****Effects of the**** RNA****interferencecassette targeting the**** *Puroindoline******a* on grain hardness and**** Puroindoline****protein in soft wheat****

Abstract

Kernel hardness,**a critical quality trait in wheat, significantly determines its processing performance and end-use baking properties.**The *Pina* gene is controlled by the *Ha* locus on chromosome 5D and is the primary gene responsible for determining wheat kernel hardness. If there is a mutationin*Pina* or *Pinb* gene, the wheat is hard kernel. In this study, **we introducedaRNA interference vector targeting the *Pina* and its linear expression cassette into soft wheat Emai 12 via particle bombardment.**For transgenic lines identified as PCR-positive, grain hardness may increase, while Puroindoline protein content may decrease. Thesestudies can help researchers deeply understand the grain hardness quality of wheat, thus facilitating the improvement of wheat quality.

**Key words**:*Puroindoline*; kernel hardness**;**linear expression cassette; transgenic lines;**RNA interference.**

Wheat is one of the world's three major food crops and**its demand continues to grow substantially**. **The hardness of wheat kernels**is one of the key traits determining wheat milling quality, primarily **determined by the *Puroindoline* (*Pin*) genes,** which include***Pina***and***Pinb*,mapped to the *Ha* locus on chromosome 5D** (Glenn and Saunders 1990; Gaines et al. 1996; Giroux and Morris 1998, Wanjugi et al. 2007,[Jolly et al. 1993](file:///D:\李小燕文档-G\Pin基因多样性投稿\genes\manuscript20180906.docx#_ENREF_36))**.The** Pina**and** Pinb**genes** encode**PINA and PINB proteins,** which are present on the surface of starch granules in the**developing endosperm,thus responsible for the soft grain texture in wheat.** PINA and PINB are two isoforms of proteins with an amino acid sequence similarity as high as 68%(Shewryet al. 2004; Gautier et al. 1994).**Composed of 145 amino acids, PINs possessed a relatively low molecular weight and were rich in cysteine and tryptophan.Ifa mutation is present in either or both wild-type alleles these two genes, the grain hardnessincreases, resulting in a hard kernel wheat (**Giroux and Morris1998;Beecher et al. 2002; Morris 2002**). For instance, a non-functional mutation in**Pinbleads to**reduced or absent accumulation of PINB protein**onstarch granules,**thereby resulting in the development of medium-hard or hard-textured wheat. The** Pina-D1b**allelic mutation,** characterized by**the absence of PINA protein, results in significantly harder wheat kernel texture**(Gazzaet al. 2005).**The *Pins* determine wheat grain hardness.** Differences in Pin genotypes among wheat varieties can lead to significant variations in grain hardness values (Giroux and Morris 1998; Morris et al. 2001a, b).**The presence of these proteins at the starch granule surface weakens the strong interfacial interactions between starch and polar lipids, resulting in varying degrees of grain hardness(**Ali et al. 2015; Geneix et al. 2015**).In a study，overexpression of the** Pina **gene, initially intended to increase PINA protein expression, ultimately triggered the co-suppression phenomenon of the endogenous** Pina **gene, counterproductively reducing its expression**(Xia et al. 2008)**.**Krishnamurthy discovered that introducing Puroindoline proteins into rice plants enhances their antimicrobial capacities, and further proposed that this might be related to the protein's ability to disrupt cell membranes (Krishnamurthy et al. 2001). Faizefound that introducing thePingene into apple significantly improved its resistance to Scab; further experiments demonstrated a clear negative correlation between the susceptibility of transgenic apple plants and the content of PinB protein (Faize et al. 2004). Luo *et al* introduced the Pina gene into durum wheat varieties Luna and Venusia2(Luo et al. 2008). They observed significantly enhanced resistance to leaf rust in transgenic wheat plants under both greenhouse and field conditions, along with a substantial increase in yield. These findings demonstrate that the PINA protein possesses a clear fungal-inhibitory function.

Previous studies have shown that **overexpression of the *Pina* gene** not only softens wheat kernels but also **worsens the dough mixing properties of flour (L**i et al. 2014; Heinze et al. 2016; Martin et al.2007). Based on the research evidence**, targeted RNA interference (RNAi) of the** Pina**gene alone can indeed transform soft wheat into harder wheat**, but the resulting hardness variation may be limited due to compensatory mechanisms and genetic background effects.

***NB>Author needs to work on his/her introduction***

The aim was to identify varieties with high grain hardness, provide elite parental materials for breeding new high-hardness wheat varieties, and determine the optimal gene combinations for high grain hardness. This will enable breeders to make effective selections in early generations through molecular markers. The results of this study will have significant implications for improving wheat variety quality.

**Methods and material**

**Plant materials**

**Linearized expressionconstruct and genetic transformation**

The interfering element in the pAHC25-*Pina*-RNAi vector used in the experiment consists of sense and antisense sequences of Pina joined by an intron (sequence detailed in Figure S1) combined with a **ubiquitin (ubi)**promoter and**a nopaline synthase (Nos)** terminator**. The linear expression cassettewere released from the parental vector by restriction digestion, containing exclusively the promoter, sense sequences, an intron, antisense sequences, and terminator.** The structure is illustrated in Figure 1.To verify the vector construct, we digested it using **HindIII**restriction sites flanking the interfering elementand the digestion results were depicted in the Figure 2.

**Regeneration of transgenic plants and PCR detection**

The pAHC25-*Pina*-RNAi vector and itslinear expression cassette were introduced into the scutellum of soft-grained wheat (*Triticum aestivum* L. cv. Emai 12) using particle bombardment technology. Through wheat tissue culture, transgenic regenerated seedlings were obtained, and PCR detection of foreign gene fragments was performed on the genomic DNA of the T0 generation. **Seeds of T0 transgenic positive plants were germinated in a phytotron to generate T1 regenerated plants, followed by PCR analysis of the T1 generation plants.Genomic DNA was extracted with the** Cetyltrimethylammonium Ammonium Bromide (**CTAB) method to detect whether the Pina vector and its linear expression cassette were successfully transferred into transgenic regenerated plants**(Giroux andMorris 1998)**.**Thenested PCR and semi-nested PCR were employed toconfirmthe intact integration of the Pina vector and its linear expression cassette in the genome of regenerated plantsraise from poor sequence specificity. The PCR amplification was conducted in a 25 µl reaction volume containing 90 ng of genomic DNA, 0.15 μM primers, and 12.5µl2×Es TaqMasterMix(Tiangen, Beijing, China).The program is, denaturing at 94 ℃ for 5 min, following by 36 cycles of94 ℃ for 30 s, 59 ℃ for 15 s and 72 ℃for 25 s , with a final extension of 4 min. Then **the PCR products were sent to AuGCT Biotechnology** company**(Beijing, China) for sequencing to determine whether the sequencing results matched the target sequence.**

**Hardness measurement**

**Grain hardness was performed on T₂ seeds harvested from PCR-positive T₁ seedlings of wheat.The**grain hardness of 10 mature seeds harvested from each **transgenic regenerated line**were measuredusing the Perten Single Kernel Characterization System 4100 (Perten Instruments,Inc., Springfield,IL, USA) according to the approved method of AACC N°55-31.The wheat Emai 12 is soft-textured wheat with kernel hardness index (HI) ≤40. If the **RNA interference** is successful, the kernel hardness of transgenic regenerated wheat plants will be harder, exceeding 40 on the hardness scale.

**SDS-PAGE and Western blotting**

**For seeds exhibiting a certain increase in grain hardness value, total seed proteins were extracted using the half-seed technique.** The total seed proteins were extracted according toHe’s method (He et al. 1999).Their separation wasmanipulated using a 12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) to detect whether the total proteins correspond to that of the *Triticum aestivum* L. cv. Emai 12 positive control seeds (Figure S2). Then the proteins were transferred onto a nitrocellulose membrane and blocked with Buffer A (0.1% Tween-20, 5% non-fat milk, 1\*TBS)for two hours. The membrane was incubated with PINA and PINB antibodies diluted 2,000-fold at room temperature for two hours, respectively. The membrane was washed twice with TBST and incubated with a goat anti-rabbit secondary antibody diluted 2,000-fold at room temperature for two hours, followed by twice washes.The PINA antibody was developed by Miao, while the PINB antibody was prepared by Li (Li et al. 2019). The positive control for this Western blotting was the wheat variety Emai12 with the wild-type *Pina-D1a*/*Pinb-D1a* genotype.Quantitative analysis of PINA and PINB proteins was performed using the Bio-Rad ChemiDoc XRS+ system.

**Results**

**The PCR identification of transgenic regenerated plants**

The first set comprised semi-nested primers, including the outer primer pair D-F/Nos-R and the inner primer pair Nos-F/Nos-R (primers detailed in Table 1), ultimately produced a target band of 253 bp (Figure 3).The second set consisted of **nested primers,** including the **outer primer pair**Inno-F/Inno-R and the **inner primer pair**D5-F/D5-R, yielding a final PCR product of **510 bp** amplified by the D5 primers.

**Following PCR screening across T0 and T1 generations,**the **pAHC25-*Pina*-RNAi vector**yielded**four putative transgenic-positive plants,** while its**linear expression cassette**produced**three putative transgenic-positive plants.**

**kernel hardness**

The selected controls included wild-type Emai 12 seeds, tetraploid durum wheat Luna, and transgenic-negative plants, all cultivated simultaneously in the same climate chamber to ensure consistent environmental conditions. The kernelHI of these seeds was presented in Table 2. The data indicated that certain transgenic-positive lines exhibited higher kernel HI compared to both wild-type and transgenic-negative controls. **The** kernel HI **for E2 transgenic line of pAHC25-*Pina*-RNAi exhibited a significant increase in grain hardness, while the E3 transgenic line showed a slight increase. In contrast, the E1 and E4 lines of pAHC25-*Pina*-RNAi displayed almost no change in grain hardness. Among these transgenic lines transformed with the linear expression cassette, the L1 line demonstrated a marked rise in kernel HI, the L3 line a marginal increase, and the L2 line no significant alteration.**

**Western blotting of Puroindoline**s

SDS-PAGE analysis was performed to determine whether the protein profiles of transgenic-positive wheat seeds were consistent with those of wild-type Emai 12 seeds. As shown in Figure S2, the protein bands of transgenic wheat seeds were identical to those of wild-type seeds.

Analysis of total seed proteins using PINA- and PINB-specific antibodies revealed distinct expression patterns across transgenic lines, as shown in Figure 4. In the E3 transgenic line, PINA protein expression was reduced in some seeds, with several wells showing no detectable expression. Conversely, PINB protein expression increased correspondingly in these seeds. Similarly, in the L2 transgenic line, a decrease in PINA expression coincided with a compensatory rise in PINB levels.

**Discussion**

Genetically, wheat kernel hardness is primarily controlled by the *Pin* gene, while environmental factors also exert considerable influence (Qury et al.2015). Therefore, when comparing transgenic-positive seeds with transgenic-negative and wild-type wheat seeds, it is essential that they are cultivated under identical controlled greenhouse conditions. This approach helps minimize environmental impacts on their comparative analysis to some extent.The kernel hardness index for T2-generation transgenic-positive seeds are presented in Table 2. Compared to the control lines, the kernel HI of E2 and L1 lines showed an increase of 20. These lines may not yet be homozygous, which likely contributes to their relatively large standard deviations. The E3 and L3 lines both exhibited small variation in kernel hardness values, likely reflecting an average measurement resulting from the mixed testing of both transgenic-positive and non-transgenic seeds.This aligns with previous findings by Gasparis, demonstrating that silencing of the ***Pina*** gene can increase thekernel HIof soft wheat(Gasparis et al. 2011).

The quantity of PIN protein in T2-generation transgenic-positive seeds is shown in Figure 2. A common phenomenon observed in both lines 1 analyzed is that when the expression level of PINA protein decreases or is even absent, the expression level of PINB protein exhibits a compensatory increase. **This may indicate a compensatory effect between PINA and PINB proteins, but there is still insufficient experimental evidence to confirm it. This phenomenon differs from what was reported in Sebastian's study on *Pin*RNAi, which did not mention such compensatory effects. Therefore, it might represent a phenomenon observed only in a limited number of lines**(Gasparis et al. 2011).**The grain hardness of transgenic line E3 increased significantly, which may be attributed to the complete suppression of PINA protein expression in some seeds. According to previous studies, when one of the PIN proteins is not expressed, wheat grains exhibit a hard phenotype (Morris 2002). Therefore, even though PINB protein expression was upregulated, the complete interference of PINA protein ultimately led to the hard-grain phenotype in wheat.The grain hardness of transgenic line L2 showed minimal change, likely because although PINA protein expression was reduced in some seeds, the compensatory** increase **of PINB expression resulted in almost no phenotypic alteration.** Whether it is an RNA interference vector or its linear expression cassette, when introduced into wheat, either may lead to a decrease in the PINA protein content. Moreover, a reduction in PINA content might lead to a compensatory increase or decrease in PINB protein.

**Conclusion**

The **pAHC25-*Pina*-RNAi vector** and its **linear expression cassette** were introduced into the common soft-kernel wheat cultivar *Triticum aestivum* L. cv. Emai 12 using the **gene gun bombardment. The identification of four pAHC25-*Pina*-RNAi positive lines and three linear expression cassette lines was achieved by amplifying regenerated plants via nested PCR and semi-nested PCR.The kernel HI of some transgenic seeds fromRNAi vector or linear expression cassette** exhibited a certain degree of elevation**, with some variation observed across different transgenic lines.**The interference of*Pina* demonstrated that the *Pina* gene is functionally associated with grain hardness once again. Western blotting analysis revealed the presence of completely silenced PINA protein lines in the progeny of transgenic plants harboring the ***Pina* interference vector.** Furthermore, in transgenic lines where PINA protein content was reduced, PINB protein content may**compensatorily**increase or decrease.

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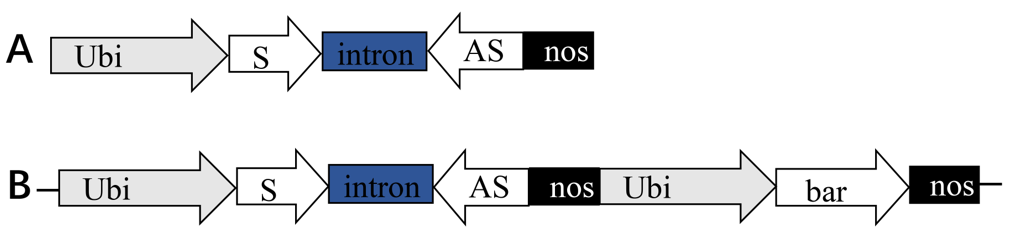


Figure 1 Composition of **pAHC25-*Pina*-RNAi (B) and its linear expression cassette (A); Ubi indicates**the ubiquitin promoter; S is the sense strand, intron is the sequence of intron, AS is the antisense strand, the S, intron and AS constitue the **linear expression cassette**of*Pina*RNAi**sequence for RNA interference;**nosindicatesthe nosterminator; bar indicates the bialaphos resistance gene sequence.

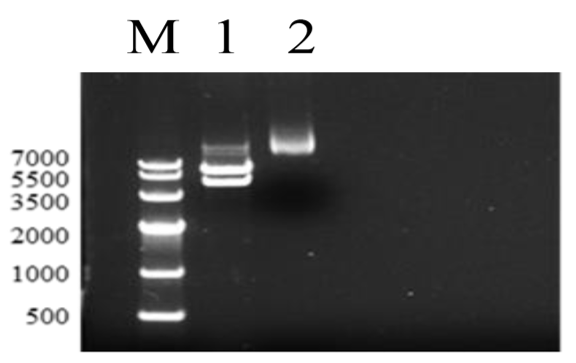


Figure 2 Amplification of linear expression constructs;

Lane M: DNA Marker; Lane 2: original RNAi vector; lane 1: linear expression cassette.

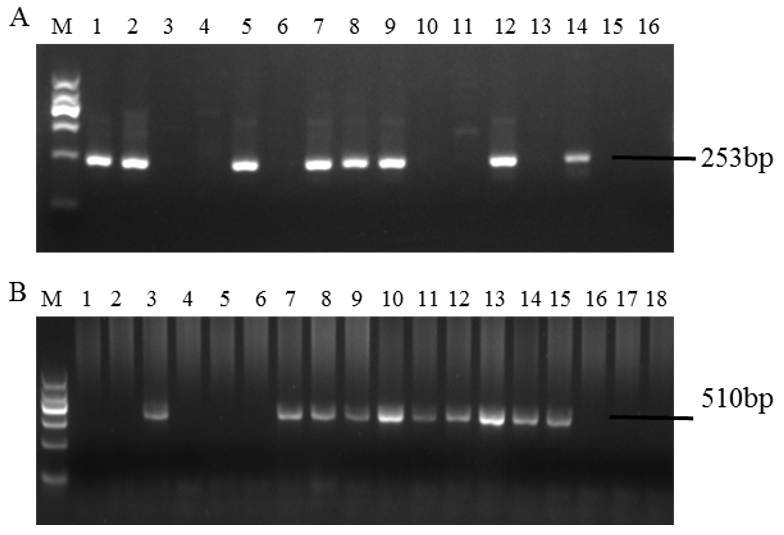


Figure3 The PCR identification of T0 regenerant plants**;**

A:Detection of semi-nested PCR (D, Nos), lane M: DNA Marker Ⅱ, lane 1-15: transgenic regenerated plants, lane 16: distilled water for negative control;

B: Detection of nested PCR (Inno,D5), lane M: DNA Marker Ⅱ, lanes 1-13: transgenic regenerated plants, lane 18: distilled water for negative control.

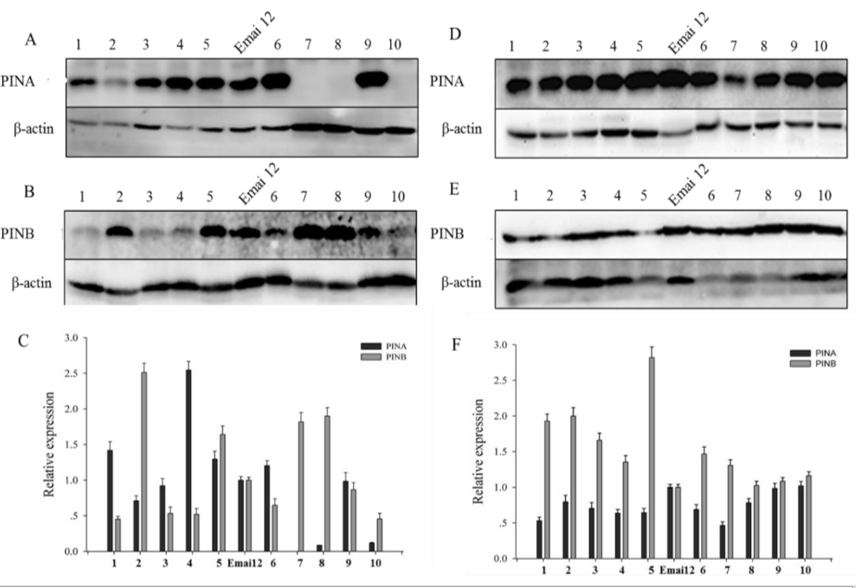


Figure 4 Western blotting analysis of T2 generation transgenic wheat PIN protein expression

A, B and C: Western blotting analysis of PINA (A) and PINB (B) protein and relative quantitative analysis (C) of the two proteins by densitometry in seeds of transgenic line E3 and wild-type, lanes 1-10: Transgenic regenerated plants E3, laneEmai 12: **non-transgenic Emai12;**

D, E and F: Western blotting analysis of PINA (D) and PINB (E) protein and relative quantitative analysis (F) of the two proteins by densitometry in seeds of transgenic line L2 and wild-type,lanes 1-10: transgenic regenerated plants L2, laneEmai 12: **non-transgenic Emai12;**

Table 1 Primer sequences

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Primer Name | Primer sequence（5'-3'） | amplicon | annealing temperature | amplification area |
| Inno-F | GGTTCTGAGATAGGGTAAACTCCACT | 704 bp | 61°C | Intron +antisense+ Nos |
| Inno-R | TCATCGCAAGACCGGCAA |
| D5-F | ATGTGCCCGACAAGGCTAGG | 510bp | 61°C | Intron + antisense + Nos |
| D5-R | TTTATTGCCAAGTGTTTGAACGA |
| D-F | GATCTAGTAACATAGATGAC | 428 bp | 55°C | Intron + antisense +Nos |
| D-R | TTAAATGTATAATTGCGGGACT |
| Nos-F | GATCGTTCAAACATTTGGCA | 253bp | 57°C | Nos |
| Nos-R | GATCTAGTAACATAGATGAC |

Table 2 The kernel hardness index of T2 transgenic wheat

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| T2 | Hardness | | Weight | | Moisture | | Diameter | |
| Average | Std.Dev. | Average | Std.Dev. | Average | Std.Dev. | Average | Std.Dev. |
| Emai12 | 38.2 | 12.2 | 32.4 | 8.8 | 10.5 | 0.2 | 2.9 | 0.4 |
| VC | 39.7 | 11.5 | 38.1 | 8.7 | 11.1 | 0.3 | 2.8 | 0.3 |
| Luna | 89.2 | 15.2 | 42.1 | 8.7 | 11.2 | 0.3 | 2.5 | 0.3 |
| E1 | 40.5 | 11.8 | 35.9 | 7.2 | 11.1 | 0.2 | 2.8 | 0.3 |
| E2 | 59.4 | 22.1 | 34.9 | 5.9 | 11.3 | 0.2 | 2.7 | 0.2 |
| E3 | 48.6 | 20.5 | 38.5 | 5.9 | 11.2 | 0.2 | 3.0 | 0.2 |
| E4 | 37.5 | 10.7 | 34.7 | 6.8 | 11.5 | 0.3 | 2.8 | 0.3 |
| L1 | 55.1 | 19.5 | 35.2 | 5.5 | 11.3 | 0.2 | 2.8 | 0.2 |
| L2 | 39.2 | 13.3 | 37.2 | 6.2 | 11.1 | 0.3 | 2.9 | 0.2 |
| L3 | 45.9 | 20.7 | 36.4 | 7.1 | 11.6 | 0.3 | 2.7 | 0.3 |