**Haemato-Biochemical and Immunological Profiling of Turkeys (*Meleagris gallopavo*) Under Intensive Rearing System in India**

**Abstract**

The present study aimed to assess haematological, biochemical, and immunological parameters in turkeys (*Meleagris gallopavo*) reared under an intensive housing system suited to Indian conditions. Seventy-two day-old healthy turkey poults were wing-banded, weighed, and randomly allocated into three replicates of 24 birds each, and reared following standard management practices according to ICAR (2013) feeding guidelines. Observations were recorded at 6 and 12 weeks of age to evaluate physiological and immunological development. Among haematological parameters, total erythrocyte count (TEC: 3.34 × 10⁶/µl) and total leukocyte count (TLC: 13.6 × 10³/µl) were significantly higher (p<0.01) at 6 weeks, whereas haemoglobin (Hb: 10.95 g/dl), packed cell volume (PCV: 36.8%), mean corpuscular volume (MCV: 148.2 fL), and mean corpuscular haemoglobin concentration (MCHC: 28.58 g/dl) were significantly higher (p<0.01) at 12 weeks. In terms of biochemical parameters, significant age-related and replicate-wise variations were observed in blood glucose (247.79 mg/dl), AST (301.4 U/L), and ALT (5.16 U/L), while no significant differences were found in total protein, creatinine, BUN, cholesterol, or triglycerides. Immunological evaluation revealed a significant enhancement (p<0.01) in both humoral immunity (HA titre: 4.6) and cell-mediated immunity (PHA-P response: 0.93 mm) at 12 weeks compared to 6 weeks. The results demonstrate that age has a significant effect on various blood and immune parameters, and that intensive rearing systems, when properly managed, can effectively support normal physiological and immunological development in turkeys. These outcomes provide valuable reference data for further research and optimization of turkey production practices.

**Keywords:** *Biochemical, Haematological, Immunological, Intensive rearing, Turkey*

**Introduction**

The poultry sector significantly contributes to India's GDP and provides employment across various socio-economic strata. Indian poultry systems include traditional backyard setups and intensive commercial farming, with the latter evolving over the last four decades into a highly organized, vertically integrated industry. Globally, poultry production comprises chickens (63%), ducks (11%), geese (9%), turkeys (5%), pigeons (3%), and guinea fowls (3%) (Besbes, 2009). Turkey (Meleagris gallopavo), though lesser-known in India, is economically and nutritionally valuable, ranking behind chicken, duck, guinea fowl, and quail in global importance (Anandh and Jagatheesan, 2015). Globally, turkeys account for about 2% of the poultry population (Hayet et al., 2021). In India, Broad Breasted Bronze, Broad Breasted White, and Beltsville Small White are the most common breeds reared (Anandh and Jagatheesan, 2015). Turkey meat is increasingly favoured for its high protein and low-fat content, offering 24% protein, 6.6% fat, and 162 kcal per 100 grams (Asaduzzaman et al., 2017). It is also preferred for its superior foraging and fibre digestion capabilities (Thomas et al., 2014). The aesthetic appeal and nutritional value of turkey have bolstered its appeal (Ogundipe and Dafwang, 1980). Blood profile analysis is essential for monitoring bird health. Hematological parameters vary due to multiple factors such as age, sex, breed, and environmental conditions (Olorode and Longe, 2000). Standard values in turkeys include Hb: 9.1–13.2 g/dL, PCV: 32.4–40.6%, erythrocyte count: 2.5–3.3×10⁶/µL, leukocyte count: 15.6–23.1×10³/µL, heterophils: 41–55%, lymphocytes: 37–50%, and monocytes: 3–6% (Arora, 2010). Deviations in these values can signal stress, infection, dehydration, or immunosuppression. Biochemical markers help assess organ functionality. Normal values include total protein: 3.5–5.6 g/dL, albumin: 1.5–2.8 g/dL, glucose: 190–240 mg/dL, cholesterol: 120–180 mg/dL, uric acid: 4.5–8.2 mg/dL, AST: 180–240 U/L, and ALT: 12–20 U/L (Pavlak et al., 2005). Elevated AST and ALT indicate liver dysfunction, while high uric acid suggests renal impairment. Routine monitoring aids early diagnosis and informs therapeutic and nutritional strategies. The turkey’s immune system includes innate and adaptive branches. Adaptive immunity involves cell-mediated and humoral responses, with the latter focusing on antibody production for combating infections. Monitoring antibody titers post-vaccination is a standard practice to evaluate immune response (Yang et al., 2017). Despite its advantages, turkey farming in India is underdeveloped, lacking robust scientific management. Enhancing disease resistance and understanding correlations between humoral immunity and reproductive performance are critical for sustainable development. Scientific rearing practices and targeted genetic improvements are essential for advancing commercial turkey farming under Indian conditions (Thomas et al., 2014).

**MATERIALS AND METHODS:**

The study was carried out at Imaliya Farm, Department of Poultry Science, NDVSU, Jabalpur, using 72 day-old turkey poults procured from the university hatchery and reared under intensive housing up to 12 weeks of age. Uniform brooding, feeding, and watering practices were followed across all replicates. Poults received Lixen powder and Vitamin B-complex during the first three days, and a coccidiostat (Diclazuril @ 1 kg/ton feed) was added during brooding. Vaccination included ND (B1 strain) at day-old, Fowl pox at 4–5 weeks, ND (R2B) at 6 weeks, and Cholera between 8–10 weeks. Skimmed milk powder (40 g/15 L) was added to drinking water 15 minutes before each vaccination to improve efficacy.

**Experimental Design**

All 72 healthy poults were weighed, wing-banded, and randomly assigned into three replicates, each containing 24 birds. The experiment was conducted in a floor housing system with individual pens arranged on both sides of a central passage. Before the start of the experiment, the entire poultry house was thoroughly cleaned, whitewashed, disinfected, and dried, and left vacant for three days to ensure biosecurity. Sawdust was used as litter material and was uniformly spread on the floor to a depth of approximately 2 cm. Each replicate was provided with identical brooding, feeding, and watering arrangements to maintain uniformity. The turkey poults were fed a standard diet formulated as per ICAR (2013) feeding standards. Two phases of feeding were followed: Poult phase (0–6 weeks): Turkey chick ration, Grower phase (6–12 weeks): Turkey grower ration. The composition of the experimental diet is detailed in (Table 1)

**Table 1:** Composition of turkey chick and turkey grower diet

|  |  |  |  |
| --- | --- | --- | --- |
| **S. No.** | **Ingredients**  **(Part/100kg)** | **0-6 weeks diet**  **(CP 24%, 2800 kcal ME/kg)** | **6-12 weeks diet**  **(CP 22%, 2800 kcal ME/kg)** |
| 1 | Maize | 43.5 | 47 |
| 2 | Soyabean meal | 37.5 | 32.6 |
| 3 | Deoiled rice polish | 16.1 | 17.5 |
| 4 | Mineral mixture | 1.50 | 1.50 |
| 5 | Vitamins mixture | 0.25 | 0.25 |
| 6 | Lime stone powder | 0.35 | 0.35 |
| 7 | Dicalcium phosphate | 0.40 | 0.40 |
| 8 | Salt | 0.30 | 0.30 |
| 9 | Coccidiostat (diclazuril) | 0.10 | 0.10 |
| Total | - | 100 | 100 |

**MEASUREMENTS AND OBSERVATIONS**

The following observations were recorded during the experimental period

1. **Hematological Parameter of Turkey:**

For hematological analysis in turkeys, approximately 3 ml of pooled blood was collected from six birds per replicate (not exceeding 1% of each bird’s body weight) via wing vein puncture into sterile heparinised vacutainer tubes on days 6 and 14 at 7:00 A.M., ensuring minimal stress to the birds. The samples were immediately transported in ice to the lab for processing. Part of the blood was used for hematological parameters including RBC, WBC, DLC, Hb, PCV, MCV, MCH, and MCHC, while the rest was used for plasma separation and biochemical tests. For Total Erythrocyte Count (TEC), blood was diluted with a specific RBC fluid and counted using a hemocytometer under high power, while Total Leukocyte Count (TLC) used a different diluting fluid and was counted under low power in the corner squares. Packed Cell Volume (PCV) was determined by centrifuging blood in Wintrobe tubes at 3000 rpm for 30 minutes, measuring the erythrocyte mass as a percentage. Hemoglobin (Hb) estimation involved mixing blood with 0.1 N HCl in a Sahli’s hemoglobinometer, forming acid hematin, and diluting until color matched standard tubes to determine concentration in g/dl.

1. **Erythrocyte indices**

The following erythrocyte indices were calculated: -

1. **Mean corpuscular volume (MCV) in femtolitre (fL)**

Or

1. **Mean corpuscular haemoglobin concentration (MCHC) in g/dl**

Or

1. **Biochemical parameters of Turkey:**

Serum was isolated from the blood sample collected at 6 and 12 weeks of age from 6 birds each replicate in 2000 rpm for 15 minutes. Following biochemical parameters were recorded.

1. **Blood glucose:**

Blood glucose was measured by using Glucose Oxidase (Trinder, 1969) method and the result was expressed as milligram per deciliter (mg/dl). In Trinder’s method, glucose in the sample is oxidized to yield gluconic acid and hydrogen peroxidase in the presence of glucose oxidase. Take 1 ml glucose working reagent and add 0.01 ml of sample in a test tube. Mix well and incubate for 15 minutes at 370C.Read absorbance of the standard and each test at 505 nm against reagent blank. Glucose was calculated by the following formula:

Concentration of Standard (mg/dl)

1. **Total Protein:**

In this study, the Erba Mannheim LiquiXX Total Protein Biuret Method, an end-point diagnostic reagent kit, was employed for the in vitro quantitative estimation of total protein in broiler serum. The procedure involved pipetting 1000 µl of the working reagent into three separate tubes labeled Blank, Standard, and Test. To the Blank tube, 20 µl of distilled water was added; to the Standard tube, 20 µl of protein standard; and to the Test tube, 20 µl of serum sample. The contents of each tube were mixed thoroughly and incubated at 37°C for 10 minutes. Following incubation, the absorbance of the Standard and Test solutions was measured at 546 nm using the reagent blank as reference. This colorimetric method allowed for precise quantification of total protein based on the biuret reaction.

Concentration of standard (g/dL)

1. **Alanine Transaminase (ALT):**

SGPT/ALT was estimated by 2, 4-DNPH (Reitman and Frankel, 1957), using commercial kit (Span Diagnostics Ltd., India) (Table 2)

**Table 2:** Alanine Transaminase (ALT)

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **S.No.** | **Pipette into tube marked** | **Blank** | **Standard** | **Test** | **Control** |
| Volume in ml | | | | | |
| **1.** | Reagent 1 | 0.25 | 0.25 | 0.25 | 0.25 |
| **2.** | Serum/Plasma | **-** | **-** | 0.05 | **-** |
| **3.** | Standard (150 U/L) | **-** | 0.05 | **-** | **-** |
| Mixed well and incubated at 37˚C for 60 minutes | | | | | |
| **5.** | Reagent 2 | 0.25 | 0.25 | 0.25 | 0.25 |
| **6.** | Deionised water | 0.05 | **-** | **-** | **-** |
| **7.** | Serum/Plasma | **-** | **-** | **-** | 0.05 |
| Mixed well and standard at room temperature (15 - 30˚C) for 20 minutes | | | | | |
| **9.** | Solution 1(1ml Reagent 3 +9ml DW | 0.25 | 0.25 | 0.25 | 0.25 |
| **10.** | Mixed well and read the O.D. against purified water in a photometer at 505 nm, within 15 minutes | | | | |



1. **Aspartate Transaminase (AST):**

SGOT/AST was estimated by 2, 4-DNPH (Reitman and Frankel, 1957), using commercial kit (Span Diagnostics Ltd., India) (Table 3)

**Table 3:** Aspartate Transaminase (AST)

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **S.No.** | **Pipette into tube marked** | **Blank** | **Standard** | **Test** | **Control** |
| Volume in ml | | | | | |
| 1. | Reagent 1 | 0.25 | 0.25 | 0.25 | 0.25 |
| 2. | Serum/Plasma | - | - | 0.05 | - |
| 3. | Standard (114 IU/L) | - | 0.05 | - | - |
| Mixed well and incubated at 37˚C for 60 minutes | | | | | |
| 5. | Reagent 2 | 0.25 | 0.25 | 0.25 | 0.25 |
| 6. | Deionized water | 0.05 | - | - | - |
| 7. | Serum/Plasma | - | - | - | 0.05 |
| Mixed well and standard at room temperature (15 - 30˚C) for 20 minutes | | | | | |
| 9. | Solution 1(1ml Reagent 3 + 9ml DW) | 0.25 | 0.25 | 0.25 | 0.25 |
| Mix and read the O.D. against purified water in a photometer at 505 nm, within 15 minutes | | | | | |



1. **Creatinine:**

Creatinine was measured colorimetrically by using Jaffe’s method (Bower, 1980) and the results were expressed as milligram per deciliter (mg/dl). Creatinine reacts with alkaline picrate to produce a Jaffe’s reaction. Take 1 ml reagent and add 0.1 ml of sample in a test tube. Mix well and read initial absorbance (A1) 20 seconds after mixing and final absorbance (A2) 80 seconds after mixing. Creatinine was calculated by the following formula:

Creatinine (mg/dl) = (ΔA of Test / ΔA of Standard) × Concentration of Standard (mg/dl)

Where:

ΔA = A₂ - A₁

(A₂ = absorbance after reaction, A₁ = blank or initial absorbance)

1. **Blood Urea Nitrogen (BUN):**

Blood Urea Nitrogen was measured by using Diacetyl Monoxime Method (Karr,1924) and the results were expressed as miligram per deciliter (mg/dl). Take 1 ml reagent and add 0.02 ml of sample in a test tube. Mix well and aspirate standard followed by samples. BUN was calculated by the following formula:

The correctly formatted formula for calculating Blood Urea Nitrogen (BUN) is:

BUN (mg/dl) = (ΔA of Test / ΔA of Standard) × Concentration of Standard (mg/dl)

Where:

ΔA = A₂ - A₁

(A₂ = absorbance of the sample or standard, A₁ = absorbance of the blank)

1. **Total Cholesterol:**

Erba Mannheim cholesterol *DES* dynamic extended stability CHOD-pad method (with LCF) end point diagnostic reagent kit was used for *in vitro* quantitative determination of cholesterol in broiler serum in this study (Table 4)

**Table 4:** Total Cholesterol

|  |  |  |  |
| --- | --- | --- | --- |
| **Pipette into tubes marked** | **Blank** | **Standard** | **Test** |
| Working Reagent | 1000 µl | 1000 µl | 1000 µl |
| Distilled water | 20 µl | - | - |
| Standard | - | 20 µl | - |
| Test | - | - | 20 µl |

1. Mix well, incubate at 370C for 10 minutes.

2. Aspirate blank followed by standard and tests.

3. Read the absorbance of standard and each test tube against blank at 505 nm on bichromatic analyzer.

1. **Triglyceride:**

Erba Mannheim Triglycerides *DES* dynamic extended stability with lipid clearing agent GPO–trinder method, end point diagnostic reagent kit was used for *in vitro* quantitative determination of triglycerides in broiler serum in this study (Table 5)

**Table 5:** Triglyceride

|  |  |  |  |
| --- | --- | --- | --- |
| **Pipette into tubes marked** | **Blank** | **Standard** | **Test** |
| Working Reagent | 1000 µl | 1000 µl | 1000 µl |
| Distilled water | 10 µl | - | - |
| Standard | - | 10 µl | - |
| Test | - | - | 10 µl |

1. Mix well, incubate at 370C for 10 minutes.

2. Read the absorbance of standard and each test tube at 505 nm on dichromatic analyzer against reagent blank.

concentration of standard (mg/dL)

1. **Immune response**
2. **Humoral immune response**
3. **Preparation of sheep red blood cell (SRBC) suspension:**

Blood from jugular vein of healthy sheep was collected in Alsever’s solution. The blood was centrifuged at 2500 rpm for about 10 minutes. The supernatant was discarded and red blood cells were washed thrice in PBS. Suspension of SRBC (1% v/v) in PBS was prepared and stored in refrigerator at 4˚C until use.

1. **Immunization and harvesting of immune serum:**

1.0 ml suspension of SRBC was injected intravenously to 6 birds in each replicate to study the primary antibody response to SRBC. After 5 days 2 ml blood was collected from the wing vein. The blood was allowed to clot, the serum was collected and frozen (-20˚C) until analyzed for the antibody titres to SRBC.

**Haemagglutination test (HA test):**

The antibody titre to SRBC was determined by HA methods (Siegel and Gross, 1980).

**Procedure:**

1. 50 µl of PBS was pipetted out in each well of the micro titter plate.
2. 50 µl of serum was added in the first well.
3. Two fold serial dilution were made up to row 11 and 12 was kept as control.
4. 50 µl of 1% SRBC was added in each well and mixed by gentle tapping
5. The plates were covered and then kept at 37˚C for 1 hour for incubation.
6. The plates were read under bright light.
7. The reciprocal of highest dilution showing clear agglutination was the ends titre. The titre were expressed as log 2.
8. **Cell-mediated immune response:**

The cellular immune response was assessed by cutaneous basophilic hypersensitivity test by using PHA-P (Phytohaemagglutinin, lectin from *Phaseolus vulgaris*). 6 birds from each replicate were selected and the toe thickness of both left and right foot at 3rd and 4th interdigital spaces were measured by micrometer. Immediately after measurements 100 mg of PHA-P suspended in 0.1 ml of phosphate buffer saline (PBS) and 0.1 ml of PBS was injected into right and left foot (acted as control), respectively. The web swelling of both the foots were measured 24 hours after injection. The cell-mediated immune response (CMIR) or Foot Web Index was determined by using the following formula

FWI = (R2-R1) - (L2-L1) (Corrier and DeLoach, 1990).

where R2- thickness after 24 hours of PHA-P injection

R1- thickness before injection of PHA-P

L2- thickness after 24 hours of PBS solution

L1- thickness before injection of PBS solution

**STATISTICAL ANALYSIS:**

Statistical analysis of the data was done by analysis of variance using completely randomized design (CRD) as per Snedecor and Cochran (1994). Differences among the treatments were tested for significance by Duncan’s Multiple Range Test (1955).

**RESULTS AND DISCUSSION:**

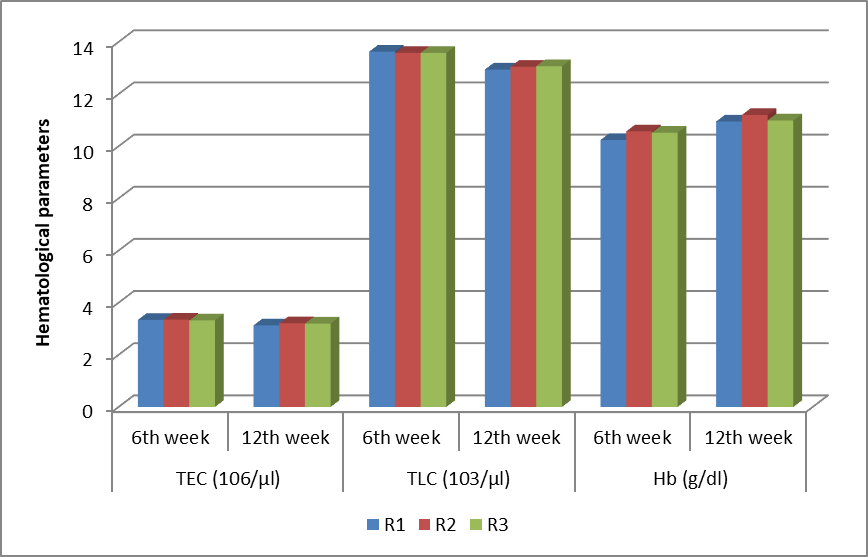
**1. Haematological Studies**

The haematological profile observed in this study presents values for Total Erythrocyte Count (TEC), Total Leukocyte Count (TLC), Haemoglobin (Hb), Packed Cell Volume (PCV), Mean Corpuscular Volume (MCV), and Mean Corpuscular Hemoglobin Concentration (MCHC) across three replicates (R1, R2, R3) at 6 and 12 weeks of age, as shown in (Table 6 and Figures 1 and 2). TEC values showed a gradual decline from 6th to 12th week across all replicates, with R2 recording a significantly higher TEC at the 12th week (p<0.01). TLC values remained consistent across replicates, with no significant differences throughout the experimental period. For haemoglobin concentration, R2 demonstrated significantly higher levels at the 6th week (p<0.01), while all replicates converged to comparable values by the 12th week. PCV values at the 6th week were significantly higher in R2 (p<0.01), though no significant variation was found at the 12th week. MCV values remained statistically similar across replicates, suggesting stable red cell size throughout. MCHC values showed significant differences at both 6th and 12th weeks, with R3 registering the highest MCHC at 12 weeks (p<0.01). These findings corroborate earlier studies by Lazar et al. (2012) in hybrid B.U.T 6 turkeys and other research by Olaniyi et al. (2012), Sogunle et al. (2008), and Safiyu et al. (2020), who noted minimal impact of rearing systems on turkey haematology. The TEC, TLC, Hb, and PCV values fall within normal physiological ranges previously established for indigenous and crossbred turkeys (Isidahomen et al., 2013). Contrarily, studies on chickens by Addass et al. (2012) and Etim et al. (2014) indicated higher haematological indices in indoor-reared birds, highlighting species-specific responses. Moreover, Milenkaya et al. (2013) reported sex and reproductive stage-related variations in haematological indices, which could explain discrepancies due to physiological or environmental differences from the current study.

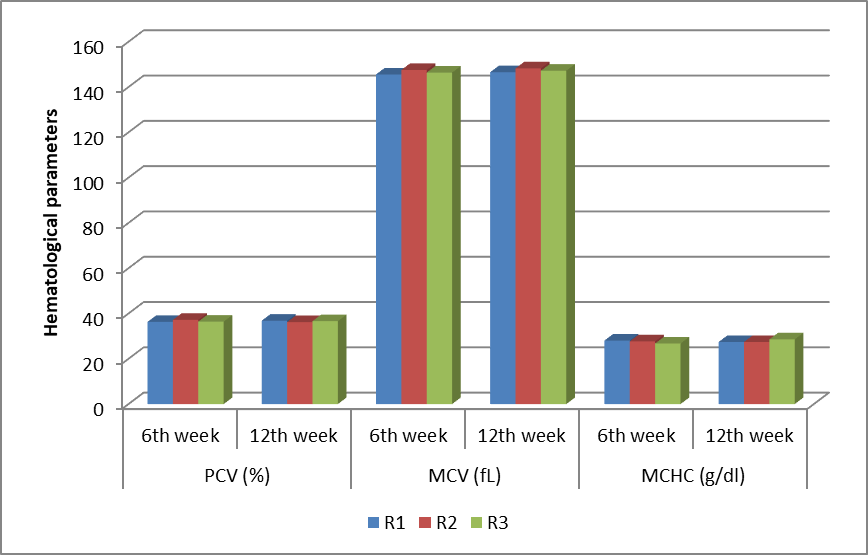
**Table 6: Hematological parameters of Turkey in different replicates at 6 weeks interval**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Parameters** | **Age** | **R1** | **R2** | **R3** |
| **TEC**  **(106/µl)** | **6th week** | 3.34±0.03 | 3.35±0.03 | 3.32±0.00 |
| **12th week** | 3.13b±0.02 | 3.21a±0.02 | 3.20a±0.00 |
| **TLC**  **(103/µl)** | **6th week** | 13.63±0.16 | 13.58±0.16 | 13.58±0.00 |
| **12th week** | 12.94±0.13 | 13.05±0.13 | 13.07±0.00 |
| **Hb**  **(g/dl)** | **6th week** | 10.24b±0.15 | 10.57a±0.15 | 10.52ab±0.00 |
| **12th week** | 10.95±0.13 | 11.20±0.13 | 10.99±0.00 |
| **PCV**  **(%)** | **6th week** | 36.29b±0.39 | 37.20a±0.39 | 36.41ab±0.00 |
| **12th week** | 36.83±0.40 | 36.20±0.40 | 36.62±0.00 |
| **MCV**  **(fL)** | **6th week** | 145.54±2.25 | 147.54±2.25 | 146.37±0.00 |
| **12th week** | 146.54±2.27 | 148.25±2.27 | 147.20±0.00 |
| **MCHC**  **(g/dl)** | **6th week** | 28.04a±0.51 | 27.70ab±0.51 | 26.75b±0.00 |
| **12th week** | 27.41b±0.47 | 27.41b±0.47 | 28.58a±0.00 |

Means with atleast one common superscripts in a column are nonsignificant (p<0.01)



**Figure 1: Graph showing the Hematological parameters (TEC, TLC & Hb) of Turkey in different replicates at 6 weeks interval**



**Figure 2: Graph showing the Hematological parameters (PCV, MCV & MCHC) of Turkey in different replicates at 6 weeks interval**

**2. Biochemical Studies**

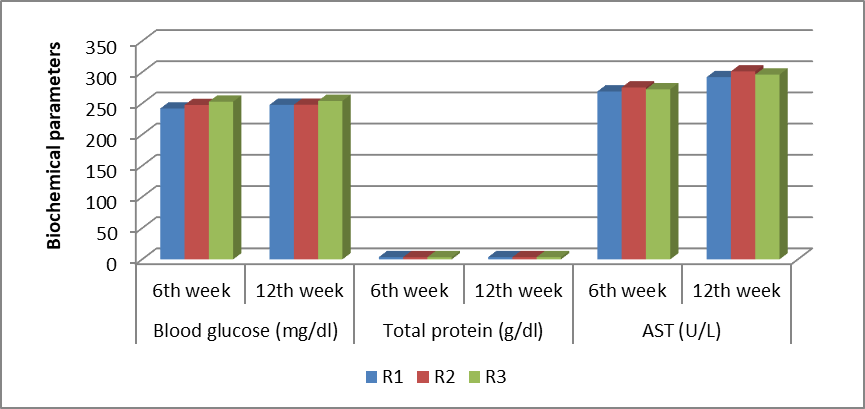
The biochemical parameters of turkeys were assessed at 6-week intervals to evaluate their physiological and metabolic responses under intensive rearing, with data represented in (Table 7 and Figures 3–5). Blood glucose levels showed a statistically significant increase in R3 at the 6th week (p<0.01), while no significant differences were noted at the 12th week. Total protein values remained consistent across all replicates throughout the study period, indicating stable protein metabolism. Aspartate transaminase (AST) and alanine transaminase (ALT) activities increased from the 6th to the 12th week in all replicates, though no significant differences were observed, suggesting uniform liver function. Creatinine levels remained identical (0.15 ± 0.00 mg/dL) across all replicates and time points, reflecting stable renal function. Similarly, blood urea nitrogen (BUN) values showed minimal variation between 1.33 and 1.35 mg/dL, with no significant differences among groups. Total cholesterol levels ranged from 128.91 to 137.87 mg/dL, while triglyceride concentrations varied between 171.25 and 175.16 mg/dL, neither showing significant replicate-based variation. These observations align with Bounous et al. (2000), who documented glucose values of 215–500 mg/dL and cholesterol levels of 60–220 mg/dL in wild juvenile turkeys, with noted seasonal fluctuations. The cholesterol values recorded in the present study suggest physiological normalcy and absence of metabolic stress. Comparable ALT, AST, total protein, and globulin values were also reported by Schmidt et al. (2010), while Ibrahim et al. (2012) provided similar serum values in turkeys raised under semi-arid Nigerian conditions. Slightly higher biochemical indices observed by Agina et al. (2015) in domestic turkeys such as total cholesterol (185.7 mg/dL), creatinine (1.05 mg/dL), AST (83.68 IU/L), and ALT (13.08 IU/L) could be attributed to breed differences, dietary variations, or environmental influences, underscoring the relevance of localized physiological benchmarks in turkey farming.

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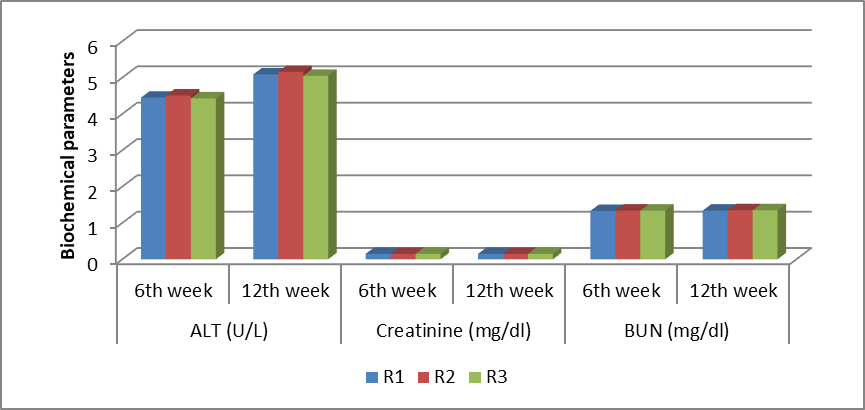
**Table7: Biochemical parameters of Turkey in different replicates at 6 weeks interval**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Parameters** | **Age** | **R1** | **R2** | **R3** |
| **Blood glucose (mg/dl)** | **6th week** | 241.91b±4.37 | 247.50ab±4.37 | 252.83a±0.00 |
| **12th week** | 247.79±4.11 | 247.91±4.11 | 254.04±0.00 |
| **Total protein (g/dl)** | **6th week** | 3.67±0.09 | 3.55±0.09 | 3.55±0.00 |
| **12th week** | 3.63±0.08 | 3.59±0.08 | 3.63±0.00 |
| **AST (U/L)** | **6th week** | 269.25±3.68 | 275.70±3.68 | 272.70±0.00 |
| **12th week** | 292.20±4.80 | 301.41±4.80 | 296.30±0.00 |
| **ALT (U/L)** | **6th week** | 4.45±0.09 | 4.51±0.09 | 4.43±0.00 |
| **12th week** | 5.09±0.09 | 5.16±0.09 | 5.05±0.00 |
| **Creatinine (mg/dl)** | **6th week** | 0.15±0.00 | 0.15±0.00 | 0.15±0.00 |
| **12th week** | 0.15±0.00 | 0.15±0.00 | 0.15±0.00 |
| **BUN (mg/dl)** | **6th week** | 1.33±0.00 | 1.34±0.00 | 1.34±0.00 |
| **12th week** | 1.34±0.00 | 1.35±0.00 | 1.35±0.00 |
| **Total cholesterol (mg/dl)** | **6th week** | 128.91±3.59 | 134.25±3.59 | 131.41±0.00 |
| **12th week** | 136.37±3.80 | 137.87±3.80 | 132.00±0.00 |
| **Triglyceride (mg/dl)** | **6th week** | 175.16±2.28 | 172.30±2.28 | 172.45±0.00 |
| **12th week** | 171.25±2.18 | 174.12±2.18 | 174.50±0.00 |

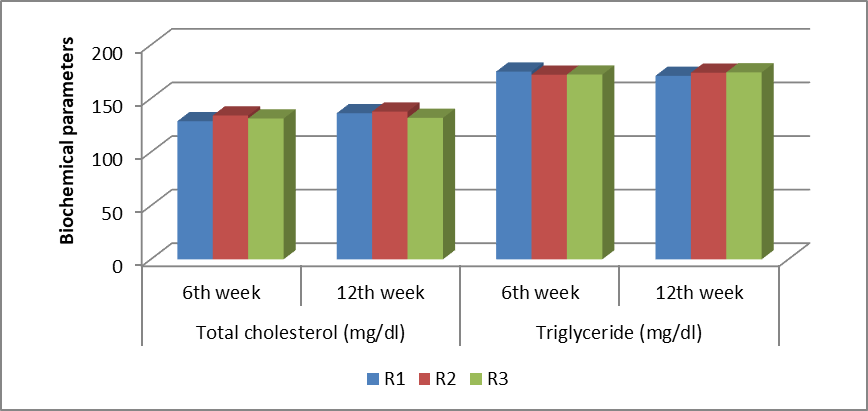
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**Figure 3: Graph showing the Biochemical parameters (Blood Glucose, Total Protein & AST) of Turkey in different replicates at 6 weeks interval**



**Figure 4: Graph showing the Biochemical parameters (ALT, Creatinine & BUN) of Turkey in different replicates at 6 weeks interval**



**Figure 5: Graph showing the Biochemical parameters (Total cholesterol & Triglyceride) of Turkey in different replicates at 6 weeks interval**

**3. Immunological Studies**

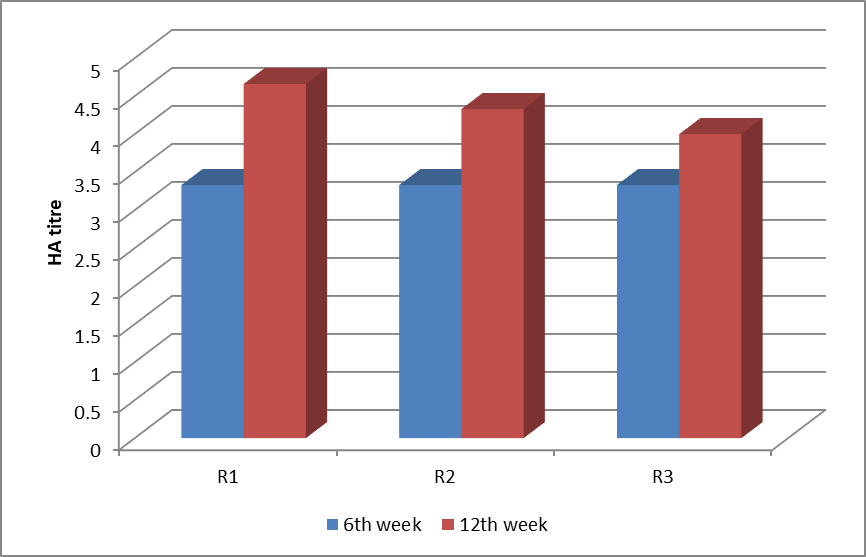
The immunological performance of turkeys reared under an intensive system was evaluated at 6 and 12 weeks using two key indicators humoral and cell-mediated immunity with data presented in (Tables 8 and 9 and visualized in Figures 6 and 7). Humoral immunity, assessed through haemagglutination (HA) titre against sheep red blood cells (SRBCs), showed values of 3.33 ± 0.59 (6th week) and 4.66 ± 0.74 (12th week) in R1, 3.33 ± 0.59 and 4.33 ± 0.74 in R2, and 3.33 ± 0.00 and 4.00 ± 0.00 in R3, with no statistically significant differences across replicates, indicating consistent antibody responses. Cell-mediated immunity, measured via toe web swelling in response to intradermal phytohaemagglutinin-P (PHA-P) injection, showed significant variation; R1 exhibited the highest swelling response at 12 weeks (0.93 ± 0.01 mm, p<0.01), suggesting stronger cellular immunity. These findings are consistent with Li et al. (2000), who reported peak antibody titres on day 10 in males and day 7 in females post-primary SRBC challenge, and day 7 in both sexes after a secondary challenge. Loa et al. (2001) documented a significant HA titre increase (~3.4) by day 3 post-SRBC injection and toe web swelling of 0.35–0.48 mm in turkeys, with no significant group-wise differences. Cheema et al. (2007) also observed comparable humoral and cell-mediated responses in both commercial and random-bred turkey strains, emphasizing that while genetic background influences immune response, environmental conditions and nutrition are crucial determinants. Bhattacharyya et al. (2018) further reported that maternal dietary interventions did not affect humoral responses to SRBC in turkey poults, supporting the present study’s finding of uniform HA titres across replicates, thus reinforcing the idea that rearing system and experimental diet had minimal effect on the humoral immune profile.

A total of 72 healthy day-old turkey poults were wing-banded, weighed, and randomly assigned into three replicates with 24 poults each, receiving uniform brooding, feeding, and watering conditions. Standard poult and grower rations were provided as per ICAR (2013) feeding norms. Blood sampling and parameter evaluation were carried out at the 6th and 12th weeks of age. Among the haematological indices, TEC (3.34 × 10⁶/µl) and TLC (13.6 × 10³/µl) were significantly higher (p<0.01) at the 6th week, whereas Hb (10.95 g/dl), PCV (36.8%), MCV (148.2 fL), and MCHC (28.58 g/dl) showed significantly elevated values (p<0.01) at the 12th week. Biochemical analysis revealed significant differences in blood glucose (247.79 mg/dl), AST (301.4 U/L), and ALT (5.16 U/L) between age intervals and across replicates (p<0.01), while parameters such as total protein (3.63 g/dl), creatinine (0.15 mg/dl), BUN (1.35 mg/dl), cholesterol (137.8 mg/dl), and triglycerides (175.1 mg/dl) remained statistically unaffected. Immunological evaluation showed that humoral immunity, measured via HA titre against SRBCs, was significantly higher (p<0.01) at the 12th week (4.0–4.6) than the 6th week. Similarly, cell-mediated immunity assessed through toe web swelling following PHA-P injection peaked at the 12th week (mean: 0.93 mm, p<0.01), indicating a progressive enhancement in immune response with age under intensive rearing conditions.

**Table 8: HA titre of Turkey in different replicates at 6 weeks interval**

|  |  |  |  |
| --- | --- | --- | --- |
| **Interval** | **R1** | **R2** | **R3** |
| **6th week** | 3.33±0.59 | 3.33±0.59 | 3.33±0.00 |
| **12th week** | 4.66±0.74 | 4.33±0.74 | 4.00±0.00 |

Means with atleast one common superscripts in a column are nonsignificant (p<0.01)

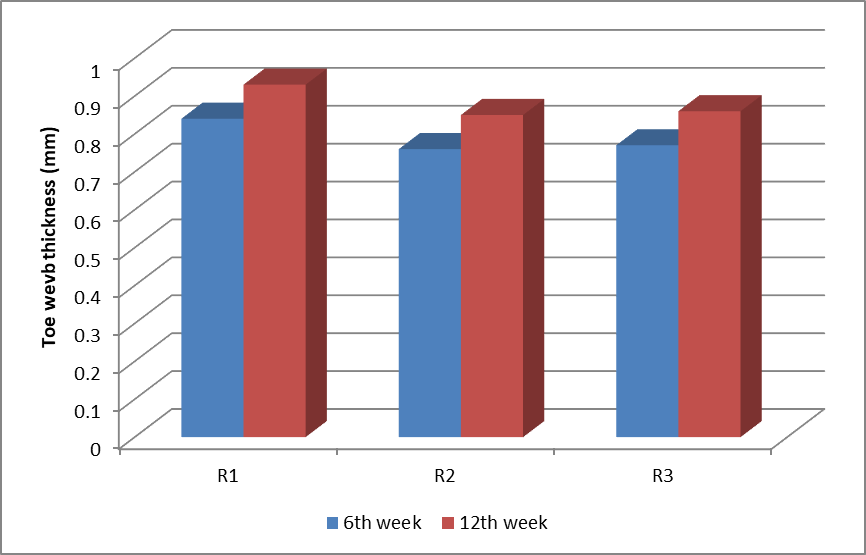


**Figure 6: Graph showing the HA titre of Turkey in different replicates at 6 weeks interval**

**Table 9: Cell-mediated immunity (by using PHA-P) (in mm) of Turkey in different replicates at 6 weeks interval**

|  |  |  |  |
| --- | --- | --- | --- |
| **Interval** | **R1** | **R2** | **R3** |
| **6th week** | 0.84a±0.01 | 0.76b±0.01 | 0.77b±0.00 |
| **12th week** | 0.93a±0.01 | 0.85b±0.01 | 0.86b±0.00 |

Means with atleast one common superscripts in a column are nonsignificant (p<0.01)



**Figure 7: Graph Showing the Cell-mediated immunity (by using PHA-P) (in mm) of Turkey in different replicates at 6 weeks interval**

**CONCLUSION**

The present study highlights the dynamic changes in haematological, biochemical, and immunological parameters of turkeys reared under an intensive system, demonstrating that age plays a significant role in physiological development. Haematological indices such as TEC and TLC were significantly higher at the 6th week, while Hb, PCV, MCV, and MCHC increased at the 12th week, reflecting progressive haematological maturation. Among the biochemical parameters, blood glucose, AST, and ALT exhibited significant variations with age and between replicates, suggesting ongoing metabolic adaptation. In contrast, total protein, creatinine, BUN, cholesterol, and triglycerides remained relatively consistent, indicating stable physiological status across different stages of growth. Immunologically, both humoral and cell-mediated responses were significantly enhanced at 12 weeks of age, confirming that immune competence strengthens as birds mature. These outcomes reinforce the importance of age-appropriate management, nutrition, and health monitoring to support optimal growth and immunity. The results also establish a valuable reference point for future research and practical strategies related to genetic selection, nutritional formulation, and disease prevention in turkeys reared under Indian conditions.

**Disclaimer (Artificial intelligence)**

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

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