**Antimicrobial Effect of Neem and Lemon Extract on Bacterial Pathogens Associated with Urinary Tract Infection in Pregnant Women attending Antenatal Clinics in Ekpoma, Nigeria.**

**ABSTRACT**

The high incidence of urinary tract infections (UTIs) amongst pregnant women as a result of physiological and hormonal changes during pregnancy, combined with the menace of antibiotic resistance to modern antibiotics, necessitates the introduction of novel alternatives to replace conventionally used antibiotics. This study investigated the antimicrobial effect of neem and lemon extract on bacterial pathogens associated with UTIs in pregnant women attending antenatal clinic at Ujeolen and Ihumudumu Health Centres in Ekpoma, Edo State. A two-week cross-sectional study was conducted using twenty (20) urine samples collected from pregnant women attending the antenatal care clinics. Bacteria causing UTIs were isolated and enumerated to assess the incidence and distribution of UTI pathogens. Isolated UTI pathogens were identified based on their biochemical properties. The crude extract of neem and lemon leaves were tested for their antimicrobial activity against the pathogens using 25, 50, 75, and 100 mg/mL concentrations. Bacterial colony count ranged from 2.4 x 105 - 4.6 x 105 CFU/mL (samples from Ujeolen) and from 3.1 x 105 - 5.2 x 105 CFU/mL (samples from Ihumudumu). Pathogens isolated from samples collected from Ujeolen Health Centre were identified as *E. coli*, *Klebsiella* sp*., Proteus* sp. and *Enterobacter* sp., while for Ihumudumu Health Centre, they were identified *E. coli*, *Klebsiella* sp*.* and *Pseudomonas* sp. All test isolates were susceptible to the extracts at the highest concentrations ranging from 75-100 mg/mL for both ethanolic extract of neem and lemon, while lower concentrations showed intermediate antimicrobial activity. The MIC of neem crude extract was 100 mg/mL for all test isolates and 50 mg/mL for the crude lemon extract. This study has confirmed the antimicrobial activity of neem and lemon crude extract on UTIs associated bacterial pathogens. Extracts of neem and lemon could serve as alternative therapy to conventional antibiotics in the treatment of UTIs.

**Keywords**: antimicrobial; lemon extract; neem extract; pathogens; pregnant women; urinary tract infections

**INTRODUCTION**

The excessive and indiscriminate use of antibiotics as emergency contraceptive that interfere with hormones changes and physiological activities during pregnancy had led to an increased in susceptibility and potentiate risks of contacting urinary tract infection among pregnant women [1]. Although UTIs are perceived as common amongst pregnant women they pose significant concern to both maternal and fetal health and if left untreated, can lead to complications such as preterm birth, low birth weight, pyelonephritis, and even maternal sepsis [2].

The rise in antibiotic resistance has outpaced the effectiveness of the current antibiotics, thus rendering many existing antibiotics ineffective [3-5]. According to the World Health Organization (WHO), this trend is considered the most pressing challenge in modern medicine [5]. Addressing this menace necessitates the introduction of novel approaches to tackle antibiotic resistance. Current research is exploring alternatives that utilize plant extract, specifically aimed at isolating and identifying novel bioactive chemicals with antimicrobial activity [6, 1], also given that nearly 50% of current nutraceuticals and pharmaceuticals are sourced from plants and their derivatives [6]. Although plant extracts hold promise in tackling this menace posed by antibiotic-resistant bacteria and despite the fact that the antimicrobial effect of most medicinal plants are unknown [7].

According to the WHO, 80% of the developing world still relies on plant-derived traditional medicine [8-9] and amongst the total estimated number of 374,000 plants [10], only 28,187 species are used by humans for medicinal purposes [11]. Additionally, 20,000 plants have been cataloged by the WHO as major sources for new drug development [5, 12]. Lewis *et al*. (2013) reported 30,000 well-defined antimicrobial extracts from over 1,340 plants [13]. Among these plants, lemon and neem leaves have been discovered to be rich in bioactive phytochemicals with promising antimicrobial properties [14].

The bioactive phytochemical constituents of neem (*Azadirachta indica*) include compounds such as azadirachtin, nimbin, and quercetin, which have exhibited antimicrobial and inflammatory-modulating effects [14]. These compounds inhibit enzymatic functions and disrupt the microbial cell membrane [15]. Similarly, lemon (*Citrus limon*) contains many limonoids, flavonoids, and citric acid, that are known to possessed antimicrobial properties [16]. Additionally, the acidic nature of lemon creates an adverse environment for bacterial growth [17]. Their accessibility, minimal side effects, and low cost represent a promising natural alternative to conventionally used antibiotics [18-19].

Pregnant women in semi-urban areas like Ekpoma, face an array notable health challenges spanning across, inadequate access to maternal healthcare services, shortage of trained maternal health personnel, and limited infrastructure that had led to hightened undiagnosed UTIs in pregnant women associated with complications like preterm labour or low birth weight [2]. The risk is heightened with current studies reporting asymptomatic cases of UTIs in pregnant women [20].

This study aimed to investigate neem and lemon extract’s effects on pathogens associated with UTIs isolated from pregnant women attending antenatal care at Ujeolen and Ihumudumu Health Centres, Ekpoma, Edo State.

**MATERIALS AND METHODS**

**Study Area**

This investigation was carried out at the Ujeolen and Ihumudumu Health Centres in Ekpoma, within the Esan West Local Government Area of Edo State and geographically, situated at a latitude of 6° 45' to 6° 0.1'N and a longitude of 6° 15' to 48°E.

**Sample Collection**

A total of 20 urine samples were randomly collected from pregnant women attending antenatal care at Ujeolen and Ihumudumu Health Centres in Ekpoma, Edo State. The 20 collected urine samples were collected as first morning urine specimens, which have increased test sensitivity and are typically more concentrated [21, 22]. A mid-stream catch method was employed for urine collection.

Fresh disease-free leaves of neem and lemon plants were collected from a farmland in Ekpoma in a sterile polythene bag.

Samples were transported immediately after collection to the Microbiology Laboratory at Ambrose Alli University for analysis.

**Isolation and Identification of Bacterial Isolates**

The streak plate method was used on nutrient and MacConkey agar for bacterial isolation. The plates were incubated at 37ºC for 24 hours and were subsequently subcultured to obtain a pure culture. The pure isolates obtained were identified based on their Gram staining and biochemical properties.

**Preparation of Plant Extract**

The cold percolation method was employed to prepare the plant extract. The plant leaves were processed into a fine powder and appropriately labeled. Sixty grams (60g) of the powder was soaked in 160 mL of anhydrous ethanol at 20°C for three consecutive days. Whatman No. 1 was used to filter the extract and the final filtrate was concentrated by evaporation at 50°C to obtain a crude extract via rotary evaporation. The extract was then stored in the refrigerator for subsequent evaluation [23].

**Preparation of Stock Concentration**

One gram (1g) of extract was dissolved in 10 mL of solvent to prepare a stock solution at 100 mg/mL concentration for both extracts. Varying concentrations were prepared from the main stock solution in the following proportions: 100, 75, 50 and 25 mg/mL.

**Antimicrobial Assay of Plant Extract**

The agar well diffusion method was used in Mueller-Hinton Agar (MHA) plates for the antimicrobial assay of plant extract. The test bacteria were cultured at 37°C overnight in a Nutrient broth to obtain a turbidity of 0.5 McFarland, equivalent to 1.5 × 108 CFU/mL. Additionally, a sterile 6 mm cork-borer was used to create six 5 mm wells, each filled with 50 μl of extract concentration ranging from 25-100 mg/mL plant extract. Dimethyl sulfoxide (DMSO) served as a negative/solvent control, while amikacin 30 mcg and nitrofurantoin 300 mcg acted as positive controls for isolates. After 30 minutes of diffusion at room temperature, the plates were incubated at 37°C for 18–24 hours, then checked for zones of inhibition (ZOI).

**Determination of MIC and MBC of the Plant Extracts**

The Minimum Inhibitory Concentration (MIC) was determined by the broth microdilution method following the Clinical and Laboratory Standards Institute (CLSI) guideline. A serial dilution of the plant extract in Mueller-Hinton agar was inoculated with bacteria (5 × 105 CFU/mL) and incubated at 37°C for 24 h. Amikacin was added as a positive control drug. After adding resazurin into the wells, the plates were incubated at 37°C for 30 minutes. Wells with bacterial growth turned pink; blue wells indicated inhibition. MIC is the lowest extract concentration, preventing growth [4].

**RESULTS**

**Bacteria Identification and Biochemical Characteristics**

Table 1 shows the biochemical characteristics of the bacterial isolate from urine samples of pregnant women attending antenatal care at Ujeolen Health Centre. The isolates were identified as *E. coli*, *Klebsiella* sp*., Proteus* sp. and *Enterobacter* sp. Table 2 shows the biochemical characteristics of bacteria isolated from urine samples obtained from pregnant women attending antenatal care at Ihumudumu Health Centre in Ekpoma. The isolates were identified as *E. coli*, *Pseudomonas* sp. and *Klebsiella* sp*.*

**Table 1:** Biochemical Characteristics of Bacteria Isolated from Urine Samples of Pregnant Women Attending Antenatal Care at Ujeolen Health Centre in Ekpoma

|  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Species | Gram | Motility | Catalase | Coagulase | Indole | Oxidase | Urease | Citrate | Glucose | Lactose | Mannitol | Sucrose |
| *Escherichia coli* | ­- | - | - | - | + | - | - | - | + | + | - | + |
| *Proteus* sp. | + | + | + | - | - | - | + | + | + | - | - | - |
| *Klebsiella* sp*.* | + | - | + | - | - | - | + | + | + | + | + | + |
| *Enterobacter* sp*.* | + | + | + | - | - | - | + | + | + | - | + | + |

**Keys: + =** Positive Reaction, **− = N**egative Reaction

**Table 2:** Biochemical Characteristics of Bacteria Isolated from Urine Samples of Pregnant Women Attending Antenatal Care at Ihumudumu Health Centre in Ekpoma

|  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Species | Gram | Motility | Catalase | Coagulase | Indole | Oxidase | Urease | Citrate | Glucose | Lactose | Mannitol | Sucrose |
| *Escherichia coli* | ­- | - | - | - | + | - | - | - | + | + | - | + |
| *Klebsiella* sp*.* | + | - | + | - | - | - | + | + | + | + | + | + |
| *Pseudomonas* sp*.* | - | + | - | - | - | - | + | - | + | - | + | - |

**Keys: + =** Positive Reaction, **− =** Negative Reaction

**Microbial Load in Urine Samples**

Table 3 presents the total colony counts of bacteria isolated from urine samples collected from pregnant women attending antenatal care at Ujeolen Health Centre, which ranged from 2.4 x 105 - 4.6 x 105 CFU/mL Ihumudumu Health Centre, which ranged from 3.1 x 105 - 5.2 x 105 CFU/mL.

**Table 3:** Bacterial Colony Count of Bacteria Isolated from Urine Samples of Pregnant Women Attending Antenatal Care at Ujeolen and Ihumudumu Health Centres in Ekpoma

|  |  |  |
| --- | --- | --- |
| Health Centre | Isolate code | CFU/mL |
| Ujeolen | U1 | 4.1 x 105 |
|  | U2 | 3.8 x 105 |
|  | U3 | 2.6 x 105 |
|  | U4 | 3.2 x 105 |
|  | U5 | 4.3 x 105 |
|  | U6 | 3.9 x 105 |
|  | U7 | 4.1 x 105 |
|  | U8 | 4.6 x 105 |
|  | U9 | 2.4 x 105 |
|  | U10 | 3.7 x 105 |
| Ihumudumu | I1 | 3.9 x 105 |
|  | I2 | 3.5 x 105 |
|  | I3 | 3.1 x 105 |
|  | I4 | 3.8 x 105 |
|  | I5 | 3.6 x 105 |
|  | I6 | 4.9 x 105 |
|  | I7 | 3.3 x 105 |
|  | I8 | 5.0 x 105 |
|  | I9 | 5.2 x 105 |
|  | I10 | 3.4 x 105 |

**Keys**: U1- U10 = Urine samples from Ujeolen Health Centre; I1- I10 = Urine samples from Ihumudumu Health Centre

**Frequency of Bacterial Occurrence**

Table 4 shows the frequency of bacteria isolated from the pregnant women. *E. coli* was the most occurring bacteria (55%) and *Proteus* sp. the least occurring (5%) bacteria isolated from the pregnant women at Ujeolen Health Centre, while *E. coli* was the most occurring (45.5%) and *Klebsiella* sp. the least occurring (18.2%) bacteria isolated from the pregnant women at Ihumudumu Health Centres.

**Table 4:** Frequency of occurrence of bacterial Isolates from urine samples of pregnant women attending antenatal care in Ujeolen and Ihumudumu Health Centres, Ekpoma

|  |  |  |  |
| --- | --- | --- | --- |
| Health centre | Isolate | Frequency | % |
| Ujeolen | *E. coli* | 11 | 55 |
|  | *Proteus* sp. | 1 | 5 |
|  | *Klebsiella* sp*.* | 5 | 25 |
|  | *Enterobacter* sp*.* | 3 | 15 |
| Ihumudumu | *E. coli* | 10 | 45.5 |
|  | *Klebsiella* sp*.* | 4 | 18.2 |
|  | *Pseudomonas* sp*.* | 8 | 36.4 |

**Antimicrobial Susceptibility of Plant Extract**

Table 5 present the antibiotic susceptibility test of ethanolic extracts of neem and lemon. All test isolates were susceptible to the extracts at the highest concentrations ranging from 75-100 mg/ml for both the ethanolic extract of neem and lemon.

**Table 5:** Antibiotic susceptibility test of the ethanolic extract of neem extracts against the tested bacterial strains from the urine samples

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Test isolate | Extract Concentration (mg/mL) | | | |
| 100 | 75 | 50 | 25 |
| *E. coli* | 28mm | 20mm | 20mm | 8mm |
| *Proteus* sp. | 28mm | 19mm | 20mm | 5mm |
| *Klebsiella* sp*.* | 26mm | 22mm | 10mm | 8mm |
| *Enterobacter* sp*.* | 27mm | 22mm | 10mm | 8mm |
| *Pseudomonas* sp*.* | 25mm | 20mm | 18mm | 7mm |

Zone of inhibition: Less than 13mm=Resistance; 14 to 17mm=Intermediate; 17 and above=Sensitive

**Table 6:** Antibiotic susceptibility test of the ethanolic extract of lemon extracts against the tested bacterial strains from the urine samples

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Test isolate | Extract Concentration (mg/mL) | | | |
| 100 | 75 | 50 | 25 |
| *E. coli* | 33mm | 28mm | 12mm | 11mm |
| *Proteus* sp. | 26mm | 22mm | 12mm | 7mm |
| *Klebsiella* sp*.* | 30mm | 29mm | 15mm | 10mm |
| *Enterobacter* sp*.* | 30mm | 25mm | 10mm | 8mm |
| *Pseudomonas* sp*.* | 31mm | 28mm | 13mm | 10mm |

Zone of inhibition: Less than 13mm=Resistance; 14 to 17mm=Intermediate; 17 and above=Sensitive

**Minimum Inhibitory Concentration (MIC) of Neem and Lemon Crude Extracts**

Tables 7 shows theMinimum Inhibitory Concentration (MIC)of neem and lemon crude extracts. The MIC of neem crude extract was 100mg/mL for all test isolates and 50 mg/mL for the crude lemon extract*.*

**Table 7:** Minimum Inhibitory Concentration of Test Isolates

|  |  |  |
| --- | --- | --- |
| Test isolate | Extract Concentration (mg/mL) | |
| Neem | Lemon |
| *E. coli* | 100 | 50 |
| *Proteus* sp. | 100 | 50 |
| *Klebsiella* sp*.* | 100 | 50 |
| *Enterobacter* sp*.* | 100 | 50 |
| *Pseudomonas* sp*.* | 100 | 50 |

**DISCUSSION**

This study investigated the antimicrobial effect of neem and lemon extract against UTIs associated bacteria isolated from pregnant women attending antenatal care clinics at Ujeolen and Ihumudumu, Ekpoma. This is in line with current research aimed at using crude plant extract as alternative therapy to conventional antibiotics.

Bacterial colony count ranged from 2.4 x 105 - 4.6 x 105 CFU/mL (samples from Ujeolen) and from 3.1 x 105 - 5.2 x 105 CFU/mL (samples from Ihumudumu), which is a significant amount (of ≥10⁴ CFU/mL), indicative of bacteriuria, indicating a potential UTI. The overall incidence of UTI amongst the pregnant women was relatively high across both hospitals. The finding is comparable to the report of studies by Okonko *et al*. [24] and Tadesse *et al*. [25] who reported a bacterial count of ≥10⁴ CFU/mL.

The bacterial isolates were *Escherichia coli, Proteus sp., Klebsiella* sp*.,* *Pseudomonas* sp., and *Enterobacter* sp*.* which are common uropathogens. *Escherichia coli* emergedas the most dominant isolate, accounting (55%) and *Proteus* sp. the least occurring (5%) bacteria isolated from the pregnant women at Ujeolen Health Centre, and same *E. coli* was the most occurring (45.5%) and *Klebsiella* sp. the least occurring (18.2%) bacteria isolated from the pregnant women at Ihumudumu Health Centres. This is consistent with the report that *E. coli* is a common uropathogen [26]. The frequency of bacterial isolates indicates the dominance of Gram-negative enteric bacteria in UTIs in the study, which is likely attributed to the hormonal and anatomical changes during pregnancy. Gram-negative bacteria possess a unique structure that facilitates attachment to the uroepithelium and prevents removal by urine flow, enhancing their pathogenicity [27]. Also, this unique feature enhances bacterial growth and tissue invasion, which can result in pyelonephritis occurring during pregnancy. Additionally, risk factors could be improper genital hygiene, direction of wipe after wash, lack of postcoital urination, and catheterization, as reported by Addo [28] and Arias [29].

The antibacterial activities of the crude extract of neem and lemon demonstrated high efficacy at high concentrations. The bacterial isolates showed sensitivity to crude extract of lemon at 100 mg/mL and 75 mg/mL and crude neem extract at 400 mg/mL and 300 mg/mL, however they showed reduced antibacterial activity at 50 mg/mL and complete resistance at 25 mg/mL for the crude lemon extract, similarly a reduced antibacterial activity was also observed for the crude extract of lemon at 200 mg/mL, and complete resistance at 100 mg/mL. These result support the potential use of crude extract of neem and lemon for the inhibition of *E. coli, Proteus sp., Klebsiella* sp*.,* *Pseudomonas* sp., and *Enterobacter* sp., as earlier reported [30,31].

**CONCLUSION**

This study has confirmed the inhibitory potential of neem and lemon crude extract in the management of UTIs caused by *E. coli, Proteus sp., Klebsiella* sp*.,* *Pseudomonas* sp., and *Enterobacter* sp. Therefore it can serves as alternative antimicrobial agents against the organisms used in this study to combat antibiotic resistant to UTIs among pregnant women.

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Disclaimer (Artificial intelligence)

Option 1:

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, manuscript.

**REFERENCES**

1. Gallucci N, Casero C, Oliva M, Zygadlo J, Demo M, Córdoba P. Interaction between Terpenes and Penicillin on Bacterial Strains Resistant to Beta-Lactam Antibiotics. Mol. Med. Chem 2006; 10:30-32.
2. Oladeinde BH. Prevalence and antimicrobial susceptibility pattern of asymptomatic urinary tract infection among pregnant women in Ekpoma, Nigeria. Journal of Basic and Clinical Reproductive Sciences. 2015; 4(1), 25–30.
3. Oliveira RG, Lima EO, Vieira WL, Freire KL, Trajano VN, Lima IO. Study of the interference of essential oils on the activity of some antibiotics used clinically. Rev. Bras. Pharmacogn 2006;16:77 82.
4. Locke JC, Lawson RH. Neem's Potential in Pest Management Programs, Proceedings of the USDA Neem Workshop. United States Department of Agriculture, Agricultural Research Service, ARS. 1990; 136.
5. Onoh RC, Umeora OU, Egwuatu VE. Antibiotic sensitivity pattern of uropathogens from pregnant women with urinary tract infection in Abakaliki, Nigeria. Infect Drug Resist 2013; 6:225.
6. Cunningham FG, Gant NF, Leveno KJ. Renal and urinary tract disorders. In: Seils A, Noujaim SR, Daris K, editors. Williams Obstetrics. 21st ed. New York, NY: McGraw-Hill Medical Publishing Division.2008;1251–1272.
7. Kaskoos RA. Essential oil analysis by GC-MS and analgesic activity of *Lippia citriodora* and *Citrus limon*. J. Essent. Oil-Bear. Plants.2019;22:273–281.
8. Wasserman S, Boyles T, Mendelson M. Pocket guide to antibiotic prescribing for adults in South Africa. Cape Town: South African Antibiotic Stewardship Program 2015; 1–60.
9. Van VF, Suliman S, Viljoen AM. The antimicrobial activity of four commercial essential oils in combination with conventional antimicrobials. Lett. Appl. Microbiol 2009;48:440 446.
10. Rodrigues E, Barnes J. Pharmacovigilance of herbal medicines: The potential contributions of ethnobotanical and ethnopharmacological studies. Drug Saf. 2103; 36:1-12.
11. Pandey A, Tripathi S. Concept of standardization, extraction, and pre-phytochemical screening strategies for herbal drug. J. Pharmacogn. Phytochem.2014;2:115–119.
12. Nicolle LE, Bradley S, Colgan R. Infectious diseases society of America guidelines for the diagnosis and treatment of asymptomatic bacteriuria in adults. Clin Infect Dis. 2005; 40(5):643–654. 10.1086/427507
13. Lewis DA, Gumede LY, Van der Hoven LA. Antimicrobial susceptibility of organisms causing community-acquired urinary tract infections in Gauteng province, South Africa. S Afr Med J. 2013;103(6):377–381. 10.7196/SAMJ.6722.
14. Biswas K, Chattopadhyay I, Banerjee RK, Bandyopadhyay U. Biological activities and medicinal properties of neem (*Azadirachta indica*). Current Science 2002;82(11): 1336-1345.
15. Subapriya R, Nagini S. Medicinal properties of neem leaves: a review. Current Medicinal Chemistry - Anti-Cancer Agents. 2005;5(2), 149–156.
16. Dahham, SS, Ali MN, Tabassum H, Khan M. Studies on antibacterial and antifungal activity of pomegranate (*Punica granatum* L.). Middle-East Journal of Scientific Research 2010;8(3), 219-224.
17. Etebu E, Nwauzoma, AB. A review on the phytochemical and pharmacological properties of lemon (Citrus limon). International Journal of Research in Pharmacy and Biosciences 2014;1(5): 1-13.
18. Nostro A, Germano MP, D’Angelo V, Marino A, Cannatelli MA. Extraction methods and bioautography for evaluation of medicinal plant antimicrobial activity. Letters in Applied Microbiology 2000;30(5): 379–384.
19. Cowan MM. Plant products as antimicrobial agents. Clinical Microbiology Reviews. 1999; 12(4): 564–582.
20. Awolude OA. Urinary tract infections among pregnant women in Nigeria: A neglected but preventable health issue. African Journal of Reproductive Health. 2021;25(3), 107–114.
21. Ouslander JG, Greengold BA, Silverblatt FJ, Garcia JP. An accurate method to obtain urine for culture in men with external catheters. Archives of Internal Medicine 1987;147(2): 286–288. <https://doi.org/10.1001/archinte.1987.00370020104045>
22. Brazier AM, Palmer MH. Collecting clean-catch urine in the nursing home: Obtaining the uncontaminated specimen. Geriatric Nursing 1995;16(5), 217–224. <https://doi.org/10.1016/s0197-4572(05)80167-3>
23. Okaba AE, Immanuel OM, Stow KM, Oku IY. Evaluation of the antimicrobial activities of selected plant extracts and honey against clinical isolates. African Scientist. 2022; 23(3): 175-179.
24. Okonko IO, Ijandipe, LA, Ilusanya OA, Donbraye-Emmanuel OB, Ejembi J, Udeze AO, Egun OC. Detection of urinary tract infection among pregnant women in Oluyoro Catholic Hospital, Ibadan, South-Western Nigeria. Malaysian Journal of Microbiology 2009;5(1), 11–17.
25. Tadesse A, Negash M, Ketema LS. Asymptomatic bacteriuria in pregnancy: assessment of prevalence, microbial agents and their antimicrobial sensitivity pattern in Gondar Teaching Hospital, northwestern Ethiopia. Ethiopian Medical Journal. 2007;45(2), 143–149.
26. Govindarajan, D.K.; Kandaswamy, K. Virulence factors of uropathogens and their role in host pathogen interactions. Cell Surf. 2022, 8, 100075.
27. Mancuso G, Midiri A, Gerace E, Marra M, Zummo S, Biondo C. Urinary Tract Infections: The Current Scenario and Future Prospects. Pathogens. 2023; 12:623. https://doi.org/10.3390/ pathogens12040623
28. Addo VN. Urinary Tract Infection in Pregnancy. Comprehensive Obstetrics in the Tropics. Dansoman: Asante and Hittscher Printing Press Limited. 2002; 261–267.
29. Arias F. Abnormalities of the urinary system during pregnancy. Practical guide to high-risk pregnancy and delivery: A South Asian perspective. 3rd ed. New Delhi: Elsevier 2008; 489–505.
30. Mosab AAM, Hadia AEA, Leila MAA, Ghanem MM. In vitro assessment of antimicrobial activity of citrus lemon against selected clinical isolates from Shendi city, Sudan. J Bacteriol Mycol Open Access. 2024;12(3):83-87. DOI: 10.15406/jbmoa.2024.12.00378
31. Jindal M, Chauhan S. In vitro comparison of the antibacterial activity of ethanolic extract of Azadirachta indica leaves with gentamycin, ampicillin, nitrofurantoin, and cotrimoxazole on bacterial pathogens isolated from urinary tract infection patients. 2017; 10:8.DOI: https://doi.org/10.22159/ajpcr.2017.v10i8.18557