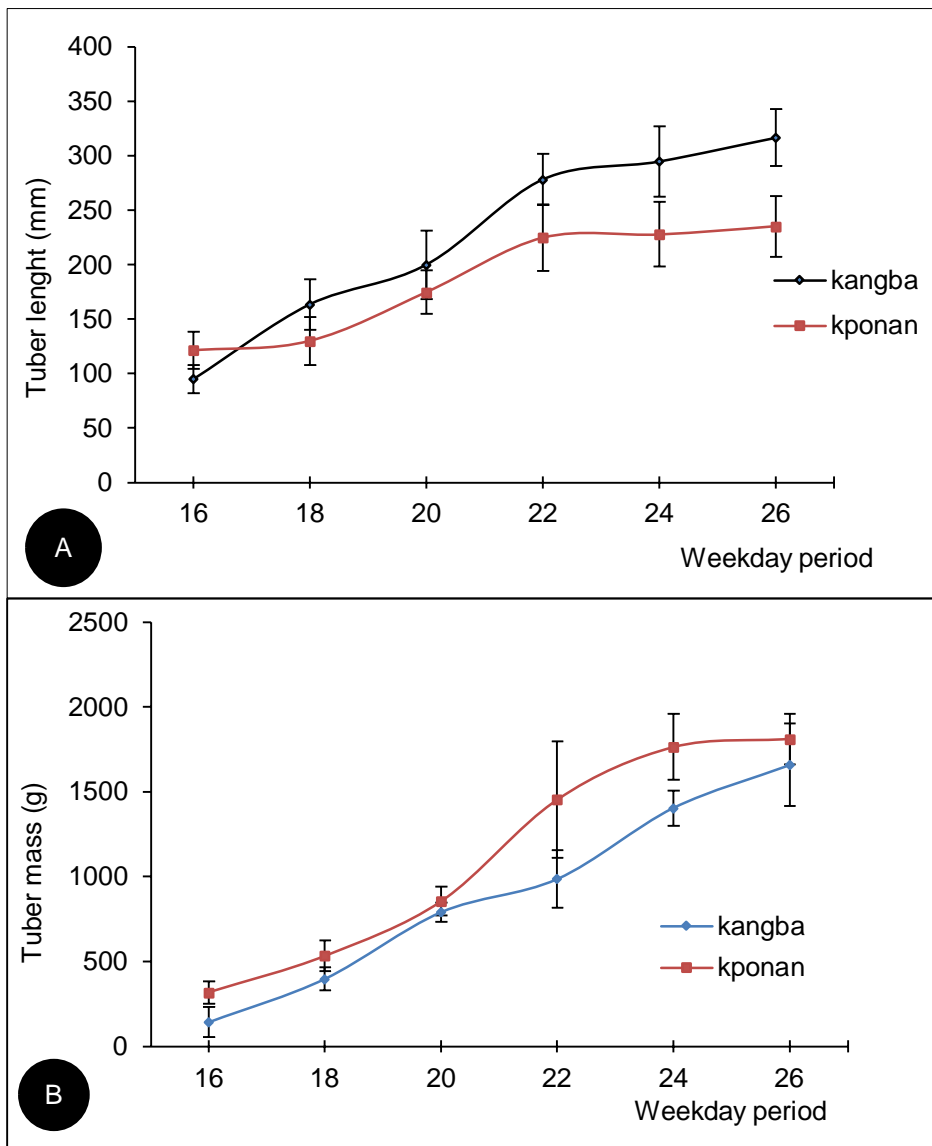
Table 2: Changes in tuber mass and length over 40 days after the suber burst stage

varieties	Days	Mass (g)	Length (cm)
Kponan	7	$1,56 \pm 1,2$	$1,35 \pm 0,1$
	15	$4,5 \pm 1,8$	$2,02 \pm 0,2$
	21	$15,3 \pm 4$	$3,2 \pm 0,3$
	30	$36, 25 \pm 5,3$	$5,2 \pm 1,4$
	40	$121,5 \pm 26,6$	$9,4 \pm 0,11$
Kangba	7	$1,47 \pm 1,1$	$1,2 \pm 0,3$
	15	$3,18 \pm 1,3$	$2,1 \pm 0,1$
	21	$10,45 \pm 3,4$	$3,02 \pm 0,5$
	30	$31,73 \pm 5,1$	$4,6 \pm 1,8$
	40	$114,49 \pm 13$	$12,3 \pm 2$

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270 3.4 Changes in tuber weight and length from the sixteenth week after planting

271 The evolution of tuber length and mass was monitored starting in the second month after the
 272 first visible sign of tuberization appeared, i.e., 16 weeks after planting (Fig. 3A). Tuber growth,
 273 which began in the first few weeks after the suber burst, intensified until the 22nd week after
 274 sowing, i.e., three months after the start of tuberization. From this period onwards, tuber
 275 growth slowed down and tended to stabilize until the 26th week after planting. The growth of
 276 kponan yam tubers is greater until the 17th week. Beyond this period, the length growth of
 277 kangba tubers becomes more significant, until tuber growth stops at the 26th week. As for the
 278 evolution of tuber mass (Fig. 3B), it becomes significant from the 20th week onwards. This
 279 increase in tuber mass tends to stabilize from the 24th week onwards for the kponan variety.
 280 For the kangba variety, tuber mass continues to increase even beyond week 26. Comparison
 281 of the average tuber mass of the two varieties shows a significant difference ($p < 0.05$).
 282



321 **Fig. 3:** Changes in tuber length and mass during tuberization. A) Tuber length; B) Tuber mass

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323 4. DISCUSSION

324 The development of the tuber was monitored from the emergence of the main stem above the
325 bag to the formation of the tuber. The three plants bearing a bud on the day of planting indicate
326 a resumption of stem growth 6 to 16 days after planting, or an average of 10 days \pm 5. The
327 resumption of stem growth depends on the level of development of the preformed bud. Based
328 on observations, the more developed the bud, the earlier the resumption of growth. However,
329 in practice, shoots that are too long are removed by farmers, as the tuber bearing the bud
330 would be weakened by excessive depletion of its reserves (Trousnot 1985). The period of
331 dormancy in the field for most yam cuttings of the two varieties Kponan (22 to 35 days) and
332 Kangba (34 to 43 days) falls within the range (14 to 91 days) reported by Hamadina (2011).
333 This period varies greatly according to previous studies and is influenced by regulatory
334 hormones (Nwogha 2022; Zangoma et al. 2025). Cuttings of the Kponan variety germinate
335 quickly compared to those of the Kangba variety. This could be due to varietal differences
336 (Wickham 2019). In *in vitro* yam stem fragment cultures, tuberization begins after three weeks
337 under normal conditions and one week when phytohormones are used (Jova et al., 2011).
338 From the date of planting to the initiation of tuberization, cuttings of the Kponan variety tuberize
339 quickly compared to those of Kangba. This could partly explain the earliness of the Kangba
340 variety. Visual observation of the onset of tuberization also shows that the tuber used as a
341 cutting disappears in Kponan, while it is still present in the Kangba variety. This could also
342 explain the earliness of Kponan. Similarly, the fiber content of the mother tuber could have an
343 impact on the earliness of tuberization initiation. Fiber-rich tubers such as Kangba tend to
344 shrivel, thus delaying this phenomenon. Those of the Kponan variety, being less fibrous, rot
345 quickly and tuberize sooner. The same applies to morphological observations of the different
346 stages of tuberization, where the tuber becomes corky earlier in the Kponan variety than in
347 the Kangba variety. However, suberization is characteristic of tuber maturity, based on
348 observations. At 22 weeks after planting, the tubers of the Kponan variety are completely
349 suberized, while those of the Kangba variety show a growing distal end. The Kponan variety
350 produces early tubers that are larger in mass than those of the Kangba variety. The Kponan
351 yam tuber is morphologically shorter and has a larger diameter than the Kangba tuber, which
352 would explain, on the one hand, the higher position of the Kangba variety on the length growth
353 curve and, on the other hand, the higher mass of the Kponan variety tubers.

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355 4. CONCLUSION

356 Generally speaking, after planting on April 14, 2007, the tuber bursts after 71 days \pm 5, or 10
357 weeks for the Kponan variety, and 95 days \pm 4 (13 weeks) for the Kangba variety. The mass
358 growth of tubers of the Kponan variety is stronger than that of the Kangba variety. The Kponan
359 variety initiates tuberization and early maturity compared to Kangba.

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