*Original Research Article*

PCA-driven Insights into Hybrid-Parent Performance in Okra [*Abelmoschus esculentus* (L.) Moench.]

.

ABSTRACT

|  |
| --- |
| **Aims**: The study aimed to evaluate genetic diversity and identify trait relationships among okra (*Abelmoschus esculentus* L. Moench) F1 hybrids and their parental lines using principal component analysis (PCA). It sought to uncover key traits contributing to yield variability and to support breeding efforts by identifying superior hybrids and combining parents suited to Sudanese agro-ecologies.  **Study Design:** The study used a line × tester mating design involving 10 parental lines and their 21 resulting F1 hybrids. Multivariate statistical tools, including PCA and hierarchical clustering, were employed to assess trait variability, relationships, and genotype grouping based on multiple morphological and yield-related parameters.  **Place of Study:** The research was conducted at the demonstration farm of the Faculty of Agriculture, University of Al Zaeim Al Azhari, located in Khartoum North, Sudan.  **Methodology:** Eleven agronomic traits were recorded, including plant height, number of fruits per plant, yield per plant, and others. Data were analyzed using analysis of variance and principal component analysis. The first four principal components were extracted, and their eigenvalues, trait loadings, and correlations were studied. Standardized Euclidean Distance and Ward’s method were used for hierarchical clustering to group genotypes based on similarity.  **Results & Discussion**: Substantial phenotypic variation was observed, with yield per plant (CV = 21.2%), fruit fresh weight (18.9%), and number of lateral branches (16.3%) showing the highest variability. PCA revealed four principal components explaining 71.85% of the total variance. PC1 (27.48%) was highly associated with yield traits, while PC2 (22.14%) captured architectural and earliness traits. PC3 and PC4 explained structural variation. Cluster analysis grouped genotypes into four major clusters. Sinnar-derived hybrids formed a compact, high-yielding group, indicating superior combining ability. Clemson crosses exhibited wide variability and heterotic potential, while Hjerat lines showed genetic divergence useful for broadening the gene pool.  **Conclusion:** Principal Component Analysis effectively revealed genetic diversity and critical trait associations among okra hybrids and parents. Sinnar was identified as a superior parent for yield improvement, while other testers contributed unique traits. The results support the use of PCA in trait prioritization, selection of promising genotypes, and development of improved okra cultivars adaptable to Sudanese and similar environments. |

*Keywords: Abelmoschus esculentus, principal component analysis, hybrid performance, yield traits, parental line selection, okra breeding*

**1. INTRODUCTION**

Okra [*Abelmoschus esculentus* (L.) Moench], a warm-season vegetable crop of tropical and subtropical origin, is widely grown across Asia, Africa, and the Middle East for its edible immature pods, nutritional value, and medicinal uses. Rich in vitamins A, C, and K, as well as calcium, magnesium, and mucilage, okra contributes significantly to food and nutritional security, especially in low-income communities (Dhankhar *et al.,* 2022). Globally, okra is cultivated on over two million hectares, with India, Nigeria, and Sudan among the top producers (FAOSTAT, 2022). In Sudan, okra is one of the most popular traditional vegetables, cultivated under both irrigated and rain fed conditions and consumed in fresh and dried forms. Despite its importance, yield levels remain low due to limited genetic improvement, poor seed systems, and a lack of systematic breeding strategies. Most farmers grow local landraces that are genetically diverse but largely uncharacterized, with little data available on hybrid performance or parent-hybrid relationships.

To address these challenges, hybridization programs have been initiated to develop improved cultivars with better yield, quality, and adaptability. However, the selection of promising hybrids and parents requires a deeper understanding of the genetic architecture of yield and morphological traits. Most of these traits are quantitatively inherited, complex, and highly influenced by environmental factors, making direct phenotypic selection difficult. In this context, Principal Component Analysis (PCA) offers a powerful multivariate statistical approach to unravel the underlying structure of trait variability, reduce data dimensionality, and reveal hidden patterns of trait association. PCA transforms correlated traits into a set of orthogonal principal components that capture the maximum variance, thereby facilitating genotype classification and trait prioritization .Its application in crop breeding has proven effective in identifying trait combinations that explain most of the variability, simplifying the selection process and supporting genetic improvement.

In okra breeding, PCA can also be used to explore the clustering of hybrids about one or both of their parents, providing insights into genetic relationships, heterosis, and transgressive segregation. However, despite its utility, the application of PCA in okra breeding remains limited in Sudan, especially in studies comparing hybrids with their parental lines. This gap restricts breeders' ability to exploit the full potential of multivariate analysis for selection and cultivar development. Therefore, this study was conducted to analyze the morphological and yield-related variation in okra hybrids and their parents using Principal Component Analysis to provide insight into genetic diversity, determine the relative contribution of each trait to total variation, examine the clustering patterns of hybrids near one or both parents and identify superior hybrids based on multivariate trait performance. The findings are expected to inform selection strategies and support ongoing efforts to enhance okra breeding efficiency and genetic gain in Sudan and similar agro-ecological regions.

2. material and methods

The present investigation was conducted at the demonstration farm of the Faculty of Agriculture at the University of Al Zaeim Al Azhari, located in Khartoum North. A total of ten genetically diverse parental lines of okra were crossed using the line × tester method developed by Kempthorne in 1957. Seven lines, namely HSD 1835, HSD1834, HSD1839, HSD 2543, HSD 2482, HSD 1840, and HSD 2550, were used as female lines, while Sinnar, Hjerat, and Clemson Spineless served as male lines/testers.

The crossing was performed in standard crossing blocks, with emasculation and pollination conducted before flower opening. Standard procedures for emasculation and pollination were adhered to, utilizing a cutter, forceps, tags, and paper bags. In total, twenty-one F1 hybrids, along with the ten parental lines, were arranged in a Complete Randomized Block Design (CRBD) with three replications. Each plot consisted of three ridges, extending three meters. The spacing adopted was 75 cm between ridges and 30 cm between plants.

Observations on the number of days to flowering were recorded on a plot basis, noting the time from the sowing date to the onset of flowering. Additionally, plant height (cm), fruit length (cm), number of nodes per plant, number of lateral branches, number of fruits per plant, fruit dry weight (g), fruit fresh weight (g), number of seeds per fruit, and yield per plant (g) were recorded as averages from ten randomly selected plants in each plot across the three replicates. The weight of the hundred-seed count (g) was measured using a sensitive balance.

Data were analyzed using Analysis of Variance (ANOVA) as described by Gomez and Gomez (1984) through the online software package Grapes 1.1.0, developed by Gopinath *et al.* (2021) at Kerala Agricultural University. Additionally, Principal Component Analysis (PCA) was performed to assess trait divergence, variability, and genotype clustering, as outlined by Pearson (1901) and Hotelling (1933). Trait-based clustering among genotypes was examined by Ward (1963), a hierarchical clustering method. The differences in scales or units across variables (traits) were adjusted using the Standardized Euclidean Distance formula, suggested by Gower (1966) and Kaufman & Rousseeuw (1990).

3. results and discussion

**3.1 Trait variability**

Table 1 presents the descriptive statistics for 11 morphological and yield-related traits among okra genotypes. The results indicate considerable phenotypic variation, which is essential for effective selection in breeding programs. For instance, plant height ranged from 23.2 cm to 45.6 cm (mean = 31.15 cm), and yield per plant ranged from 12.1 g to 27.0 g (mean = 20.29 g). The coefficient of variation (CV %) was notably high for yield per plant (21.2%), fruit fresh weight (18.9%), and number of lateral branches (16.3%), suggesting these traits are more influenced by genetic diversity and thus valuable for selection. In contrast, days to flowering exhibited a low CV (2.8%), indicating this trait was relatively stable across genotypes.

**Table 1. Descriptive statistics of morphological and yield traits in okra hybrids and their parents**

|  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Variance** | **Number of days to flowering** | **Number of lateral branches** | **Plant height (cm)** | **Number of nodes per plant** | **Fruit length (cm)** | **Number of fruits per plant** | **Fruit dry weight(g)** | **Fruit fresh weight(g)** | **Number of seeds per fruit** | **Weight of hundred-seeds(g)** | **Yield per plant (g)** |
| **Minimum** | 41.700 | 4.300 | 23.200 | 6.700 | 3.600 | 3.000 | 0.400 | 3.900 | 35.300 | 3.900 | 12.100 |
| **Maximum** | 48.000 | 8.300 | 45.600 | 11.000 | 5.800 | 6.000 | 0.600 | 5.500 | 64.700 | 6.300 | 27.000 |
| **Mean** | 45.603 | 5.958 | 31.145 | 8.677 | 4.713 | 4.539 | 0.497 | 4.848 | 49.339 | 5.116 | 20.294 |
| **Standard error mean** | 0.230 | 0.174 | 0.947 | 0.229 | 0.106 | 0.154 | 0.011 | 0.061 | 1.273 | 0.104 | 0.774 |
| **Standard deviation** | 1.278 | 0.971 | 5.270 | 1.274 | 0.593 | 0.859 | 0.060 | 0.342 | 7.087 | 0.580 | 4.308 |
| **Coefficient of variation %** | 2.8 | 16.3 | 16.9 | 14.7 | 12.6 | 18.9 | 12.2 | 7.1 | 14.4 | 11.3 | 21.2 |

**3.2 Eigenvalue structure and total variation**

Table 2 summarizes the eigenvalues and variance explained by each principal component. The first four principal components (PC1 to PC4) had eigenvalues >1 and jointly accounted for 71.85% of the total variation. The first principle component PC1 (27.48%) was the most influential, followed by PC2 (22.14%), PC3 (11.83%), and PC4 (10.39%). This pattern indicates that the first four components effectively summarize the multidimensional data and can be used to classify genotypes and interpret trait interrelationships. The drop in eigenvalues after PC4 supports retaining these components for further biological interpretation, consistent with the report by Idehen & Ola, (2021) on the Kaiser criterion (Kaiser, 1960).

Across various okra studies, PC1 alone explained a significant portion of the total variance. The first principle component (PC1) values reported ranged from (25.38%- 42.50%) was recorded in several works (Das *et al.*, 2022; Mohammed *et al.*, 2022; Kenaw *et al.*, 2023; Abhilash *et al.*, 2023). This confirms and is in lieu of the result obtained in this study.

Several researchers have reported that the first four principal components (PCs) effectively summarize multidimensional data, classify genotypes, and interpret trait interrelationships in okra (Dos Santos *et al.*, 2013; Das *et al.*, 2022; Idehen & Ola, 2021; Murtadha *et al.*, 2023; Abhilash *et al.*, 2023; Kenaw *et al.*, 2023; Syfullah *et al.*, 2025). The first four PCs frequently account for over 60% and sometimes up to 85% of the total variation (Murtadha *et al.*, 2023; Mohammed *et al.*, 2022; Kenaw *et al.*, 2023; Nwangburuka *et al.*, 2011). In Nigeria, Olayiwola *et al.*(2021) found the first four PCs accounted for 80% of the total variation, with PC1 having the highest discriminatory power. Another report from India by Ranga, *et al.*(2021) recorded as high as 84.28% cumulative variance across the first four PCs.

**Table 2. Eigenvalues and variance explained by principal components for morphological and yield traits in okra**

|  |  |  |  |
| --- | --- | --- | --- |
| **Principle components** | **Eigen value** | **percentage of variance** | **Cumulative variance percentage** |
| **PC1** | 3.023 | 27.48 | 27.48 |
| **PC2** | 2.436 | 22.142 | 49.622 |
| **PC3** | 1.301 | 11.83 | 61.452 |
| **PC4** | 1.143 | 10.394 | 71.846 |
| **PC5** | 0.728 | 6.615 | 78.461 |
| **PC6** | 0.689 | 6.266 | 84.727 |
| **PC7** | 0.607 | 5.52 | 90.247 |
| **PC8** | 0.379 | 3.446 | 93.694 |
| **PC9** | 0.361 | 3.284 | 96.978 |
| **PC10** | 0.24 | 2.186 | 99.164 |
| **PC11** | 0.092 | 0.836 | 100 |

**3.3 Contribution of traits to principal components**

Table 3 details the percentage contribution of each trait to the first eight PCs. Yield per plant followed by number of fruits per plant (22.31%), had the highest contribution to PC1 (25.75%), confirming its role as a primary determinant of genetic divergence among the studied genotypes. The specific trait identified as the highest contributor to PC1 varies from one study to another depending on the specific germplasm, traits measured, and environmental conditions of the study. However, yield-related traits such as "yield per plant," "pod yield," or "fruit yield per hectare" are consistently highlighted as major, and often the highest, contributors to PC1 in many investigations (Badiger *et al.*, 2018; Ranga, *et al.*, 2021; Idehen & Ola, 2021; Mohammed *et al.*, 2022; Kenaw *et al.*, 2023; Abhilash *et al.*, 2023 ). Other major contributors to PC1 included the number of seeds per fruit (19.07%), and fruit dry weight (14.92%). This indicates that PC1 largely represents yield performance and associated reproductive traits. Consensus from the sources indicates that yield performance and associated reproductive traits (such as number of fruits/pods per plant, fruit/pod yield, fruit/pod length, fruit/pod weight, and seed weight) are generally major, and often the most influential, contributors to the first principal component in genetic diversity analyses of okra. This makes PC1 a crucial indicator for identifying superior genotypes for breeding programs aimed at improving yield (Ranga *et al.*, 2022; Yadav *et al.*, 2024). PC2 was dominated by days to flowering (25.01%) and the number of lateral branches (23.87%), suggesting that it represents a dimension of plant architecture and earliness. Several studies indicate that PC2 is significantly influenced by, or largely loaded with a combination of plant architectural traits (e.g., plant height, branching) and earliness traits (e.g., days to flowering, days to maturity), often alongside important reproductive or yield components (Idehen & Ola, 2021; Ranga, *et al.*,2021; Mohammed *et al.*, 2022 ).The third principle component (PC3) captured variation in plant height (27.17%) and fruit fresh weight (37.14%), while PC4 was mainly explained by fruit length (34.98%) and number of nodes per plant (25.87%).

**Table 3. Percentage contributions of variables on principal components (PC1–PC8) in okra**

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Variables** | **PC1** | **PC2** | **PC3** | **PC4** | **PC5** | **PC6** | **PC7** | **PC8** |
| **Number of days to flowering** | 0.923 | 25.009 | 1.697 | 2.647 | 0.048 | 16.644 | 2.155 | 0.011 |
| **Number of lateral branches** | 0.496 | 23.875 | 5.416 | 7.331 | 5.275 | 4.296 | 1.815 | 18.74 |
| **Plant height (cm)** | 0.044 | 12.308 | 27.173 | 15.277 | 0.342 | 6.156 | 4.247 | 7.805 |
| **Number of nodes per plant** | 0.758 | 17.802 | 0.352 | 25.869 | 18.454 | 2.614 | 0.121 | 10.892 |
| **Fruit length (cm)** | 2.142 | 9.819 | 1.701 | 34.975 | 0.095 | 26.181 | 7.075 | 4.083 |
| **Number of fruits per plant** | 22.314 | 0.564 | 0.348 | 4.334 | 6.8 | 0.163 | 18.071 | 16.002 |
| **Fruit dry weight(g)** | 14.924 | 2.285 | 7.276 | 0 | 15.317 | 12.083 | 17.544 | 18.246 |
| **Fruit fresh weight(g)** | 3.652 | 1.154 | 37.141 | 5.742 | 33.185 | 3.164 | 3.612 | 0.517 |
| **Number of seeds per fruit** | 19.072 | 4.1 | 0.052 | 2.58 | 0.147 | 21.28 | 1.277 | 9.424 |
| **Weight of hundred-seeds(g)** | 9.927 | 1.628 | 18.137 | 1.17 | 18.558 | 2.838 | 33.566 | 13.572 |
| **Yield per plant (g)** | 25.749 | 1.455 | 0.708 | 0.076 | 1.779 | 4.579 | 10.517 | 0.707 |

**3.4 Trait–Principal component correlations**

Table 4 provides the correlation coefficients between each trait and the first eight PCs. High correlations between a trait and a PC indicate the trait’s influence on that component. Yield per plant showed a strong positive correlation with PC1 (r = 0.882), followed by number of fruits per plant (r = 0.821) and number of seeds per fruit (r = 0.759), reinforcing their contribution to PC1 variation and their relevance for yield improvement.

Days to flowering were strongly negatively correlated with PC2 (r = -0.780), reflecting variation in earliness among genotypes. This aligns with the findings of Abhilash *et al.*(2023), which demonstrate strong positive correlations between the number of fruits per plant and fruit yield per plant with Principal Component 1 (PC1). Additionally, recorded strong positive correlations for earliness with Principal Component 2 (PC2), which includes days to first flowering, days to 50% flowering, and days to the first fruit harvest.

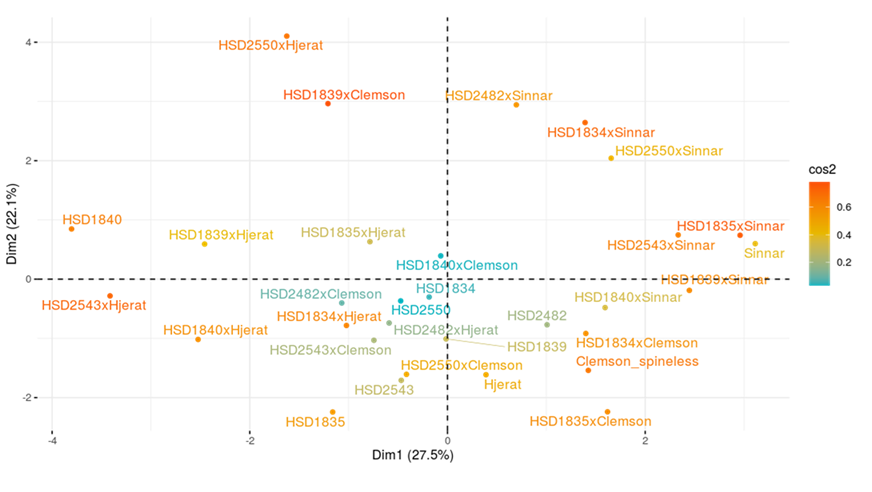
Plant height and fruit fresh weight had substantial contributions to PC3, with respective correlations of −0.595 and 0.695 respectively. Fruit length was negatively correlated with PC4 (r = -0.632), and nodes per plant had a positive association (r = 0.544), defining structural variation in this axis. These patterns affirm that PCA not only reduces data complexity but also identifies trait combinations that most influence genotype classification, which is critical in early-generation selection.

**Table 4. Trait–Principal component correlation matrix (PC1–PC8) for okra genotypes**

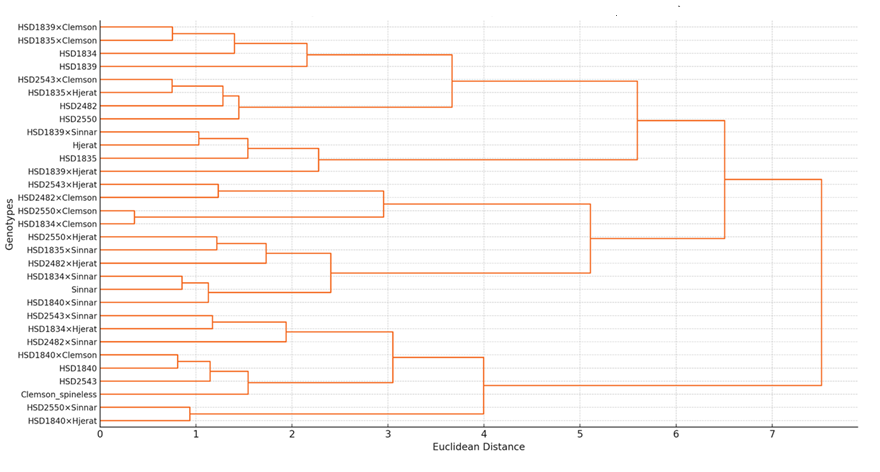
|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Variables** | **PC1** | **PC2** | **PC3** | **PC4** | **PC5** | **PC6** | **PC7** | **PC8** |
| **Number of days to flowering** | 0.167 | -0.78 | -0.149 | 0.174 | 0.019 | 0.339 | -0.114 | 0.006 |
| **Number of lateral branches** | -0.122 | 0.763 | 0.265 | 0.289 | -0.196 | -0.172 | 0.105 | 0.267 |
| **Plant height (cm)** | 0.036 | 0.548 | -0.595 | -0.418 | 0.05 | -0.206 | -0.161 | -0.172 |
| **Number of nodes per plant** | -0.151 | 0.658 | 0.068 | 0.544 | 0.366 | 0.134 | 0.027 | -0.203 |
| **Fruit length (cm)** | 0.254 | 0.489 | 0.149 | -0.632 | -0.026 | 0.425 | 0.207 | 0.124 |
| **Number of fruits per plant** | 0.821 | 0.117 | -0.067 | 0.223 | -0.222 | 0.034 | -0.331 | 0.246 |
| **Fruit dry weight(g)** | 0.672 | -0.236 | 0.308 | -0.001 | -0.334 | -0.289 | 0.326 | -0.263 |
| **Fruit fresh weight(g)** | 0.332 | -0.168 | 0.695 | -0.256 | 0.491 | -0.148 | -0.148 | 0.044 |
| **Number of seeds per fruit** | 0.759 | 0.316 | 0.026 | 0.172 | -0.033 | 0.383 | 0.088 | -0.189 |
| **Weight of hundred-seeds(g)** | 0.548 | -0.199 | -0.486 | 0.116 | 0.367 | -0.14 | 0.451 | 0.227 |
| **Yield per plant (g)** | 0.882 | 0.188 | -0.096 | -0.029 | 0.114 | -0.178 | -0.253 | -0.052 |

**3.5 Cluster and bi-plot analysis of okra hybrids and their parents**

Figure 1 shows the Bi-Plot (PC1-PC2) for Okra Hybrids and Parental Lines, while Figure 2 displays the cluster dendrogram of Okra Genotypes using PCA scores. Both figures illustrate the clustering pattern of okra hybrids and their parental lines identifying four clustering groups.  Group A includes the High-Yielding Sinnar Hybrids, HSD2482×Sinnar, HSD1834×Sinnar, HSD1835×Sinnar HSD2550×Sinnar, and Sinnar. These hybrids form a tight cluster, indicating Sinnar's strong combining ability for yield traits. Group B comprises the Clemson-Derived Diverse Hybrids, including HSD1839×Clemson, HSD1835×Clemson, and HSD1834×Clemson. These genotypes are widely separated, reflecting higher variability and heterotic expression. Group C include the divergent Hjerat Lines and Late Maturing Genotypes, which Include HSD1839×Hjerat, HSD2543×Hjerat, HSD1835, and HSD1840. These are genetically divergent, showing unique trait profiles possibly suited for genetic base broadening. Group D represent the core cluster of stable lines that Includes HSD1839, HSD2482, HSD2550, and Hjerat. These genotypes display moderate trait values and genetic stability. The dendrogram confirms PCA findings and highlights Sinnar-derived hybrids for yield improvement. Hjerat and Clemson contribute to broader trait diversity, suggesting their value in heterotic group development. The core genotypes may be used as standard checks or donor parents. The results align with previous work on these hybrids reporting significant heterosis for yield and yield components for these hybrid combinations HSD2482×Sinnar, HSD1834×Sinnar, HSD1835×Sinnar HSD2550×Sinnar, (Abdalla & ElKamil, 2025a). The role of sinner as a parent has also been highlighted and recommended as a parent in several studies (Abdalla & AlKamal, 2025b).

****

**Figure 1. Principal component analysis bi-plot (PC1- PC2) of okra hybrids and parental lines**

****

**Figure 2. Cluster dendrogram of okra genotypes using PCA scores**

4. Conclusion

The Principal Component Analysis (PCA) of morphological and yield traits in okra hybrids and their parental lines revealed significant genetic diversity and trait relationships critical for breeding programs. The first four principal components (PC1–PC4) cumulatively explained 71.85% of the total variation, with PC1 (27.48%) primarily associated with yield-related traits such as yield per plant, number of fruits per plant, and seeds per fruit, highlighting their importance for selection. PC2 (22.14%) was linked to earliness (days to flowering) and plant architecture (lateral branches), while PC3 and PC4 captured structural traits like plant height, fruit length, and node number. Hierarchical clustering and biplot analysis grouped the genotypes into four distinct clusters. The high-yielding Sinnar hybrids (HSD2482×Sinnar, HSD1834×Sinnar, HSD1835×Sinnar HSD2550×Sinnar). These findings confirm Sinnar as a superior parent for yield enhancement. This research supports the development of improved cultivars for enhanced productivity and food security. Future work should integrate genomic data to validate clustering patterns and expand trait evaluations across diverse environments to further refine breeding strategies.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

The author(s) hereby declare that generative AI technologies and digital tools, including Grammarly Version 1.2.169.1689, released June 19, 2025 and Zotero 7.0.19, were used solely for reference management, proofreading, and language editing. All content and ideas presented in the manuscript were entirely generated by the author(s), without AI assistance in writing or data interpretation.

References

Dhankhar, P., Yadav, R., Kaur, P., & Arora, A. (2022). Nutritional and medicinal significance of okra (Abelmoschus esculentus): A review. Journal of Food Science and Technology, 59(1), 1–12. <https://doi.org/10.1007/s13197-021-05123-5>

FAOSTAT. (2022). Food and Agriculture Organization of the United Nations. Retrieved from <https://www.fao.org/faostat/en>

Gomez, K.A., & Gomez, A.A. (1984). Statistical Procedures for Agricultural Research (2nd ed.). John Wiley & Sons.

Gopinath, P. P., Parsad, R., Joseph, B., & VS, A. (2021). grapesAgri1: a collection of shiny apps for data analysis in agriculture. *Journal of Open Source Software*, *6*(63), 3437.

Pearson, K. (1901). On lines and planes of closest fit to systems of points in space. *Philosophical Magazine*, 2(11), 559–572.

Hotelling, H. (1933). Analysis of a complex of statistical variables into principal components. *Journal of Educational Psychology*, 24(6), 417–441.

Ward, J. H. (1963). Hierarchical Grouping to Optimize an Objective Function. Journal of the American Statistical Association, 58(301), 236–244. <https://doi.org/10.1080/01621459> .1963.10500845

Gower, J. C. (1966). Some distance properties of latent root and vector methods used in multivariate analysis. *Biometrika, 53*(3/4), 325–338. <https://doi.org/10.1093/biomet/53> .3-4.325.

Kaufman, L., & Rousseeuw, P. J. (1990). *Finding groups in data: An introduction to cluster analysis*. Wiley, <https://doi.org/10.1002/9780470316801>.

Idehen, E. O., & Ola, O. E. (2021). Performance and character contributions to variability in okra (*Abelmoschus esculentus* L. Moench) genotypes. *Acta Horticulturae et Regiotecturae*, *24*(2), 127–133. <https://doi.org/10.2478/ahr-2021-0034>.

Kaiser, H. F. (1960). The application of electronic computers to factor analysis. *Educational and Psychological Measurement*, 20(1), 141–151. https://doi.org/10.1177/001316446002000116

Das, A., Yadav, R. K., Choudhary, H., Lata, S., Singh, S., Kumar, C., Kumari, S., Boopalakrishnan, G., Bhardwaj, R., & Talukdar, A. (2022). Population structure, gene flow and genetic diversity analyses based on agro-morphological traits and microsatellite markers within cultivated and wild germplasms of okra [*Abelmoschus esculentus* (L.) Moench.]. *Genetic Resources and Crop Evolution*, *69*(2), 771–791. <https://doi.org/10.1007/s10722-021-01263-9>.

Mohammed, J., Mohammed, W., & Shiferaw, E. (2022). Phenotypic Diversity Assessment of Okra (*Abelmoschus esculentus* (L.) Moench) Genotypes in Ethiopia Using Multivariate Analysis. *Scientifica*, *2022*, 1–9. <https://doi.org/10.1155/2022/3306793>

Kenaw, W., Mohammed, W., & Woldetsadik, K. (2023). Morpho-agronomic variability of okra [Abelmoschus esculentus (L.) Moench] genotypes in Dire Dawa, eastern Ethiopia. *Plos one*, *18*(7), <https://doi.org/10.1371/journal.pone.0288534>.

Abhilash, P. V., Talekar, N., Delvadiya, I. R., & Singh, S. K. (2023). Principal Component Analysis Approach for Yield Attributing Traits in Okra (*Abelmoschus esculentus* L.) Genotypes. *Journal of Advanced Zoology*, *44*(3), 958–964. <https://doi.org/10.17762/jaz.v44i3.1254>

Dos Santos, I. F., Dos Santos, A. M. P., Barbosa, U. A., Lima, J. S., Dos Santos, D. C., & Matos, G. D. (2013). Multivariate analysis of the mineral content of raw and cooked okra (*Abelmoschus esculentus* L.). *Microchemical Journal*, *110*, 439–443. [https://doi.org/10.1016/ j.microc.2013.05.008](https://doi.org/10.1016/%20j.microc.2013.05.008)

Murtadha, M. A., Adetoro, S., & Shittu, K. A. (2023). Multivariate analysis of seed yield and morphological characters of Okra (*Abelmoschus esculentus* L. Moench) accessions. *Agricultural Science and Technology*, *15*(2), 17–25. [https://doi.org/10.15547/ ast.2023.02.012](https://doi.org/10.15547/%20ast.2023.02.012).

Nwangburuka, C. C., Kehinde, O. B., Ojo, D. K., Denton, O. A., & Popoola, A. R. (2011). Morphological classification of genetic diversity in cultivated okra, *Abelmoschus esculentus* (L) Moench using principal component analysis (PCA) and single linkage cluster analysis (SLCA). *African Journal of Biotechnology*, *10*(54), 11165-11172. <https://doi.org/10.5897/ajb11.285>

Syfullah, K., Islam, M. S., Jahan, N., Haque, Md. I., Manik, Md. M. H., Afrose, S. N., Bashar, Md. A., Joha, M. R. K., & Sharmin, S. (2025). Exploring Genetic Diversity and Variation in Okra (*Abelmoschus esculentus* L. Moench): A Comprehensive Study on Cultivars. *Asian Journal of Research in Crop Science*, *10*(1), 37–43. <https://doi.org/10.9734/ajrcs/2025/v10i1330>.

Olayiwola, R., Yusuf, R. A., Oyetunde, O. A., Sosanya, O. S., & Ariyo, O. J. (2021). Assessment of genetic variability among accessions of okra (Abelmoschus esculentus L. Moench). Acta Horticulturae et Regiotecturae, 24(2), 141–147. <https://doi.org/10.2478/ahr-2021-0036>.

Ranga, A. D., Kumar, S., & Darvhankar, M. S. (2021). Variability among different yield and yield contributing traits of Okra (*Abelmoschus esculentus* L. Moench) genotypes. *Electronic Journal of Plant Breeding*, *12*(1), 74-81.[https://doi.org/10.37992/2021. 1201.011](https://doi.org/10.37992/2021.%201201.011)

Badiger, M., Rao, P. G., Pitchaimuthu, M., Indiresh, K. M., & Lingaiah, H. B. (2018). Multivariate assessment of yield and its components in okra [Abelmoschus esculentus (L.) moench] genotypes. Agricultural Research Journal, 55(1), 140. https://doi.org/10.5958/2395-146x.2018.00023.6

Yadav, K., S.K. Dhankhar, Davinder Singh, Uday Singh, & Amit. (2024). Genetic variability, character association and path coefficient analysis in okra. *Indian Journal of Horticulture*, *81*(03), 231–237. <https://doi.org/10.58993/ijh/2024.81.3.2>

Abdalla, A.I. & ElKamil, Y.A. (2025a). ASSESSING HETEROSIS FOR YIELD AND YIELD COMPONENTS IN OKRA (*Abelmoschus esculentus*). *International Journal of Agriculture and Environmental Research*, 11(02), 324–336.[https://doi.org/10.51193/ ijaer.2025.11202](https://doi.org/10.51193/%20ijaer.2025.11202)

Abdalla, A. I., & AlKamal, Y. A. (2025b). Combining Ability Analysis for Earliness and Growth Parameters in Sudanese Okra Collection [Abelmoschus esculentus (L.) Moench]. *Journal of Experimental Agriculture Internationa*l, 47(3), 100–111. <https://doi.org/10.9734/jeai/> 2025/v47i33320