**Genetic Divergence and Principal Component Analysis for Yield and Yield Components of Lentil (*Lens culinaris L. Medik*.) Genotypes**

**Abstract**  
**Aims:**  
The present study aimed to assess the genetic diversity among 25 lentil (*Lens culinaris L*. *Medikus*) genotypes, including three standard checks, with a focus on identifying high-yielding genotypes and those with elevated iron and zinc content, to support future breeding programs.

**Study Design:** The experiment was laid out in a randomized block design (RBD) with three replications.

**Place and Duration of Study:** The study was conducted at the Research Farm of Banda University of Agriculture and Technology, Banda, Uttar Pradesh, India, during the rabi season of 2023–2024.

**Methodology:** Analysis of variance (ANOVA) was performed to determine genotypic variation. Cluster analysis and principal component analysis (PCA) were used to study genetic diversity and relationships among the genotypes.

**Results:** Significant genotypic differences were observed for most traits, indicating ample genetic variability. Cluster analysis grouped genotypes into three distinct clusters, with Cluster II containing the most genotypes (15), followed by Clusters I and III (5 each). The highest inter-cluster distance was found between Clusters II and III, suggesting the potential for generating superior recombinants through inter-cluster hybridization. PCA revealed four principal components accounting for 72.30% of total variance. The highest seed yield was recorded in genotypes ILL-753/ILL-8461 (0.294 kg/plot), ILL-7537/ILL-800-S4 (0.280 kg/plot), and Black lentil (0.284 kg/plot). Genotypes IPL-316 (119.5 mg/kg) and PL-4 (106.3 mg/kg) had the highest iron content, while X20115-89-23-S4 (54.4 mg/kg) and ILL-10657 (52.1 mg/kg) had the highest zinc content.

**Conclusion:** The highest seed yield was recorded in genotypes ILL-753/ILL-8461 (0.294 kg/plot), ILL-7537/ILL-800-S4 (0.280 kg/plot), and Black Lentil (0.284 kg/plot). The highest iron content was observed in genotypes IPL-316 (119.5 mg/kg) and PL-4 (106.3 mg/kg), while the highest zinc content was found in genotypes X20115-89-23-S4 (54.4 mg/kg) and ILL-10657 (52.1 mg/kg)."

**Keywords:** Analysis of variance, Cluster analysis, Iron, Lentil, Principal components analysis (PCA), and Zinc

1. **Introduction**

Lentil (*Lens culinaris L. Medik*.), a diploid (2n = 2x = 14), self-pollinating, annual cool-season grain legume with a genome size of 4,063 Mbp, constitutes a major dietary staple worldwide (Faris et al., 2013). As one of the earliest domesticated crops in the Fertile Crescent, its cultivation extends back approximately 10,000 years (Cokkizgin & Shtaya, 2013). According to FAOSTAT (2023), global lentil production amounted to 5.610 million t from 5.585 million ha, of which India accounted for 1.490 million t over 1.734 million ha. When incorporated into dryland wheat (Triticum aestivum L.)‐based cropping systems, lentil have been shown to suppress grass weeds, interrupt disease cycles, improve water‐use efficiency, and increase overall yield (Chen et al., 2012; Miller et al., 2015), while also enhancing farm profitability (Tanaka et al., 2005). Iron (Fe) is an important part of hemoglobin, a protein that carries oxygen to body tissues. Globally, an estimated 50% of anemia cases are likely attributed to Fe deficiency (Stoltzfus, 2003), which makes the body more susceptible to other diseases (Hassan et al., 2018). About 33% of the world’s population suffers from zinc (Zn) deficiency, characterized by growth retardation and impaired immune function (Prasad, 2004). Lentil grains grown at multiple environments varied in total Fe and Zn concentrations among lentil genotypes, and with estimated Fe and Zn, respectively (Thavarajah et al., 2009). Significant cultivars and variations were observed in analyzing the micronutrient Fe and Zn content of lentil Genotype (Ray et al., 2014). A lentil genotype indicated significant heterogeneity in the grain Fe and Zn contents. Understanding the genetic basis of developmental and yield-related traits is vital for improving crop performance through breeding. When traits associated with yield are well-characterized, they can serve as reliable selection criteria (Poehlman, 1991; Singh et al., 1995). Research has shown that genotypic correlations often exceed their phenotypic counterparts, suggesting stronger genetic linkages that may be masked by environmental factors. For example, considerable genetic and phenotypic variability in seed yield per plant. In contrast, Bicer and Sarkar (2004) found low heritability for several traits, including flowering time, seed yield, biomass, maturity, seed weight, pod number, and plant height. Conversely, Gowda et al. (1997) documented higher heritability and genetic advance for seed yield, indicating a stronger potential for genetic improvement. Traits such as thousand-seed weight, and plant height have also shown positive and significant associations with seed yield in lentil (Ghimire *et al.* 2019). To optimize breeding efforts, it is essential to thoroughly evaluate the genetic diversity within lentil populations. This enables the effective selection of parent lines, facilitates the introduction of beneficial traits into elite varieties, and supports the long-term conservation of valuable genetic resources. Selecting genetically diverse parents for hybridization is particularly important, as such crosses increase the likelihood of producing superior offspring in future generations. Principal Component Analysis (PCA) and Cluster Analysis are essential multivariate statistical tools used in plant breeding and genetic research. PCA reduces data complexity by transforming correlated variables into a smaller number of uncorrelated components, allowing for the identification of key traits contributing to genetic variability. It enables effective ranking of genotypes based on their principal component scores. Cluster Analysis, on the other hand, groups genotypes into distinct clusters based on trait similarity, thus revealing genetic diversity and relationships among accessions. Together, these methods facilitate the selection of superior and genetically diverse genotypes, thereby improving the efficiency and accuracy of crop improvement programs.

1. **Materials and Methods**

The experiment was conducted to evaluate 25 lentil genotypes, including three check varieties, under irrigated conditions and normal soil using a randomized block design (RBD). Field trials were carried out during the *Rabi* season of 2023–2024 at the postgraduate research block of the College of Agriculture, Banda University of Agriculture and Technology, Banda, India. The experimental site is located at an elevation of 113 meters above mean sea level, between latitudes 24°53′ and 25°55′ North and longitudes 80°07′ and 81°34′ East. Each genotype was sown in a plot consisting of three rows, with each row measuring 2 meters in length and spaced 30 cm apart, covering a total plot area of 1.2 m². The study was conducted under controlled field conditions with an average temperature ranging from 20°C to 28°C and moderate rainfall, suitable for lentil growth. A recommended dose of fertilizers (20:40:20 kg/ha of N: P: K) was applied at sowing. The crop was managed following a standard package of practices for lentil cultivation, including timely sowing, optimum spacing, manual weeding, and irrigation schedules to ensure healthy crop growth and reliable experimental outcomes.

Data were recorded from five randomly selected plants per genotype for the traits under study. Statistical analyses were conducted using the R Studio software package R 4.3.0, including analysis of variance (ANOVA), cluster analysis, and principal component analysis (PCA), were performed to assess genetic variability and relationships among the genotypes. The estimation of iron and zinc content was carried out using the Agilent ICP-MS 7850 instrument, which is housed in the Central Laboratory Facility at Banda University of Agriculture and Technology (BUAT), Banda. This advanced inductively coupled plasma mass spectrometry (ICP-MS) system provides high sensitivity and precision for trace element analysis.

**Table 1. List of 25 genotypes and 3 checks with sources**

|  |  |  |
| --- | --- | --- |
| **S.No.** | **Genotype Name** | **Sources** |
| 1 | X20125-109-513 | ICARDA, AMLA, SEHORE, (MP) |
| 2 | IPL-406 | BUAT, Banda |
| 3 | IPL-316 | BUAT, Banda |
| 4 | 10072/1712/4-1 | ICARDA, AMLA, SEHORE, (MP) |
| 5 | PDL-1 | BUAT, Banda |
| 6 | 6002/7716/4-4-S2 | ICARDA, AMLA, SEHORE, (MP) |
| 8 | ILL-753/ILL-8461 | ICARDA, AMLA, SEHORE, (MP) |
| 9 | L-4076 | IARI, New Delhi |
| 10 | ILL-6778XILL-5480-S3-S4 | ICARDA, AMLA, SEHORE, (MP) |
| 11 | 6002/LiRL-21-50-1-1/17-S3-S1 | ICARDA, AMLA, SEHORE, (MP) |
| 12 | 6002/LiRL-21-50-1-1-1/24-6 | ICARDA, AMLA, SEHORE, (MP) |
| 13 | 590/8461/2-4-S1 | ICARDA, AMLA, SEHORE, (MP) |
| 14 | 4605/4380/2SPS | ICARDA, AMLA, SEHORE, (MP) |
| 15 | ILL-7537/ILL-800-S4 | ICARDA, AMLA, SEHORE, (MP) |
| 16 | X20125-146-S1 | ILL4605 x ILL6002 |
| 17 | X20125-171-17-S1 | ICARDA, AMLA, SEHORE, (MP) |
| 18 | X20115-89-23-S4 | ICARDA, AMLA, SEHORE, (MP) |
| 19 | L-4147 | IARI, New Delhi |
| 20 | ILL-10657 | BUAT, Banda |
| 21 | KM-2 | BUAT, Banda |
| 22 | PL-04 | BUAT, Banda |
| 23 | L-4729 | IARI, New Delhi |
| 24 | PSL-9 | BUAT, Banda |
| 25 | KM-1 | BUAT, Banda |
| 26 | IPL-220 | BUAT, Banda |
| 27 | Black lentil | BUAT, Banda |

1. **Results and Discussion**

**3.1. Analysis of variance**

Genetic diversity and effective selection approaches are fundamental to the success of any breeding program, as they enable the utilization of available genetic variability (23, 24). The analysis of variance (ANOVA) revealed statistically significant differences among the pigeon pea genotypes for nearly all the traits examined (Table 2), suggesting the presence of substantial inherent genetic variability within the population. This variability offers ample opportunity to identify genotypes with favorable traits for yield improvement, particularly when appropriate selection pressure is applied. These findings are consistent with previous reports by (Alemayo et al 2021 and Kumar et al. 2022). Significant genotypic differences were observed for key agronomic and nutritional traits, including days to 50% flowering (DF), number of primary branches (PB), number of secondary branches (SB), plant height (PH), number of pods per plant (PP), seeds per pod (SP), days to 50% maturity (DM), biological yield (BY), 1000-grain weight (TGW), iron content (Fe, mg/kg), zinc content (Zn, mg/kg), and seed yield (SY) (Ghimire *et al.*  2024).

**Table 2. An analysis of variance (ANOVA) was conducted for twelve morphological and yield-related traits across lentil genotypes.**

|  |  |  |  |
| --- | --- | --- | --- |
| **Source** | **Replication** | **Genotype** | **Error** |
| **df** | 2 | 24 | 48 |
| **Days to 50% flowering** | 10.253 | 23.27\* | 10.587 |
| **Primary branches per plant** | 0.2125 | 0.5476\*\* | 0.09677 |
| **Secondary branches per plant** | 3.5897 | 7.0756\*\*\* | 0.8514 |
| **Plant height (cm)** | 49.204 | 38.44\* | 9.217 |
| **Total no. of pods per plant** | 1089.5 | 29950.8\*\*\* | 333.1 |
| **Total no. of seed per pod** | 0.011111 | 0.042133\* | 0.013244 |
| **Days to maturity** | 18.8333 | 0.2133 | 1.13 |
| **Biological yield (kg)** | 0.03824 | 0.68931\*\*\* | 0.01451 |
| **1000 seed wt. (gm)** | 103.455 | 43.944\*\* | 7.312 |
| **Fe** | 774.52 | 262.25\* | 71.05 |
| **Zn** | 93.545 | 33.739\*\*\* | 0.61 |
| **Seed yield (gm/plot)** | 0.003072 | 0.150492\*\*\* | 0.001786 |

*\*Indicates significance at 5%; Days to 50% flowering (DF), Number of primary branches (PB), Number of secondary branches (SB), Plant height (PH), Number of pods per plant (PP), Seeds per pod (SP), Days to 50% maturity (DM), Biological yield (BY), 1000-grain weight (TGW), Iron content (Fe, mg/kg), Zinc content (Zn, mg/kg), and Seed yield (SY).*

**3.2. Distribution of genotypes intoclusters**

To evaluate the genetic divergence among 25 genotypes Mahalanobis' D² statistics were calculated for all possible genotype pairs. This analysis facilitated the assessment of genetic diversity within the population. Based on the results of cluster analysis, the genotypes were classified into three distinct and non-overlapping clusters. Among these, Cluster II contained the highest number of fifteen genotypes, followed by clusters I and III, each comprising five genotypes (Table 3). In a separate analysis, 25 lentil genotypes were grouped into three clusters using the dendrogram method (Figure 1). Genotypes within the same cluster exhibited greater genetic similarity, whereas those from different clusters showed broader genetic divergence. Therefore, selecting parents from separate clusters could be advantageous for breeding programs, as such crosses have a higher potential to produce genetically diverse and heterotic offspring.

**Table 3. The clustering pattern of twenty-five genotypes, including three check varieties into three clusters.**

|  |  |  |
| --- | --- | --- |
| **Cluster** | **Genotype number** | **Genotype Name** |
| **I** | 5 | X20125-109-513, 6002/LiRL-21-50-1-1/17-S3-S1, 590/8461/2-4-S1, ILL-7537/ILL-800-S4, X20115-89-23-S4 |
| **II** | 15 | IPL-406, IPL-316, PDL-1, ILL-753/ILL-8461, L-4076, ILL-6778XILL-5480-S3-S4, 4605/4380/2SPS, L-4147, ILL-10657, PL-04, L-4729, PSL-9, KM-1, IPL-220, Black lentil |
| **III** | 5 | 10072/1712/4-1, 6002/LiRL-21-50-1-1-1/24-6, X20125-146-S1, X20125-171-17-S1, 6002/7716/4-4-S2 |

* 1. **Inter-cluster and Intra-cluster distance**

The intra and inter-cluster distances between all possible pairs of three clusters were computed and presented in Table 4. While the minimum inter-cluster distance demonstrated a close link between the groupings, the largest inter-cluster distance indicated broad diversity. The maximum intra-cluster distance was found in Cluster III (4.580) followed by Cluster II (4.227), and the minimum intra-cluster distance was found in Cluster I (3.793). The maximum inter-cluster distance was found between cluster III (5.445), and the minimum inter-cluster distance was found between II (4.990). Similar results of utilization of principal component analysis combined with hierarchical cluster analysis in genetic diversity studies were reported by Rao, (Maurya *et al.* 2018) in lentil.

**Table 4. The average intra and inter-cluster distances for f clusters**

|  |  |  |  |
| --- | --- | --- | --- |
| **Cluster** | I | II | III |
| I | **3.793105** | 4.990322 | 5.088511 |
| II |  | **4.227174** | 5.445513 |
| III |  |  | **4.580984** |

*\*Bold values in the table represent intra-cluster distances*.

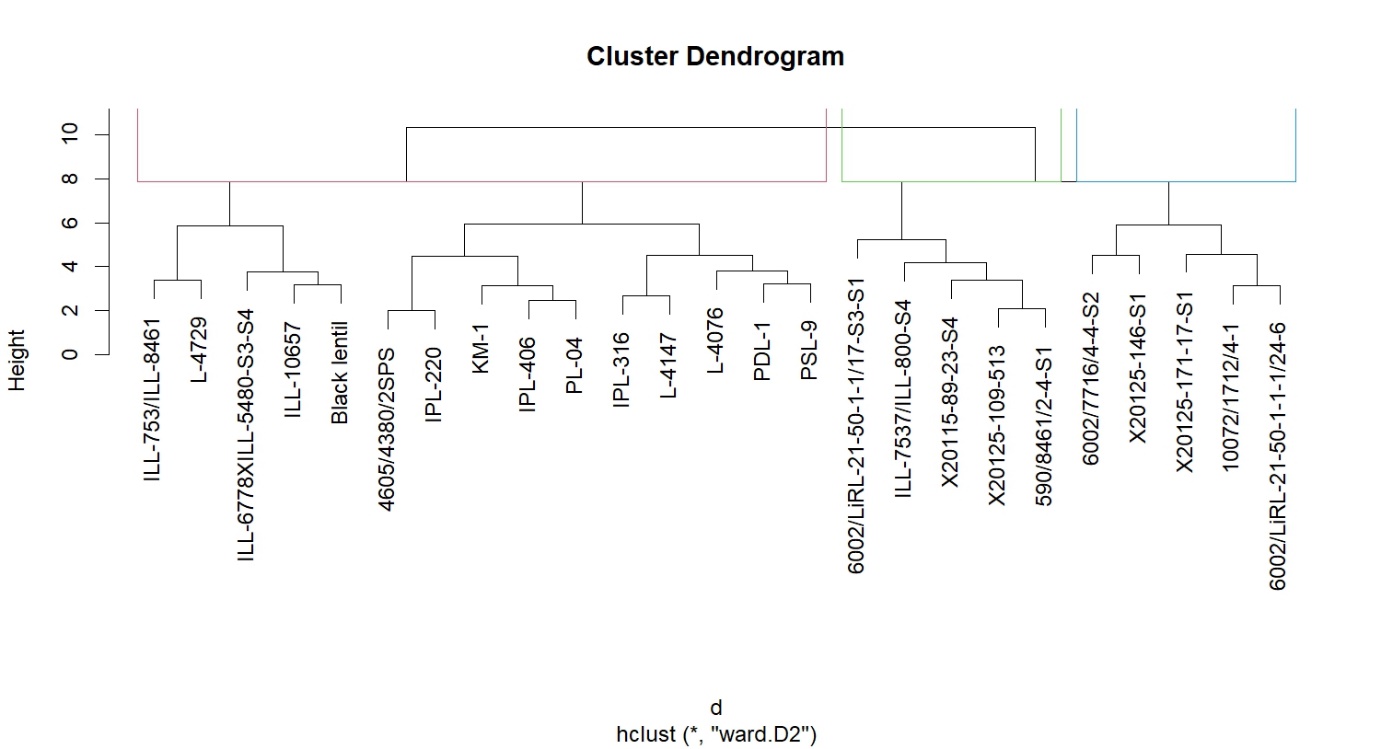
* 1. **Cluster Means for Different Characters**

The cluster means for all twelve traits are shown in Table 4. Among the clusters, cluster Cluster II exhibited the highest mean values for several traits, including the number of pods per plant (120.77), primary branches per plant (3.79), and seeds per pod (1.93). In contrast, cluster I had the highest mean values for iron content (94.60) and zinc (53.83). Cluster III displayed maximum values for biological yield (0.76), thousand seed weight (86.76), and overall yield (0.21), while clusterhad the lowest values for these traits in Table 5. The results indicate that no single cluster contained genotypes with all desirable traits. Therefore, crossing genotypes from different clusters is recommended to develop superior genotypes.

**Table 5. Cluster means for different characters in lentil genotypes.**

|  |  |  |  |
| --- | --- | --- | --- |
| **Cluster** | **I** | **II** | **III** |
| **DF** | 55.46\* | 58.50 | 61.00\*\* |
| **PB** | 3.51\* | 3.79\*\* | 3.50 |
| **SB** | 7.21 | 8.41\*\* | 6.76\* |
| **PH** | 36.98\* | 42.99\*\* | 42.64 |
| **PP** | 78.74\* | 120.77\*\* | 116.09 |
| **SP** | 1.87\* | 1.93\*\* | 1.91 |
| **DM** | 105.29\* | 107.53\*\* | 110.24 |
| **BY** | 0.66\* | 0.69 | 0.78\*\* |
| **TQW** | 25.47 | 23.86\* | 29.79\*\* |
| **Fe** | 94.60\*\* | 87.02 | 86.76\* |
| **Zn** | 53.83\*\* | 46.07\* | 51.47 |
| **SY** | 0.21\* | 0.22 | 0.26\*\* |

*\*Lowest value, and \*\*Highest value*



**Figure 1. Dendrogram showing the relationship between twenty-five genotypes including three checks of lentil genotypes.**

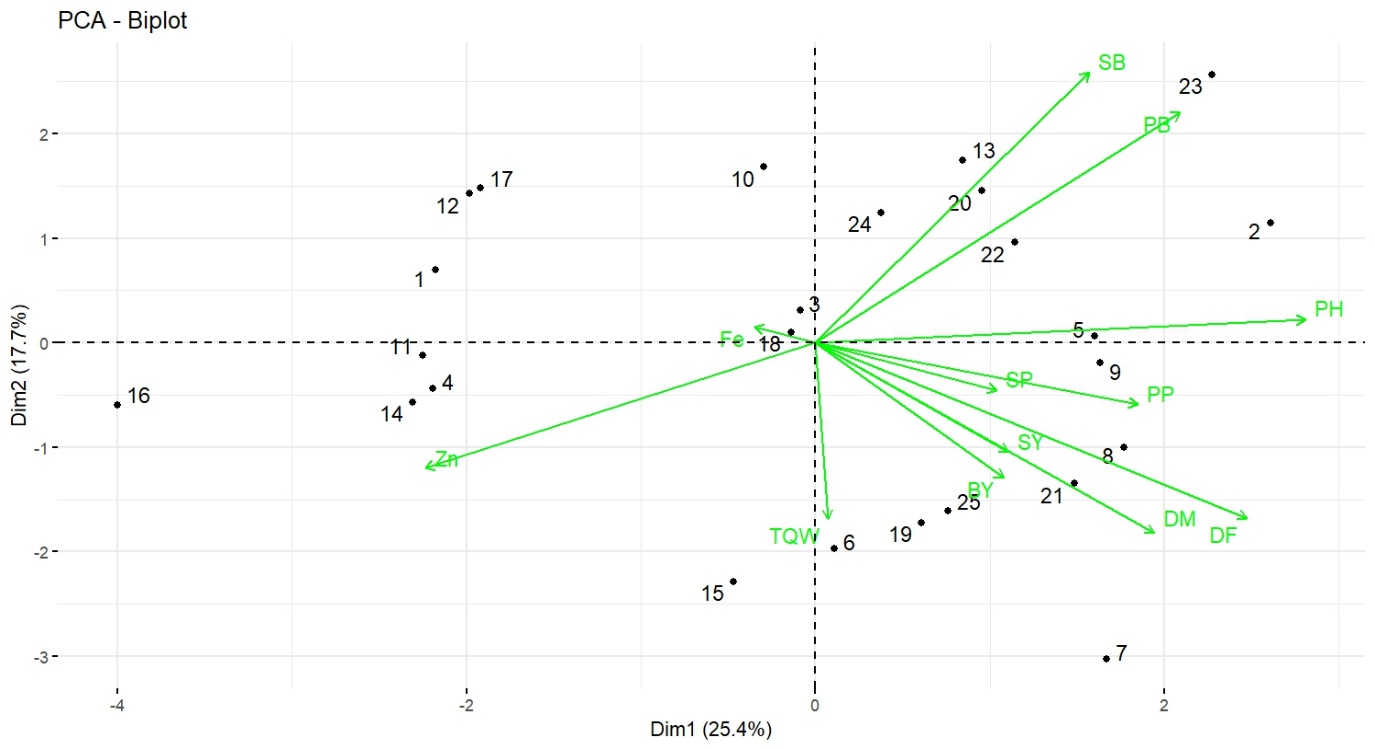
* 1. **Principal component analysis**

This method was first developed by Pearson (1901) and later utilized by Hotelling (1936). Several researchers are currently using it to select superior genotypes (Vianna *et al.*  2013 and Li *et al.* 2020). Principal Component Analysis (PCA) is a powerful tool in modern data analysis, as it is a well-known multivariate statistical technique used to identify the minimum number of components that can rank genotypes based on PC scores and explain the most variability. Principal components are generally estimated from either a correlation matrix or a covariance matrix. Considering the importance of PCA, this study focuses on lentil genotypes to identify the yield and yield-related traits responsible for the differences in yield among these genotypes. The estimates from the principal component analysis for the quantitative traits are presented in Table 6. The table reveals that the data from twelve traits were transformed into twelve principal components. The first four principal components, with eigenvalues greater than one, accounted for 72.30% of the total variation present in the studied genotypes. The first principal component accounted for the highest variation (25.43%), followed by PC2 (17.65%) and PC3 15.68%). The eigenvalues for the first four principal components are as follows: the first (3.05), the second (2.12), the third (1.88), and the fourth (1.63). The traits contributing to the variance for each principal component are shown in Table 5. According to Ramdath *et al.,* 2020 Principal component analysis revealed that the first three components explained 62.8% of the total variation, with SHAUC contributing the highest at 33.2%. These findings suggest that in vitro SHAUC, along with the combined effects of RDS and RS, could serve as effective indicators of the digestibility characteristics in cooked lentils.

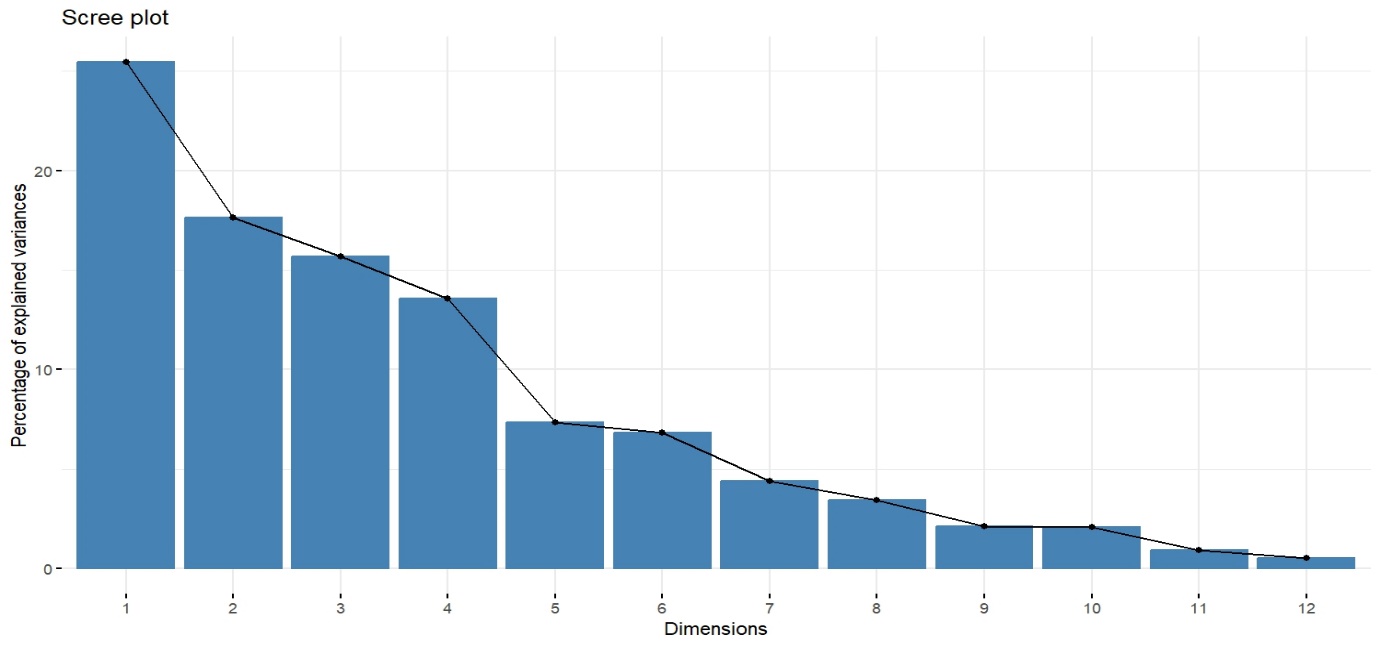
**Table 6. Principal component analysis for twelve quantitative traits in lentil genotypes**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Charcter** | **PC I** | **PC II** | **PC III** | **PC IV** |
| **Eigenvalue** | 3.05 | 2.12 | 1.88 | 1.63 |
| **Variance (%)** | 25.43 | 17.65 | 15.68 | 13.55 |
| **Cumulative variance (%)** | 25.43 | 43.08 | 58.76 | 72.31 |
| **Eigenvectors** | | | | |
| **DF** | **0.409** | -0.333 | 0.241 | 0.185 |
| **PB** | 0.345 | **0.438** | 0.021 | 0.151 |
| **SB** | 0.259 | **0.514** | 0.101 | 0.107 |
| **PH** | **0.464** | 0.044 | 0.003 | 0.172 |
| **PP** | **0.305** | -0.117 | 0.259 | -0.475 |
| **SP** | **0.172** | -0.090 | -0.119 | -0.565 |
| **DM** | **0.321** | -0.362 | 0.263 | 0.222 |
| **BY** | **0.178** | -0.257 | -0.561 | -0.051 |
| **TQW** | 0.012 | -0.335 | 0.174 | **0.239** |
| **Fe** | -0.058 | 0.030 | -0.262 | **0.481** |
| **Zn** | -0.370 | -0.238 | **0.249** | 0.114 |
| **SY** | **0.183** | -0.208 | -0.552 | 0.062 |

Principal Component I (PC I), which produced the most variability, was associated with days to 50% flowering, plant height, pods per plant, seeds per pod, days to maturity, biological yield, and seed yield per plot. The number of primary branches and secondary branches mainly contributed to PC II, while zinc content was significant in PC III. In PC IV, 1000 seed weight and iron content were prominent (Table 6). These results indicate that these traits contributed most to the divergence and accounted for the majority of the variability. Thus, the yield level was determined by these four components, representing a weighted average of the traits. The selection of the number of principal components (PCs) to retain is commonly guided by the magnitude of the eigenvalues, with components having eigenvalues greater than 1.00 considered significant. Since the total number of variables corresponds to the sum of the eigenvalues, this criterion helps in identifying the most informative components. In the present study, only the first four PCs had eigenvalues exceeding 1.00, indicating that they captured the majority of the variability among the lentil germplasm for yield-related traits (Table 6). These components represent the most influential dimensions in the dataset. Therefore, the traits contributing to these four principal components should be given priority in lentil breeding and improvement programs to enhance selection efficiency and genetic gain



**Figure 2. PCA Biplot of 25 genotypes including three checks of lentil genotypes for yield and yield-related traits.**

Principal Component Analysis (PCA) is an effective multivariate technique for identifying and determining the independent principal components that govern plant attributes individually. It can be used to select effective traits through indirect selection of superior genotypes. The gaps between traits indicate their associations; for instance, if two variables are positioned away from the origin and form an acute angle (less than 90°), they are positively correlated. This is exemplified by biological yield, as indicated by PC1 and PC2, which reflects the contribution of these traits toward divergence (Figure. 2). The length of the vector represents the magnitude of each character. The scree plot shows that the major contributions to divergence were primarily from PC1, followed by PC2, PC3, and PC4. A scatter plot of twelve characters using PC1 and PC2 is presented in (Fig. 2). The Scree plot reflected the eigenvalue of different principal components and the percent cumulative variability as shown by PCA of lentil genotypes based on quantitative traits in Figure 3.

**Figure 3. The scree plot shows the eigenvalues and cumulative variability of principal components in the PCA of lentil genotypes.**

**Conclusion**

The present study revealed significant of anova among 25 lentil genotypes and three checks, indicating a rich source of variability that can be effectively harnessed in lentil improvement programs. The significant genotypic variation observed for most traits suggests the presence of ample genetic potential for enhancing seed yield and micronutrient content, particularly iron and zinc, through selective breeding. Cluster analysis divided the genotypes into three distinct groups, with the greatest inter-cluster distance observed between Clusters II and III, indicating the possibility of producing high-quality recombinants through inter-cluster hybridization. Principal component analysis further supported the diversity, with four components explaining over 72.31% of the total variation. High-yielding genotypes such as ILL-753/ILL-8461, ILL-7537/ILL-800-S4, and Black lentil lines, as well as nutrient-rich genotypes like IPL-316 and X20115-89-23-S4, demonstrated their potential as valuable genetic resources. These findings provide a strong foundation for selecting elite parental lines to develop improved cultivars with superior yield potential and enhanced nutritional quality. The integration of such diverse and high-performing genotypes into breeding programs could contribute significantly to food and nutritional security, particularly in regions dependent on lentils as a primary protein and micronutrient source. Future efforts should focus on utilizing these diverse and high-performing genotypes in strategic hybridization and selection schemes to develop improved lentil cultivars with enhanced yield and nutritional traits.

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