**Original Research Article**

**Spatial Heterogeneity of Soil Micronutrients in Shivalik Foothill Zone of Himachal Pradesh: A Cross-Site Investigation of Soil Fertility**

**ABSTRACT**

Agricultural soils are subjected to constant management practices which influences their nutrient status throughout cropping cycles. Further, micronutrients play a crucial role in determining optimal crop growth and yield for majority of the commercial crop roster. Keeping this under consideration, an investigation was carried out for micronutrient status of soils of Indora block of district Kangra, Himachal Pradesh. Ninety-nine representative soil samples were collected from the selected sites. The profile samples were analyzed for DTPA micronutrients which was found to be sufficient in Cu and Fe. Whereas, for DTPA-extractable Zn and Mn it ranges from 1.12 to 4.30 mg kg-1and 2.04 to 7.20 mg kg-1. These findings emphasize the importance of soil testing and site-specific nutrient management to address potential deficiencies, optimization of crop productivity in the region and promote policy making protocols for the local farming community.

*Keywords****:*** *Micronutrients, Spatial variability, Soil fertility, Nutrient status, Sustainability, Soil survey*

1. **INTRODUCTION**

Micronutrients are essential trace elements that plants require in very small amounts, but they play a significant role in increasing crop growth, yield, and quality. When properly applied, micronutrients not only help in boosting agricultural productivity but also improve the nutritional quality of food and feed, which is important for human and animal health. Yield and quality of the crop can decrease if the supply of any one of these elements is inadequate, but crop species and cultivars vary considerably in their susceptibility to deficiencies (Shukla et al., 2019).

However, micronutrient deficiency has become a major concern globally. One of the main reasons is intensive farming, continuous cropping, and reduced use of organic amendments by the farming communities. These practices have led to a gradual decline in micronutrient levels in the soil (Sharma and Chaudhary, 2007). In India most of the soils are already low in fertility, especially in terms of micronutrients. Years of continuous cultivation without adequate micronutrient management and the heavy reliance on chemical fertilizers that provide only major nutrients like nitrogen (N), phosphorus (P), and potassium (K) have further deteriorated the soil condition and have deteriorated the soil micronutrient content and which ultimately adversely affects the crop growth (Sharma et al., 2023).

As a result, large areas of Indian agricultural land now exhibit deficiencies of important micronutrients such as zinc (Zn), iron (Fe), and boron (B). This not only affects crop production but also reduces the nutritional value of the food via depreciated bio-assimilation, thereby impacting the health of people and animals who depend on certain staple crops. Therefore, proper micronutrient management based on the crop type, soil condition, severity of deficiency, source, method, timing, rate, and frequency of application is essential for sustainable agricultural production and maintaining long-term soil and human health (Rahman et al., 2020). Productivity, stability and sustainability in many Indian soils have been affected by deficiency of micronutrients which may be either by primary (due to their low total contents) or secondary (caused by soil factors reducing their availability to plants) causes. Besides soil characteristics, the availability of micronutrients to plants is also influenced by their distribution within the soil profile (Singh et al. 1989). Therefore, it is important to study how agricultural management practices affect the availability of micronutrients and their accumulation in commonly consumed food crops.

Considering the aforementioned status and condition of the micronutrient mediated agricultural fertility domain the present study was conducted to evaluate the micronutrient status (Zinc, Copper, Iron and Manganese) of agricultural sites at block level (Indora, H.P., India) and execute subsequent descriptive statistical analysis.

1. **MATERIALS AND METHODS**

**2.1 Location and climatic condition:** The geographical setting of Kangra district lies within the coordinates of 31˚ 21′ to 32˚ 59′ N latitude and 75˚ 47′ to 77o 45’E longitude. This district is positioned on the southern slopes of the Himalayan Mountain range. The altitude in Kangra district varies significantly, ranging from approximately 500 meters above sea level to as high as 5000 meters above sea level. The total geographical area of Kangra district is 5,739 sq km, which is 10.31 per cent of the total area of the state. The average annual rainfall of district is 1751 mm.

**2.2 Study area:** Ninety-nine representative soil samples were collected at a depth of 0-20 cm from ninety-nine locations of Indora block of the farmers who were growing megha variety of cauliflower in more than 400m2 land. Soil samples thus collected during the month of August before transplantation of the crop, were air dried and passed through 2 mm sieve and stored in small muslin cloth bags for further analysis.

**2.3 Sample analysis:** Micronutrient concentrations in soil were determined by combining 10 g of air-dried soil with 20 ml of an extractant solution composed of 0.005M DTPA (Diethylenetriaminepentaacetic acid), 0.1M triethanolamine, and 0.01M calcium chloride, buffered at pH 7.3. The mixture was shaken for two hours to facilitate the extraction of plant-available zinc, iron, manganese, and copper. Following filtration, the concentrations of these micronutrients in the filtrate were quantified using atomic absorption spectrophotometry (novAA-350, Analytik Jena), allowing for an accurate assessment of their availability in the soil. (Lindsay and Norvell, 1978)

**Table 1. Critical limit for available micronutrients (Lindsay and Norvell, 1978)**

|  |  |
| --- | --- |
| **Availability** | **Micronutrients (mg kg-1)** |
| **Cu** | **Fe** | **Zn** | **Mn** |
| **Low** | 0.1-0.3 | 2-4 | 0.5-1 | 0.5-1.2 |
| **Medium** | 0.3-0.8 | 4-6 | 1-3 | 1.2-3.5 |
| **High** | 0.8-3 | 6-10 | 3-5 | 3.5-6 |
| **Very High** | >3 | >10 | >5 | >6 |

1. **RESULTS AND DISCUSSION**

Table 2. Status of DTPA-extractable Copper, Iron, Zinc and Manganese of selected sites of Indora block, H.P.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Sample No.** | **Village** | **DTPA-extractable Cu** | **DTPA-extractable Fe** | **DTPA-extractable Zn** | **DTPA-extractable Mn** |
| **(mg kg-1)** | **(mg kg-1)** | **(mg kg-1)** | **(mg kg-1)** |
| S1. | Sargotran | 0.92 | 23.38 | 1.3 | 2.25 |
| S2. | Sargotran | 1.10 | 25.38 | 1.35 | 2.32 |
| S3. | Sargotran | 0.85 | 20.32 | 1.28 | 2.35 |
| S4. | Pandohar | 1.65 | 32.56 | 2.55 | 3.24 |
| S5. | Pandohar | 1.38 | 28.95 | 1.66 | 2.91 |
| S6. | Pandohar | 1.25 | 27.75 | 1.58 | 2.85 |
| S7. | Badukhar | 1.47 | 29.65 | 1.78 | 3 |
| S8. | Badukhar | 1.00 | 24.41 | 1.38 | 2.3 |
| S9. | Badukhar | 1.40 | 30.12 | 2.2 | 3.05 |
| S10. | Samoon | 1.98 | 32.22 | 2.51 | 3.11 |
| S11. | Samoon | 1.53 | 31.25 | 2.19 | 2.37 |
| S12. | Baleer | 0.89 | 21.12 | 1.4 | 2.55 |
| S13. | Baleer | 1.34 | 26.65 | 1.51 | 2.62 |
| S14. | Tamladar | 1.47 | 31.22 | 2.26 | 2.38 |
| S15. | Madoli | 2.00 | 33.74 | 2.85 | 2.74 |
| S16. | Madoli | 0.97 | 24.1 | 1.32 | 2.17 |
| S17. | Tajwan | 2.11 | 30.95 | 2.18 | 2.32 |
| S18. | Tajwan | 0.88 | 20.11 | 1.21 | 2.1 |
| S19. | Tajwan | 2.85 | 35.56 | 3.55 | 5.52 |
| S20. | Lehrian | 2.45 | 34.14 | 3.21 | 5.35 |
| S21. | Lehrian | 2.15 | 28.68 | 2 | 2.52 |
| S22. | Sanaur | 0.92 | 22.45 | 1.51 | 2.8 |
| S23. | Sanaur | 0.90 | 22.4 | 1.41 | 2.23 |
| S24. | Sanaur | 1.45 | 31.12 | 2.23 | 2.65 |
| S25. | Paral | 0.95 | 21.44 | 1.38 | 2.11 |
| S26. | Kursan | 1.11 | 28.47 | 2.14 | 2.34 |
| S27. | Kursan | 2.48 | 33.12 | 2.61 | 3.12 |
| S28. | Kursan | 2.24 | 30.25 | 2.15 | 3.45 |
| S29. | Chanour | 1.65 | 28.21 | 1.89 | 3.64 |
| S30. | Chanour | 1.13 | 27.49 | 1.77 | 2.19 |
| S31. | Chanour | 1.69 | 29 | 2.18 | 2.66 |
| S32. | Ghurohan | 1.29 | 28.88 | 2.16 | 2.68 |
| S33. | Ghurohan | 1.38 | 30.2 | 2.06 | 2.41 |
| S34. | Dagla | 2.39 | 35.65 | 3.35 | 5.45 |
| S35. | Dagla | 0.87 | 21.12 | 1.37 | 2.34 |
| S36. | Dagla | 1.24 | 28.64 | 2.31 | 3.64 |
| S37. | Manwal jattan | 1.33 | 30.01 | 2.44 | 3.58 |
| S38. | Manwal jattan | 2.09 | 32.44 | 2.54 | 3.57 |
| S39. | Surdwan | 1.11 | 25.48 | 1.4 | 2.42 |
| S40. | Surdwan | 0.98 | 22.36 | 1.38 | 2.34 |
| S41. | Mijhli band | 2.58 | 33.58 | 2.66 | 3.55 |
| S42. | Thakurdwara | 2.68 | 33.74 | 2.71 | 4.21 |
| S43. | Thakurdwara | 1.57 | 29.98 | 2.54 | 3.66 |
| S44. | Mand miami | 0.89 | 21.14 | 2.12 | 3.22 |
| S45. | Mand miami | 3.01 | 38.14 | 3.85 | 5.65 |
| S46. | Ghoran | 2.95 | 37.89 | 3.77 | 3.88 |
| S47. | Ghoran | 1.84 | 30.27 | 2.45 | 2.69 |
| S48. | Ghoran | 1.39 | 28.86 | 2.23 | 2.44 |
| S49. | Ghoran | 0.99 | 22.23 | 2.02 | 2.36 |
| S50. | Bhapu | 3.00 | 38.05 | 3.87 | 2.65 |
| S51. | Bhapu | 2.27 | 31.16 | 2.68 | 2.37 |
| S52. | Bhapu | 2.55 | 32.23 | 2.74 | 2.66 |
| S53. | Bhapu | 1.69 | 29.65 | 2.39 | 3.44 |
| S54. | Bhapu | 1.84 | 29.89 | 2.52 | 3.65 |
| S55. | Mangwal | 0.85 | 21.54 | 1.36 | 2.14 |
| S56. | Mangwal | 1.95 | 30.14 | 2.66 | 3.68 |
| S57. | Mangwal | 2.00 | 30.54 | 2.71 | 3 |
| S58. | Indora | 3.05 | 37.45 | 4 | 6.32 |
| S59. | Indora | 2.33 | 36.65 | 3.66 | 5.52 |
| S60. | Indora | 1.54 | 28.74 | 2.68 | 5.12 |
| S61. | Indora | 1.00 | 27.47 | 2.54 | 4.11 |
| S62. | Indora | 1.29 | 28.24 | 2.39 | 3.54 |
| S63. | Indpur | 3.01 | 37.4 | 4.11 | 6.34 |
| S64. | Indpur | 2.56 | 31.47 | 2.7 | 3.65 |
| S65. | Indpur | 2.74 | 32.22 | 2.58 | 2.11 |
| S66. | Thath | 2.91 | 37.74 | 4 | 6.11 |
| S67. | Thath | 0.88 | 25.21 | 1.5 | 2.34 |
| S68. | Thath | 3.00 | 38.78 | 4.24 | 6.54 |
| S69. | Thath | 0.89 | 24.44 | 1.48 | 2.44 |
| S70. | Haler | 0.92 | 22.24 | 1.28 | 2.3 |
| S71. | Haler | 2.02 | 30.21 | 2.19 | 3.57 |
| S72. | Haler | 1.68 | 29.74 | 2.25 | 3.61 |
| S73. | Raja khasa | 0.99 | 20.54 | 1.5 | 2.04 |
| S74. | Gharoh | 1.78 | 28.85 | 1.66 | 2.45 |
| S75. | Gharoh | 2.32 | 31.12 | 2.68 | 4.12 |
| S76. | Gharoh | 3.11 | 38.89 | 4.3 | 7.2 |
| S77. | Gharoh | 1.87 | 29.16 | 2.35 | 2.66 |
| S78. | Gharoh | 2.65 | 32.25 | 2.45 | 2.45 |
| S79. | Gharoh | 2.85 | 33.54 | 2.55 | 3.56 |
| S80. | Purani gangath | 0.95 | 21.14 | 1.73 | 2.24 |
| S81. | Purani gangath | 0.88 | 21.03 | 1.68 | 2.31 |
| S82. | Purani gangath | 1.86 | 29.85 | 2.47 | 2.45 |
| S83. | Purani gangath | 3.00 | 38.25 | 4.28 | 7.15 |
| S84. | Purani gangath | 2.05 | 32.34 | 2.87 | 2.74 |
| S85. | Khadoli | 2.55 | 33.87 | 3.05 | 5.22 |
| S86. | Khadoli | 1.55 | 28.89 | 2.51 | 2.35 |
| S87. | Khadoli | 1.95 | 30.25 | 2.65 | 2.36 |
| S88. | Khadoli | 0.94 | 22.36 | 1.5 | 2.1 |
| S89. | Rohan | 1.15 | 24.35 | 1.8 | 2.11 |
| S90. | Rohan | 1.58 | 24.33 | 1.84 | 2.25 |
| S91. | Rohan | 2.70 | 34.12 | 3.21 | 5.32 |
| S92. | Taloti | 2.66 | 33.66 | 3.1 | 4.25 |
| S93. | Taloti | 2.11 | 32.21 | 2.89 | 3.66 |
| S94. | Indorian | 2.65 | 34.45 | 4.01 | 5.85 |
| S95. | Indorian | 1.60 | 28.74 | 2.74 | 2.85 |
| S96. | Tappa | 1.65 | 29.02 | 2.8 | 2.96 |
| S97. | Tappa | 1.45 | 28.88 | 2.85 | 2.88 |
| S98. | Gangwal | 2.35 | 35.87 | 4.14 | 6.45 |
| S99. | Gangwal | 2.95 | 36.22 | 4.21 | 7.11 |

\*DTPA: Diethylenetriaminepentaacetic acid.

Table 2. exhibits the assessed DTPA-extractable Copper, Iron, Zinc and Manganese levels of soil samples collected from the selected sites. The highest Copper content was detected at S10 (3.11 mg kg-1) and lowest Copper content was expressed by S3 (0.85 mg kg-1). Copper serves as a crucial component in processes such as hormone signalling, electron transport, Mitochondrial respiration and forms structural element in regulatory proteins (Rahman et al., 2020). Copper acts as driving factor for crop growth by directly influencing tillering and pollen viability of crops (Das, 2014). Primary anthropogenic sources of Copper addition to agricultural soils includes: Copper-rich fungicide, treated sewage sludge and liquid pig feed manure application (Panagos et al., 2018). Also, Copper concentration of agricultural soils positively correlates with degree of precipitation within the region and the age of the farming site, while it is inversely proportional to the increase in soil organic matter content (Neeman et al., 2024). Further, the highest and lowest Iron content was recorded from sites S76 (38.89 mg kg-1) and S18 (20.11 mg kg-1), respectively. Within plant physiology, Iron plays a crucial role in chlorophyll biosynthesis, photosynthesis, chloroplast development and cellular respiration (Kim and Guerinot, 2007; Gill and Tuteja, 2011). Moreover, inadequate Iron supply can lead to Iron deficiency in plants resulting in interveinal chlorosis in leaves which subsequently compounds to crop yield losses (Lan et al., 2011). Direct Iron addition to agricultural soils can be executed via addition of compound Iron fertilizers (CF) and sulphate iron (FeSO4). Iron can readily form exchangeable carbon-bound and Manganese-bound compounds which dictate iron availability within the soil matrix (Liu, 2002). Furthermore, the highest and lowest Zinc levels were exhibited by S76 (4.3 mg kg-1) and S18 (1.21 mg kg-1), respectively. Zinc influences processes such as stomatal function, cell membrane stabilization, water uptake and seed germination (Hassan et al., 2020). To a restricted degree, enzyme activation, gene expression, signal transduction and plant defence are also regulated by Zinc (Khan, 2022). Zinc sorption is higher in alkaline soil compared to acidic soils (Fan et al., 2018). Application of chelates such as Zn-EDTA and Zn-EDDS can enhance Zinc availability in pre-dominantly deficient soils and have a lower ecological impact due to their biodegradable properties (Múnera, 2022). Zinc is one of the defining nutritional factors determining the yield and quality of cauliflower crop as it enhances metabolism and improves pathogen resistance (Parveen et al., 2022). The combined application of Fe and Zn can lead to a synergistic effect, where the presence of one micronutrient enhances the uptake and utilization of the other. This is particularly evident in soils that are deficient in both elements, where their co-application mobilizes native nutrients and increases their bioavailability to plants (Choudhary et al., 2015). Lastly, the highest and lowest Manganese concentration was detected at S76 (7.2 mg kg-1) and S73 (2.04 mg kg-1), respectively. Manganese primarily plays a significant role in oxidation and reduction processes in plants (electron transport during photosynthesis) and is an essential component of Photosystem-II (Mousavi et al., 2011). Manganese fertilization can improve crop yield by augmenting photosynthetic efficiency of the plant (Mousavi et al., 2007). Improved photosynthetic efficiency positively correlates to the biomass accumulation capacity of the plant which directly influences the overall yield. Commercial sources for addition of Manganese to agricultural platforms include: Manganese sulphate, Manganese oxide and organic amendments (Rashed et al., 2021). Moreover, microbial interactions have proved to increase the availability of innate soil Manganese via causing reduction system pH and organic acid production (Khoshru et al., 2023).

Fig. 1. Site-wise micronutrient status of Indora block, H.P.

\*DTPA: Diethylenetriaminepentaacetic acid.

Table 3. Descriptive statistical measures of soil micronutrients

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  **Statistical property**  | **DTPA-extractable Cu** | **DTPA-extractable Fe** | **DTPA-extractable Zn** | **DTPA-extractable Mn** |
| **Range** | 0.85 - 3.11 | 20.11 - 38.89 | 1.21 - 4.3 | 2.04 - 7.2 |
| **Mean** | 1.79 | 29.52 | 2.43 | 3.37 |
| **SE±** | 0.07 | 0.5 | 0.08 | 0.14 |
| **C.V(%)** | 39.65 | 16.91 | 34.26 | 40.32 |

**\*** DTPA: Diethylenetriaminepentaacetic acid; SE±: Standard error; C.V. (%): Coefficient of variation.

* 1. **DTPA-extractable Copper**

The data presented in table 3 reveal that the DTPA-extractable copper content ranged from 0.85 to 3.11 mg kg-1 with mean value of 1.79 mg kg-1. The per cent distribution of DTPA-extractable copper content shows that 100 per cent of the soil samples were found to be in high category. Mahajan *et al.* (2007) also reported high levels of DTPA-extractable Cu in the soils of Himachal Pradesh. Comparatively higher copper content may be due to higher organic carbon because Cu forms Cu-humus complex of relatively high stability with humus that decreases its susceptibility to fixation or precipitation in the soil. The results are in the line with those of Thakur and Bhandari (1986), Mahajan (2001) and Sharma and Kanwar (2010).

* 1. **DTPA-extractable Iron**

Table 3 further reveals that the DTPA-extractable Fe content ranged from 20.11 to 38.89 mg kg-1 with mean value of 29.52 mg kg-1. The percentage distribution of DTPA-extractable Fe content shows that 100 per cent of the soil samples were found to be in very high category which may be attributed to neutral to slightly alkaline soil reaction beside presence of gypsiferrous and ferruginous parent materials containing hematite and limonite minerals (Wadia, 1966). Mahajan *et al.* (2007) also found that the soils of North West Himalayas were high in DTPA- extractable Fe content. Chander *et al.* (2014) and Kakar *et al.* (2018) reported that the soils of different zones of Himachal Pradesh were high in DTPA-extractable iron content.

* 1. **DTPA-extractable Zinc**

The data in table 3 reveals that the DTPA-extractable Zn ranged from 1.21 to 4.3 mg kg-1 with mean value of 2.43 mg kg-1. The percentage distribution of DTPA-extractable Zn content is illustrated in fig. 2 which shows that 81.00 per cent of the soil samples were found to be in medium category while 19.00 per cent were found to be in high category.Thakur and Bhandari (1986) reported that DTPA extractable Zn varied from 0.56 to 6.76 ppm in Saproon valley of Himachal Pradesh, while Mahajan (2001) found high levels of zinc in Mandi district soils.

Fig. 2. DTPA-extractable Zinc percentage distribution

* 1. **DTPA-extractable Manganese**

A perusal of data in table 3 reveals that the DTPA-extractable Mn content ranged from 2.04 to 7.2 mg kg-1 with mean value of 3.37 mg kg-1. The percentage distribution of DTPA-extractable Mn is illustrated in fig. 3 which shows that 55.00 per cent of the soil samples were in medium category while, the remaining 45.00 per cent were in high category. Similar values of DTPA extractable Mn were also observed by Behera and Shukla (2013), Dixit (2021) and Kumar *et al.* (2022) in soils of Himachal Pradesh.

Fig. 3. DTPA-extractable Manganese percentage distribution

1. **CONCLUSION**

India is facing reduced crop yields and lower micronutrient levels in food due to the intensive farming and modern cropping practices with high yielding crop cultivars and unbalanced fertilizer application which ultimately results in widespread deficiency of micronutrients. While, Zn deficiency has declined in soils of the country because of regular and more use of Zn fertilizer whereas deficiency of Fe and Mn increased slightly which can be overcome by spraying of suitable chemicals at recommended levels by foliar application which reduce the deficiency of these nutrients in the soil. Therefore, the information obtained by this study about micronutrients status would be useful for the farmers in developing nutrient management techniques for better and long-term soil health sustainability.

**REFERENCES**

Behera, S. K., & Shukla, A. K. (2013). Depth-wise distribution of zinc, copper, manganese and iron in acid soils of India and their relationship with some soil properties. Journal of the Indian Society of Soil Science 61(3), 244-252.

 Chander, G., Sharma, S., Sharma, V., Verma, S., Sharma, S., & Verma, T. S. (2014). Micronutrient cations status in vegetable growing soils of subhumid and wet-temperate zones of Himachal Pradesh. Himachal journal of Agriculture Research 40, 79-83.

Choudhary, H. D., Sharma, S. R., Jat, R. S., & Jat, G. (2015). Effect of soil and foliar application of zinc and iron on yield, quality and economics of fennel. Annals of plant and Soil Research, 17(2), 200-203.

Das, S. K. (2014). Role of micronutrient in rice cultivation and management strategy in organic agriculture - A reappraisal. Agricultural Sciences, 5(9): 765.

 Dixit, (2021). Assessment of soil health under vegetable cultivation in sub-montane and subtropical zone of Himachal Pradesh. M.Sc. Thesis. Dr. Yashwant Singh Parmar University of Horticulture and Forestry, Nauni, Solan, Himachal Pradesh, India.

Fan, T. T., Cui, P. X., Sun, Q., Zhou, D. M., Alves, M. E., & Wang, Y. J. (2018). Combining path analysis and x‐ray absorption spectroscopy to unravel the Zn sorption mechanism on soils. Soil Science Society of America Journal, 82(4), 796-802.

Gill, S. S., & Tuteja, N. (2011). Cadmium stress tolerance in crop plants probing the role of sulfur. Plant Signal. Behav. 6, 215–222. doi: 10.4161/psb.6.2.14880

Hassan, M. U., Aamer, M., Chattha, M. U., Haiying, T., Shahzad, B., Barbanti, L., Nawaz, M., Umair Hassan, M., Aamer, M., Umer Chattha, M., Haiying, T., Shahzad, B., Barbanti, L., & Guoqin, H. (2020). The critical role of zinc in plants facing the drought stress. Agriculture, 10(9), 396.

 Kakar, R., Tripathi, D., Chandel, S., & Sultanpuri, A. (2018). Nutritional status of tomato leaves in North Western Himalayas. Annals of plant and soil research 20, 187-190.

Khoshru, B., Mitra, D., Nosratabad, A. F., Reyhanitabar, A., Mandal, L., Farda, B., & Mohapatra, P. K. D. (2023). Enhancing manganese availability for plants through microbial potential: A sustainable approach for improving soil health and food security. Bacteria, 2(3), 129-141.

Kim, S. A., & Guerinot, M. L. (2007). Mining Iron: Iron uptake and transport in plants. FEBS Lett. 581, 2273–2280. doi: 10.1016/j.febslet.2007.04.043

 Kumar, A., Arshad, M., Singh, S. P., & Sepehya, S. (2022). Macronutrient status and their relationship with leaf nutrient content of cauliflower (Brassica oleraceae var. botrytis) growing areas of Kunah river flood plains in Hamirpur Himachal Pradesh. Journals of Soils Crops 32, 45-58.

Lan, P., Li, W., Wen, T. N., Shiau, J. Y., Wu, Y. C., and Lin, W. (2011). iTRAQ protein profile analysis of Arabidopsis roots reveals new aspects critical for iron homeostasis. Plant Physiol. 155, 821–834. doi: 10.1104/pp.110.169508

Lindsay, W. H., & Norvell, W. A. (1978). Development of DTPA soil test for zinc, iron, manganese and copper. Soil Science Society of American Journal 42, 421-428.

Liu, W. (2002). Study on the transformation and iron provision mechanism of iron fertilizers in the calcareous soil. Plant Nutrition and Fertilizing Science.

 Mahajan, A. (2001). Characterization of Balh valley soils of District Mandi. M.Sc. Thesis. Department of Soil Science, CSK Himachal Pradesh Krishi Vishvavidyalaya, Palampur, India. 133p.

Mahajan, A., Sharma, S. K., Gupta, R. D., & Sharma, R. (2007). Morphological, physical and chemical properties of soils from North West Himalayas. Bulgarian Journal of Agricultural Science 13, 607-618.

Mousavi, S. R., Galavi, M., & Ahmadvand, G. (2007). Effect of zinc and manganese foliar application on yield,

Mousavi, S. R., Shahsavari, M., & Rezaei, M. (2011). A general overview on manganese (Mn) importance for crops production.

Muhammad, F., Haq, M. M. U., Khan, M. D., Shafique, M., Bashir, S., Alizai, S. U., & Raza, M. M. (2022). Unlocking the potential of zinc (zn) efficiency for enhanced crop production in low-zn environments. Indus Journal of Agriculture and Biology, 1(1), 33-45.

Neaman, A., Schoffer, J. T., Navarro-Villarroel, C., Pelosi, C., Peñaloza, P., Dovletyarova, E. A., & Schneider, J. (2024). Copper contamination in agricultural soils: A review of the effects of climate, soil properties, and prolonged copper pesticide application in vineyards and orchards. Plant, Soil & Environment, 70(7).

Panagos, P., Ballabio, C., Lugato, E., Jones, A., Borrelli, P., Scarpa, S., & Montanarella, L. (2018). Potential sources of anthropogenic copper inputs to European agricultural soils. Sustainability, 10(7), 2380.

Parveen, K., Kumar, N., Sharma, D., & Ledwani, L. (2022). Evaluation of efficacy of “Cassia renigera” leaf extract mediated ZnO nanoparticles as nano fertilizer for cauliflower plant. ChemistrySelect, 7(23), e202200367.

Pinto, W. S. (2021). Soil zinc availability from inorganic and chelated zinc sources and its accumulation in maize, evaluated by Kinetica: a computational modeling platform.

quality and enrichment on potato (Solanum tuberosum L.). Asian Journal of Plant Sciences, 6, 1256-1260.

Rahman, R., Sofi, J. A., Javeed, I., Malik, T. H., & Nisar, S. (2020). Role of micronutrients in crop production. International Journal of Current Microbiology and Applied Sciences, 8, 2265-2287.

Rahman, R., Sofi, J. A., Javeed, I., Malik, T. H., & Nisar, S. (2020). Role of micronutrients in crop production. International Journal of Current Microbiology and Applied Sciences, 8, 2265-2287.

Rashed, M. H., Hoque, T. S., Jahangir, M. M. R., & Hashem, M. A. (2019). Manganese as a micronutrient in agriculture: crop requirement and management. Journal of Environmental Science and Natural Resources, 12(1-2), 225-242.

Sharma, A., Sepehya, S., Sharma, R., & Kumar, A. (2023). Impact of various cropping systems on the status and distribution of selected micronutrients in subtropical region of Himachal Pradesh, India. Int. J. Plant Soil Sci, 35(19), 454-463.

Sharma, J. C., & Chaudhary, S. K. (2007). Vertical distribution of micronutrient cations in relation to soil characteristics in lower Shiwaliks of Solan district in North-West Himalayas. Journal of the Indian society of soil science, 55(1), 40-44.

 Sharma, V., & Kanwar, B. B. (2010). Copper status and its relation with soil properties in pea growing soils of high hills dry temperate zone of Himachal Pradesh. Indian Journal of Agricultural Research 44, 32-37.

Shukla, A. K., Behera, S. K., Satyanarayana, T., & Majumdar, K. (2019). Importance of micronutrients in Indian agriculture. Better Crops South Asia, 11(1), 6-10.

Singh, K., Kuhad, M. S., & Dhankar, S. S. (1989). Influence of soil characteristics on profile distribution of DTPA-extractable micronutrient cations. Indian Journal of Agricultural Sciences 59, 331-334.

Thakur, N. R., & Bhandari, A. R. (1986). Available nutrient elements status of soils of a temperate vegetable seed producing valley of Himachal Pradesh. Journal of the Indian Society of Soil Science 34, 428-30.

 Wadia, D. N. (1966). Geology of India. SLBS and McMillan and Company. London. 383p.