**A Brief Review on the Invasive Fall armyworm, *Spodopetra frugiperda* (J.E.Smith)**

**Abstract**

The fall armyworm (*Spodoptera frugiperda*), a highly destructive and invasive pest species, has rapidly become a major concern for global food production systems. Originally endemic to the Americas, this noctuid moth has aggressively expanded its geographical range causing widespread crop losses and threatening the livelihoods of millions of smallholder farmers. Known for its polyphagy, the fall armyworm feeds on over 80 host plant species, with a strong preference for cereal crops such as maize, rice, sorghum, and wheat. Its ability to migrate over long distances, coupled with rapid lifecycle turnover and resistance development to common chemical pesticides, makes *S. frugiperda* a formidable pest in both rainfed and irrigated agro-ecosystems. The species exhibits two genetically distinct strains—the rice strain and the corn strain—which differ in host preference and geographical behavior, complicating control measures. Moreover, the lack of early warning systems and limited access to effective control technologies in newly invaded regions exacerbates the pest's impact. This review aims to provide a concise overview of the taxonomy, biology, invasion history and all further details about *Spodoptera frugiperda*, emphasizing the importance of sustainable and adaptive pest management approaches in the face of ongoing agricultural challenges.

Keywords: ***FAW, INVASIVE, ENTOMOLOGY, SPODOPTERA, NOCTUID, LEPIDOPTERA***

1. **Introduction**

Maize, *Zea mays* considered as Queen of Cereals, is grown for a variety of purposes like human consumption, cattle and poultry feed in India. Industrial processing may also include extraction of starch, corn syrup, dextrose and also corn oil. Being a crucial staple food, it is cultivated in among 160 countries with a diverse soil and climatic conditions [**Mishra et al. 2024**]. *Zea mays* contribute about 36% of global grain production [**Malleta, 2024**]. According to existing reviews it is seen that almost 13% of total production is used for industrial processing, 24% is consumed as human food, 11% as cattle feed and 52% as poultry feed [**Dass 2013**]. Development of Quality Protein Maize using the OPAQUE 2 Gene has overcome the deficiency of essential amino acids [**Maitra et al. 2019**].

Southern states of India such as Karnataka and Madhya Pradesh account a highest percentage of maize plantation. It is prominently seen that during 2020-2021 about 9.86 million ha of maize was grown yielding almost 31.50 mt and 29.5 q/ha productivity in India [**Mishra et al. 2024**]. In 2050, Projected Demand for maize is estimated as 121 mt [**Amarasinghe and Singh, 2008**]. Usually, it is grown as a rainfed crop under residual moisture conditions in India. In Odisha, it is evidently found that southern states are more actively involved in cultivation of maize. Cultivated area is about 2.8 lakh hectare with production of about 7.79 lakh tonne of maize with a productivity of about 2785 kg/ha [**Odisha Agricultural Statistics, 2013-2014**]. Districts like Boudh, Gajapati, Ganjam, Kalahandi, Malkangiri, Koraput, Nabrangpur, Kandhamal, Rayagada and Nuapada is considered to be the production hub of maize in Odisha [**Maitra et al. 2019**].

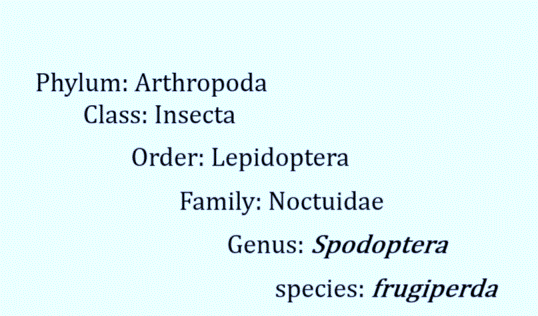
Despite its great contribution to the economy, it is seen that almost 141 insect pests cause a varying degree of damage to the crop from the stage of sowing to harvest **[Reddy and Trivedi, 2008].**  A newly introduced pest, the invasive fall armyworm is considered to be creating havoc in the maize production. *Spodoptera frugiperda* (Noctuidae: Lepidoptera) is a notorious, polyphagous pest feeding voraciously on more than 80 crops and plant species [**Prasanna, 2018**]. The name *Spodoptera frugiperda* is derived from the feeding habits of the larval lifestage, *frugiperda* in Latin means ‘LOST FRUIT', as the pest can cause severe damage and yield loss. It is also seen that almost 353 plants can be referred as the host of the invasive FAW [**Kansiime et al.2019**] Prominent damage is seen during Larval stage feeding in leaves making papery windows subsequently defoliating the plant [**Reddy et al. 2019**]. The insect has army marching behaviour feeding voraciously on the plant preferably during the night and midnight hours. Recent studies on FAW, have revealed that an adult can fly over long distances of about 300 miles.

It is native to Tropical and Subtropical regions of America. In 2016, it was first reported in Africa [**Sisay et al. 2018**]. Currently, the pest has reached to almost 44 countries [**Sisay et al. 2019**]. Through commercial aircrafts and cargo containers, the pest has spread from America to Africa, and later wind acted as the dispersal medium [**Day et al. 2018**]. In Indian continent, the first report of FAW was seen in Karnataka during 2018. Later, it spread to states like Bihar, Odisha, Chhattisgarh, WB [**CABI,2020**]. In India, a sum total of 170,000 ha of maize crops was affected by FAW among 10 different states [**Sangomla and Kukreti, 2019**]

**2. THE INVASIVE FALL ARMYWORM, *Spodoptera frugiperda***

**2.1 Taxonomic Position**

According to **Nagoshi et al. 2007,** two strains of FAW are found such as Rice strain and Corn Strain. Rice strain feeds on Rice and related species, while the corn strain feeds on maize, cotton and sorghum.The systematic taxonomic position is given below:

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**2.2 Biology of Fall armyworm**

As it is a lepidopteran pest the lifecycle of FAW consists of four stages: egg, larva, pupa and adult. The duration of lifecycle varies with season. In summer season, 30 days, while in autumn and spring the lifecycle is completed in 60 days and when the temperature falls in winter the lifecycle is completed in about 90 days.

**2.2.1 Egg Stage**

The eggs of FAW is generally dome shaped with a flattened base. The eggs are creamy white in colour with reticulate ribs, covered by abdominal hairs which is grey to pink in colour which protects the eggs. These are the anal tuft scales or setae. An adult female lay about 100-200 eggs in a mass. Eggs are mostly laid on the upper and lower sides of the leaf. In some cases, eggs were also found in stalk and funnel of maize plant. [**Prasanna et al. 2018; Bajracharya and Bhat, 2019; CABI,2018b].** In 20-30 degree Celsius, the eggs are matured in 2-3 days.

**2.2.2 Larval Stage**

It is considered as the most destructive stage. The newly hatched larvae at first and second instars are green in colour, which gradually turns to blackish brown in colour when it is in third to sixth instars. A Prominent inverted Y shaped mark is found on the front. The epidermal pattern is rough and may have granular texture with four dark raised spots in form of square in the front. The newly hatched larvae is voracious and has burrowing behaviour [**Prasanna et al. 2018**].

**2.2.3 Pupal Stage**

After 14 days of complete larval cycle it falls to the ground. Pupa is of oval shape generally reddish brown in colour present inside a cocoon located at a depth of 2-10 cm in soil [**Prasanna et al. 2018, CABI,2018b**]. The cocoon is made by the binding soil particles. In case, if the soil is hard, the leaf debris along with other materials are webbed together on the soil surface,

**2.2.4 Adult Stage**

Emerged adults are nocturnal while the colour may vary. Male adults have shaded grey and brown forewing with prominent white triangular spots at the tip and centre of wing, this feature is restricted to males. The wingspan may be of 40mm. As moths are migratory, having ability to fly long aerial distances [**Prasanna et al. 2018**].

**2.3 Damage Symptoms and Severity:**

In Maize, the severity starts after the eggs are hatched. The typical symptoms is the formation of papery windows of variable sizes and rugged edges with oval round appearance on the leaves, making them detached from the plant. Eventually, depending on the infestation severe defoliation can be observed with excessive faecal material left out in the plants. Severe defoliation is also the result of voracious feeding by the larval instars. After these symptoms the growth of the crop is eventually stopped leading to malformed development of the cob or tassel [**Reddy,2019**]. Depending on the Instars the symptoms vary, for example translucent patches are seen when it is generally fed by 1st and 2nd Instar larvae while elongated large hole is seen when it is infested by larvae of 3rd to 6th Instars. The most detrimental and devastating stage of the pest is the Larval stage. Depending on growth stage of the plant, the larvae can be seen in different parts of the maize crop including Leaf whorls, young leaves, tassels and cob [**Goergen er al. 2016**]**.** An average of 11.57% of yield reduction is generally seen when the incidence of the pest is in between the range of 27% - 56% [**Buadron et al. 2019**]. In 2019, **Chimweta et al.2019** revealed that almost 60% of yield reduction is seen when the damage to leaf, silk and tassel range from 25-50%.

**Table 1: Scale for the leaf damage assessment of the crop through fall armyworm (*S. frugiperda*)**

|  |  |  |
| --- | --- | --- |
| **Scale Description** | | **Pictorial Representation** |
| **0** | **No visible leaf damage** |  |
| **1** | **Only pinhole damage on leaves** |  |
| **2** | **Pinhole and shot hole damage to leaf** |  |
| **3** | **Small elongated lesions (5-10 mm) on 1-3 leaves** |  |
| **4** | **Mid-sized lesions (10-30 mm) on 4-7 leaves** |  |
| **5** | **Large elongated lesions (>30 mm) or small portions eaten on3-5 leaves** |  |
| **6** | **Elongated lesions (>30 mm) and large portions eaten on 3-5 leaves** |  |
| **7** | **Elongated lesions (>30 cm) and 50% of leaf eaten** |  |
| **8** | **Elongated lesions (30 cm) and large portions eaten on 70 % of leaves** |  |
| **9** | **Most leaves with long lesions and complete defoliation observed** |  |
| **Source: Sagar et al., 2020; Prasanna et al., 2018** | | |

**2.3 Favourable Condition for development of the pest:**

The outbreaks and the intensity of damage by the pest is often governed by climatic factors. Characteristics such as Mortality, Growth, abundance, number of generations and successful survival depends on environmental conditions. [**Ramirez et al, 2017**]. Overwintering mechanism governs the greater invasion of the pest. It can thrive well in wet and cool weather conditions. Outbreak Severity is seen after heavy rainfall with humid weather conditions [**Westbrook and Sparks, 1986**]. For the successful reproduction and survival of the pest, a warm and humid growing season with high rainfall are best suited. Generally, when the temperature is less than 10  
 degree Celsius the development is ceased [**Assefa and Ayalew, 2019**]. In tropical areas almost ten generations are found as compared to only two generations in temperate areas which indicates warm temperature conditions favours increasing of the generations [**Assefa and Ayalew, 2019**]. It is also seen that for pupation and adult emergence a threshold temperature of about 10.9 degree Celsius is favourable [**CABI, 2020**]. When the temperature is about 21-26 degree Celsius the egg hatches within two to four days. For larval development a temperature of 28 degree Celsius is preferred while, the threshold temperature of about 14.6 degree Celsius is considered optimum for pupation. Litreature studies have also revealed that if the temperature increases to about 30 degree Celsius the wings are deformed [**Assefa and Ayalew, 2019**].

**2.4. Natural Enemies of FAW**

The larvae and eggs of *S.frugiperda* is parasitised by a number of insects. Listed in the Table 2 and 3.

**Table 2 : Indicating list of Parasitoids, Predators and Entomopathogens**

|  |  |  |
| --- | --- | --- |
| **Parasitoids** | | |
| 01 | *Apanteles marginiventris* | **Sisay, 2018; Pokhrel et al. 2022** |
| 02 | *Campoletis grioti* |
| 03 | *Chelonus insularis* |
| 04 | *Meteorus autographae* |
| 05 | *Ophion* spp. |
| 06 | *Eiphosoma vitticolle* |
| 07 | *Cotesia icipie* |
| **Predators** | | |
| 01 | Ground Beetles | **Sisay, 2018; Pokhrel et al. 2022** |
| 02 | *Labidura riparia* |
| 03 | *Podisus maculiventris* |
| 04 | *Orius insidiosus* |
| **Entomopathogens** | | |
| Fungi | *M.anisopliae* | **Sisay, 2018; Pokhrel et al. 2022** |
| *B. bassiana* |
| Virus | *S. frugiperda* multiple nucleo polyhedrovirus |
| Bacteria | *B. thuringiensis* |

**Table 3 : Indicating list of Natural Enemies and their effective Host stage**

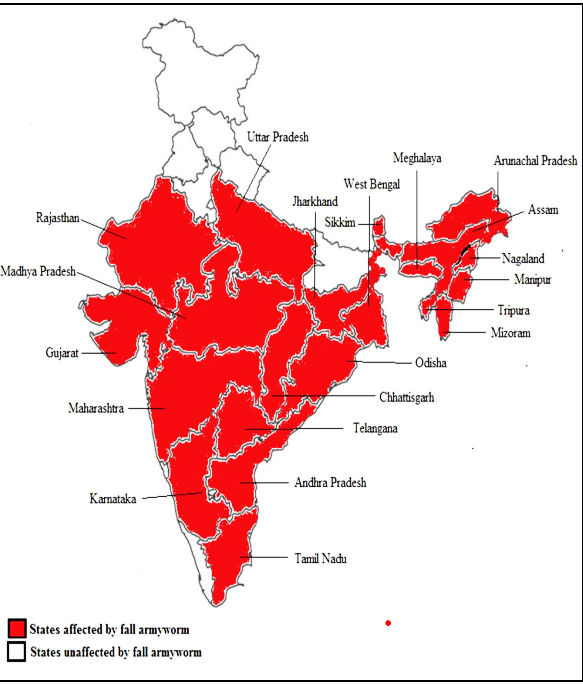
|  |  |  |
| --- | --- | --- |
| **S. No.** | **Scientific name** | **Host stage** |
| 1 | *Telenomus* sp. | Egg parasitoid |
| 2 | *Trichogramma sp* | Egg parasitoid |
| 3 | *Glyptapanteles creatonoti* (Viereck) | Larval parasitoid |
| 4 | *Forficula sp.* | Larval predator |
| 5 | *Coccygidium melleum* (Roman) | Endo larval parasitoid |
| 6 | *Campoletis chlorideae* Uchida | Endo larval parasitoid |
| 7 | *Eriborus* sp | Endo larval parasitoid |
| 8 | *Odontepyris* sp. | Larval parasitoid |
| 9 | *Exorista sorbillans* (Wiedemann) | Endo larval parasitoid |
| 10 | *Forficula* sp. | Predator |
| 11 | *Harmonia octomaculata* (Fabricius) | Predator |
| 12 | *Coccinella transversalis* Fabricius | Predator |
| 13 | *Nomuraea rileyi* (Farlow) Samson | Entomo-pathogen on larva |
| 14 | Nucleopolyhedrovirus (NPV) | Virus on larva |
| Repalle et al., 2020 | | |

**2.5 Present Status of FAW in India.**

FAW with its great ability spreads into new territory moving east and north in India. In mid-May of 2017, it was first reported by Sharanabasappa et al, 2018 in Shivamogga, Karnataka. Now, FAW is reported from Tamil Nadu, Odisha, AP. Maharastra, Telangana, Gujarat.

TABLE 4. **The Table shows the fall armyworm-affected Indian states during the month and year**

|  |  |
| --- | --- |
| **Name of the Indian states** | **Month and Year** |
| Karnataka | May 2018 |
| Chhattisgarh | August 2018 |
| Gujarat | September 2018 |
| Tamil Nadu | November 2018 |
| Maharashtra | September 2018 |
| Odisha and West Bengal | 2018 |
| Andhra Pradesh | March, 2019 |
| Mizoram, Nagaland, Tripura, Manipur and Meghalaya | May 2019 |
| Arunachal Pradesh | June 2019 |
| Telangana | 2019 |
| Madhya Pradesh, Uttar Pradesh, Assam, Sikkim, Rajasthan and Jharkhand | 2019 |
| Repalle et al., 2020 | |



**Figure 1. Indicating the states of India affected by FAW**

**2.6. Cause of Dominance**

The dominance of *Spodoptera frugiperda* (fall armyworm) can be attributed to several key biological and ecological traits. It is highly adaptable to a wide range of temperatures, with cloud cover, moderate rainfall, and low temperatures accelerating population outbreaks. Its short life cycle, multivoltine nature (10–12 generations per year), and absence of diapause allow for continuous reproduction. The larvae exhibit cryptic behavior, feed gregariously, and are capable of cannibalism, which enhances survival. Additionally, the pest has a high reproductive potential and a broad host range, being oligophagous, enabling it to thrive on multiple crops. These features collectively contribute to its rapid spread and dominance in diverse agro-climatic regions

**Conclusions**

The global emergence of *Spodoptera frugiperda* highlights the complex challenges posed by invasive species in modern agriculture. Its rapid spread, high reproductive capacity, and wide host range have resulted in severe economic and ecological consequences, particularly in regions unprepared for its arrival. Understanding the pest’s biology, behavior, and strain variations is crucial for anticipating its movements and potential impact. Continued research and international collaboration are vital to deepen scientific knowledge and enhance preparedness against such invasive threats, ensuring the resilience of cropping systems in the face of ongoing biological invasions.

**References**

1. Amarasinghe, U.A. and Singh, O.P. 2008. Changing consumption patterns of India: implications on future food demand. In: India’s Water Future: Scenarios and Issues (eds. Amarasinghe, U.A., Shah, T. and Malik, R. P. S.). International Water Management Institute, Colombo, Sri Lanka, pp. 131–146.
2. Assefa, F., Ayalew, D. (2019). Status and control measures of fall armyworm (Spodoptera frugiperda) infestationsin maize fields in Ethiopia: A review. Cogent Food & Agriculture, 5(1), 1641902.
3. Bajracharya, A.R., Bhat, B. (2019). The first record of Fall Armyworm Spodoptera frugiperda in Nepal. Khumaltar,Nepal, 2019 NARC <http://narc.gov.np/the-first-record-of-fall-armyworm-spodoptera-frugiperdain-nepal/>
4. Baudron, F., Zaman-Allah, M.A., Chaipa, I., Chari, N., Chinwada, P. (2019). Understanding the factors influencingfall armyworm (Spodoptera frugiperda JE Smith) damage in African smallholder maize fields andquantifying its impact on yield. A case study in Eastern Zimbabwe. Crop Protection, 120, 141-150
5. CABI. (2020). Spodoptera frugiperda (fall armyworm). Invasive Species Compendium. Retrieved from: <https://www.cabi.org/isc/datasheet/29810>
6. Chimweta, M., Nyakudya, I.W., Jimu, L., Bray Mashingaidze, A. (2019). Fall armyworm [Spodoptera frugiperda(JE Smith)] damage in maize: management options for flood-recession cropping smallholder farmers.International Journal of Pest Management, 1-13.
7. Dass, S. 2013. Maize and its diversified uses, In: Maize production systems for improving resource-use efficiency and livelihood security, Eds., Kumar, Ashok, Jat, S. L., Kumar, Ramesh, Yadav, O.P., Directorate of Maize Research, pp. 1-3.
8. Day, R., Abrahams, P., Bateman, M., Beale, T., Clottey, V., Cock, M., Gomez, J. (2017). Fall armyworm: impactsand implications for Africa. Outlooks on Pest Management, 28(5), 196-201.
9. Kansiime, M. K., Mugambi, I., Rwomushana, I., Nunda, W., Lamontagne‐Godwin, J., Rware, H., Day, R. (2019).Farmer perception of fall armyworm (Spodoptera frugiderda JE Smith) and farm‐level management practicesin Zambia. Pest management science, 75(10), 2840-2850.
10. Maitra, Sagar & Shankar, Tanmoy & Manasa, Pilli & Sairam, Masina. (2019). Present Status and Future Prospects of Maize Cultivation in South Odisha.
11. Maletta HE. From Hunger to Food Security: A Conceptual History (2024 revision). Available from: SSRN 2484166.
12. Nagoshi, R.N., Silvie, P., Meagher, R.L., Lopez, J., Machado, V. (2007). Identification and comparison of fallarmyworm (Lepidoptera: Noctuidae) host strains in Brazil, Texas, and Florida. Annals of the EntomologicalSociety of America, 100(3), 394-402.
13. Pokhrel, S., Khadka, G. B., Sherpa, D. D., Sah, N., Gautam, I., Upadhyaya, S. D., & Khanal, R. (2022). Invasion of Fall armyworm [Spodoptera frugiperda (JE Smith, 1797)(Lepidoptera: Noctuidae)](JE Smith, 1797)(Lepidoptera: Noctuidanead) Management Strategies in Maize Fields of Nepal. Turkish Journal of Agriculture-Food Science and Technology, 10(4), 629-636.
14. Prasanna, B. M., Huesing, J. E., Eddy, R., & Peschke, V. M. (2018). Fall armyworm in Africa: a guide for integrated pest management.
15. Prasanna, B., Huesing, J.E., Eddy, R., Peschke, V.M., 2018. Fall Armyworm in Africa: a Guide for Integrated Pest Management. Mexico, CDMX: CIMMYT., First Edit, Pp. 45–62. [www.maize.org](http://www.maize.org).
16. Prasanna, B.M., Huesing, J.E., Eddy, R., Peschke, V.M. (2018). Fall armyworm in Africa: a guide for integrated pestmanagement.
17. Ramirez-cabral, N. Y. Z., Kumar, L., Shabani, F. (2017). Future climate scenarios project a decrease in the risk offall armyworm outbreaks. The Journal of Agricultural Science, 155(8), 1219-1238
18. Reddy YVR, Trivedi S. Maize Production Technology. Academic Press. 2008, 0-192.
19. Reddy, J. (2019). Fall Armyworm control methods and symptoms. Agrifarming. Retrived from <https://www.agrifarming.in/fall-armyworm-control-methods-and-symptoms>
20. Repalle Naganna, Jethva DM, Bhut JB, Pankaj S Wadaskar, Akash Kachot. Present status of new invasive pest fall armyworm, Spodoptera frugiperda in India: A review. J Entomol Zool Stud 2020;8(2):150-156.
21. Sagar, G.C., Aastha, B., Laxman, K., 2020. An introduction of fall armyworm (Spodoptera frugiperda) with management strategies: a review paper. Nippon Journal of Environmental Science, 1 (4), Pp. 1010. <https://doi.org/10.46266/njes.1010>
22. Sambit Mishra, RK Rout, BP Gantayat, LK Das, T Badjena, Chandana Behera, RL Moharana. Exploring trends and constraints of maize production in Odisha. Int J Res Agron 2025;8(1S):292-296. DOI: [10.33545/2618060X.2025.v8.i1Se.2387](https://doi.org/10.33545/2618060X.2025.v8.i1Se.2387)
23. Sangomla, A., Kukreti, I. (2019).Fall Armyworm attack: The damage done. <https://www.downtoearth.org.in/coverage/agriculture/fall-armyworm-attack-the-damage-done-63445>
24. Sisay, B. (2018). Evaluation of different management options of fall armyworm,(JE Smith)(Lepidoptera: Noctuidae) and assessment of its parasitoids in some parts of Ethiopia (Doctoral dissertation, Haramaya University).
25. Sisay, B., Simiyu, J., Malusi, P., Likhayo, P., Mendesil, E., Elibariki, N., Tefera, T. (2018). First report of the fallarmyworm, Spodoptera frugiperda (Lepidoptera: Noctuidae), natural enemies from Africa. Journal ofApplied Entomology, 142(8), 800-804.
26. Sisay, B., Tefera, T., Wakgari, M., Ayalew, G., Mendesil, E. (2019). The efficacy of selected synthetic insecticidesand botanicals against fall armyworm, Spodoptera frugiperda, in maize. Insects, 10(2), 45.
27. Westbrook, J.K., Sparks, A.N. (1986). The role of atmospheric transport in the economic fall armyworm(Lepidoptera: Noctuidae) infestations in the southeastern United States in 1977. Florida Entomologist, 492-502.