***Original Research Article***

**Impact of Organic Additives on Soil Organic Carbon and Enzyme Activities in Intensive Ginger Agro-ecosystems**

**Abstract**

 Organic manure is a good source of nutrients, which enhances soil productivity, increases the soil organic carbon content, enhances the activities of soil microorganisms, and supplies major plant nutrients. Nutrient management is one of the key determinants of crop production. Variability in nutrient additions through fertilisers and organic manures appears to be an influencing factor on soil fertility status and ginger productivity. This study was conducted to assess different magnitudes of organic and inorganic nutrient inputs on soil carbon stock and soil enzyme activities. A survey-based study was conducted. The data obtained was subjected to one-way ANOVA using Excel software version of Windows MS Office -2007. In the Banavasi area of Uttara Kannada, Ginger farmers adopted different nutrient management practices in ginger cultivation were grouped into Local Farmers-1: High OM and High Fertilisers, Local Farmers-2: Low OM and Moderately High Fertilisers and Migrated Farmers: Very Low OM and Very High Fertilisers. Soil samples were collected at two sampling depths (0-15 cm and 15-30 cm) in three groups of ginger farmers’ fields. The mean rainfall of this region is 2500 mm, with up to 103 rainy days in a year. Annually, this region has 25 hot days with temperatures more than 350 C and 124 warm days ranging 30-350 C and remains below 350 C during the rest of the year for about 214 days. The soil organic carbon content was higher in soils of Local Farmers-1 compared to Local Farmers-2: Low OM and Moderately High Fertilizers and Migrated Farmers: Very Low OM and Very High Fertilizers thus, the study revealed that application of organic manures in intensively managed ginger production systems helps to enhance soil carbon stocks and maintain soil biological activities.

**Key words:** Organic manures, fertiliser, Ginger production, Soil-C stocks, Urease, Phosphatase, Dehydrogenase

**INTRODUCTION:**

Ginger (*Zingiber officinale*) is an important commercial spice crop and has also long been used as a medicinal plant. It is a perennial herb member of the *Zingiberaceae* family, and its thick tuberous rhizomes are very popular for medicinal uses and as a spice and additive agent for flavouring foods and drinks (Arcusa et al.,2022). Especially, the fresh and dry rhizomes of ginger are widely used in folk medicine for the treatment of colds, sore throats, asthma, joint pain, and also to stimulate appetite (Egamberdieva and Jabborova, 2018). As a spice, ginger is one of the most widely used condiments for various foods and beverages (Jabborova *et al.,* 2021). Further, ginger extracts from both fresh and dried rhizomes are used extensively in the food, beverage, and confectionery industries (Jabborova and Egamberdieva, 2019). In terms of mineral nutrients, the rhizome is rich in calcium, phosphorus and potassium. Ginger cultivation is expanding throughout the world as its demand is spreading for medicinal and beverage industries are increasing. (Asfaw and Demissew, 2009). India is the largest producer, consumer and exporter of ginger (Srinivasan *et al.* 2019). Our country accounts for almost 30 per cent of the world’s total ginger production (Singh *et al.* 2015). In Karnataka, it is grown in an area of 23.09 thousand hectares with an annual production of 58.39 thousand metric tonnes (Anon., 2018)

Nutrient management is essential in achieving the best growth and productivity in any crop (Srinivasan *et al.* 2019). Nutrient management is essential in achieving the best growth and productivity in ginger, in addition to soil type. Because it is a crop that exhausts nutrients, it needs a sufficient supply during key growth periods (Prasath et al., 2024; Ravi et al., 2022). A good nutrient management practice not only sustains crop production but also helps to improve soil health and enhance nutrient use efficiency (Dinesh *et al*. 2012). The productivity of ginger is affected due to poor nutrient management Dinesh *et al.,* 2012a), as it is a nutrient-exhaustive crop and therefore requires an adequate supply of nutrients at specific stages of its growth (Weiss, 1997). Inappropriate agricultural intensification coupled with reckless use of fertilisers can deteriorate soil quality in terms of soil physicochemical and biological properties (Lal 2015). Injudicious use of fertilisers can damage the natural ecosystem and cause soil and water pollution (Pathak and Ram 2013).

Soil structure plays a pivotal role in determining the overall productivity and sustainability of agricultural systems (Ramasamy et al.,2024). In this context, Organic manure is a good source of nutrients which enhances soil productivity, increases the soil organic carbon content, enhances the activities of soil microorganisms and supplies major plant nutrients (Sanchez and Miller 1986). Further, Organic practices increase the soil carbon content and the overall biological activities (Bai *et al.* 2019). Even organic wastes from different sources can be used to improve soil fertility and productivity (Palm *et al.* 2001). The organic manure feeds both plants and the soil microorganisms simultaneously and thus, enhances soil health and its quality, leading to overall improvement of the agroecosystem. Exclusive use of fertilisers can significantly reduce microbial activity (as measured by dehydrogenase, acid phosphatase and other enzymes (Parham *et al.* 2002; Ning *et al.* 2017).

 The biochemical properties are more sensitive to environmental stress and provide rapid and accurate estimates of soil quality. These soil biochemical parameters have been considered as potential indicators of soil quality (Nannipieri *et al.* 2002). Application of organic manures helps to improve soil structure, soil moisture retention, and available water content and reduce bulk density. Enhancing SOC through organics is crucial for both sustaining intensive production as well as mitigating greenhouse gases (GHGs). However, there are or no little efforts on the influence of the combined use of organics and inorganics and their influence on soil-C stocks and soil enzymes at the farmer’s field. Thus, a survey-based study was carried out to assess the magnitude of organic and inorganic inputs application on soil organic carbon stocks and soil enzyme activities in soils cropped with ginger.

**MATERIALS AND METHODS**

**General description of the study area**

Ginger is widely grown in the Hilly zone of Karnataka, where the soils mostly belong to alfisols and oxisols, having slightly acidic, medium-textured and well-drained soil conditions. Thus, these regions provide optimum conditions for ginger cultivation. In Karnataka, ginger is widely grown in Shivamogga, Chikkamagaluru, Uttara Kannada, Udupi and Mysuru districts. In the Uttara Kannada district, the Ginger farmers are spread over a large area. However, the study was restricted to the Banavasi region of Uttara Kannada district the mean rainfall of this region is 2500 mm, with up to 103 rainy days in a year. Annually, this region has 25 hot days with temperatures more than 350 C and 124 warm days ranging 30-350 C and remains below 350 C during the rest of the year for about 214 days

**Selection and categorisation of ginger farmers**

 An initial survey was carried out in the study areas to identify the farmers involved in ginger cultivation. During the survey, the information on yield and proper nutrient management practices adopted was collected. It was interesting to note that the rhizome yield varied from as high as 30-35 tonnes ha-1 in high nutrient applied fields to as low as 15-18 tonnes ha-1in low nutrient applied plots. Nutrient management appeared to be a key factor in determining the productivity compared to other management practices such as seed material, irrigation schedules and plant care measures. Thus, based on the organic manures and fertilisers applied, the farmers were grouped as **Local Farmers-1**: High organic manures and high fertilisers (G1), **Local Farmers-2**: Low organic manures and moderately high fertilizers (G2), **Migrated Farmers**: Very low organic manures and very high fertilizers (G3). and the details are given in Table 1 and presented in Fig.1.

**Soil sample collection**

The soil samples were collected from each of the selected ginger fields (15 farmers) at two depths (0-15 cm and 15-30 cm) after the harvest of the crop during January-February 2020. At each location, the samples were collected at 8-10 spots and pooled to get one composite sample, and the collected soil samples were dried in the shade, powdered and then sieved through a 2 mm sieve.

**Soil C stocks assessment**

 **Soil organic carbon**

The soil organic carbon was determined by the wet oxidation method (Walkley and Black, 1934). A known weight of 0.2 mm sieved soil sample was treated with an excess volume of standard potassium dichromate solution in 500 ml conical flasks, and 25 ml of concentrated sulphuric acid was added. The soil suspension was kept undisturbed for 30 minutes. The unused potassium dichromate was determined by back titration using standard ferrous ammonium sulphate solution in the presence of ferroin indicator. The soil organic carbon contents were expressed in per cent.

 **Soil carbon stock**

Carbon stock for a given soil layer was estimated by multiplying the soil organic carbon content by the corresponding soil bulk density of that particular layer (Batjes, 1996). The total soil carbon stock was estimated by using the formula as given below.

Carbon stock (t ha-1) = Volume of soil in ha x Soil BD x SOC

**Soil enzyme analysis**

 The stored soil samples were pre-incubated at 25 °C for 2 days, after adjusting the soil moisture content to field capacity, to rejuvenate soil biological activity. These pre-incubated soils were analysed for dehydrogenase, acid phosphatase and urease enzyme activities. The dehydrogenase activity was determined by incubating soil samples at 37 °C with 3 per cent of 2, 3, 5- 5-triphenyl tetrazolium chloride (TTC) aqueous solution. The amounts of triphenyl formazon (TPF) formed were extracted with methanol and measured at 485 nm (Casida et al. 1964).

 The acid phosphatase activity was determined by the *p-*Nitrophenol Phosphate (*p*-NPP) method (Eivazi and Tabatabai 1977). A known weight of the soil sample (< 2 mm) was incubated with four ml of modified universal buffer (pH 6.5 for any assay of acid phosphatase). One ml of *p*-NPP solution (as substrate) and 0.2 ml of toluene (to suppress further microbial activity). After one hour, the p-nitrophenol (*p*-NP) formed in the supernatant was extracted by centrifugation and the intensity of yellow colour was measured at 420 nm.

 For the urease enzyme, the soil samples (2mm) were incubated with urea and citrate buffer solution for three hours at 37 ºC. The soils were extracted and the amount of ammonia produced was measured colourimetrically by the phenate-NaOCl method (Hofmann 1963).

**Statistical analysis**

 The data obtained was subjected to one-way ANOVA. using Excel software version of Windows MS Office -2007.

**RESULTS AND DISCUSSION**

**Soil carbon stock**

 The soil carbon stock of ginger farmers' fields soil samples was estimated by multiplying the SOC content with the corresponding BD based soil mass in the top 30 cm layer of a 1 ha area.

**Soil organic carbon**

 The application of organic manures significantly influenced the SOC content, and it ranged from 0.77 to 1.33 per cent. The soil organic carbon content in surface soils was found significantly higher in fields of local farmers-1 (1.33 ± 0.08 %) compared to local farmers-2 (0.82 ± 0.06 %) and migrated farmers (0.77 ± 0.10 %). The soil OC content was found to be substantially lower in sub-surface soils compared to surface soils. The corresponding organic carbon content in sub-surface soils was 1.08 ± 0.05 per cent, 0.50 ± 0.07 per cent and 0.46 ± 0.06 per cent in local farmers- 1, local farmers- 2 and migrated farmers, respectively. However, the carbon content in the soils of local farmer-1 was found to be significantly higher compared to the other two groups.

 The variations in SOC among different ginger growers may be attributed to additions of different levels of organic manure (Niranjana *et al.* 2018). The quality of organic matter is also an important factor in any ecosystem (Dattaraja *et al.* 2018). As its additions determine the soil organic matter content (Grewal et *al.,*1981 and Kaushik et *al.,*1984). The surface soils recorded higher soil organic carbon content compared to sub-surface soils. This was due to the various carbon input distributions in terms of root biomass, root exudates, rhizospheric deposition, followed by slow decomposition due to poor oxygenic conditions for microbial oxidation at deeper soil layers. The results are consistent with the findings of Padbhushan *et al*. (2016) and Mahanta *et al.* (2013). Further, the upper layer remains in dynamic equilibrium with biological and anthropological activities and thus is generally richer in C than the lower layers.

**Soil bulk density**

 The bulk densities of surface soils were significantly higher in migrated farmers (1.24 ± 0.07 Mg m-3) compared to soils of local farmers-2 group (1.19 ± 0.03 Mg m-3) and soils of local farmers-1 group (1.15 ± 0.07 Mg m-3). Similarly, the bulk density values in sub surface soils were found to be higher than surface soils with respective values ranging from 1.29 ± 0.06 Mg m-3, 1.24 ± 0.04 Mg m-3 and 1.22 ± 0.04 Mg m-3 among migrated farmers, local farmers-2 and local farmers-1 respectively (Table.2). The variation in bulk density among three groups of ginger growers could be attributed to the differences in organic manure applications. The organic matter addition improves soil structure, pore size distribution and soil water transmission (Srikanth *et al.,* 2000). The low bulk density values in soils of local farmers-1 may be attributed to better soil structure as a result of higher organic inputs (Sharma *et al*., 2001). The soil organic carbon content among different groups was in concurrence with these observations. Lower bulk density in surface soils may be due to loosening of soils during ginger harvest by manual hand tools and machines would have reduced soil bulk density (Bhavya *et al*., 2018). On the otherhand, Higher bulk density in sub-surface soils might be due to compaction as a result of the pressure exerted by the upper layers (Patil and Jagdish, 2004). It may also be due to the fact that the organic matter is mostly added to the surface layers and only a small portion of it would reach the sub-surface layers (Tejada *et al*, 2008 and Nagaraja, 1997).

**Soil carbon stock**

 Soil carbon stock was found highest in Local Farmers-1 and in surface (23.17 ± 1.60 t ha-1) and sub surface (18.14 ± 2.03 t ha-1) soils respectively) and least was noted in migrated farmers in surface (03.62 ± 1.68 ) t ha-1 and sub surface (6.67 ± 1.70 t ha-1)soils respectively. The C stock at 0-15 cm depth was higher in surface soil compared to sub-surface soils. However, the total carbon stock at 0-30 cm soil depth among different ginger grown soils varied significantly in the order Local Farmers-1 (41.30 ± 3.39 t ha-1 ) > Local Farmer-2 (23.26 ± 3.06 t ha-1 ) > Migrated Farmers (20.08 ± 3.21 t ha-1 ). 57. Table 3.

 The variation in soil carbon stocks in different soils might be due to the application of organic manure. The organic manure additions contribute to enhanced soil organic carbon contents.. Similar results were observed in studies conducted by Roy *et al.* 2007 and Dattaraja *et al.* 2018). However, soil carbon stocks were higher in surface soils compared to sub-surface soils This might be due to the direct relation between the organic carbon and soil carbon stocks, therefore, soil carbon stocks also follow the same trend of organic carbon, it decrease with increasing depth (Balloli *et al.,* 2007).

**Soil enzymatic activities**

 Soil enzymatic activities and Biochemical parameters are considered to be very sensitive indicators of both short-term and long-term changes in soil quality.

**Urease enzyme activity:**

 Urease activities are involved in the hydrolysis of urea in the soil. Among the different groups, urease activity was significantly higher in migrated farmers' fields with an activity of 132.81 ± 8.59 µg NH4+ released g-1 soil hr-1 in surface and 90.56 ± 9.67 µg NH4 + released g-1 soil hr-1 sub-surface soils, respectively. Contrastingly, fields of local farmers recorded least with values of 112.97 ± 6.26 and 83.06 ± 4.27 µg NH4 + released g-1 soil hr-1 in surface and sub-surface soils. Higher urease activity in migrated farmers' fields may be due to high use of fertiliser inputs, including urea. which promoted the urease activity (Shen *et al.,* 2010). The application of both organic manure and fertilisers appears to be substantial in ginger cultivation, which consequently resulted in high urease activity (Sharan *et al.,* 2020). In addition, Urease enzyme activity was higher in surface soils than sub-surface soils. The higher soil organic carbon content and available-N in ginger cultivated soils studied would further strengthen these observations. Lesser urease activity in sub-surface layers might be due to the fact that application of organic manures and fertilisers is confined to the surface layer (Rao *et al.,* 1989).

**Dehydrogenase activity:**

Dehydrogenase enzyme is used as an index of soil biological activity and is expressed in µg TPF g-1 of soil day-1. Its activity was found to be significantly higher in ginger soils of local farmers (15.21 ± 2.92 µg TPF g-1 of soil day-1 and 9.30 ± 0.40 µg TPF g-1 of soil day-1 in surface and sub surface soils, respectively) while the least was noticed in soils of migrated farmers (9.12 ± 2.81 and 6.73 ± 1.73 µg TPF g-1 of soil day-1 in surface and sub surface soils, respectively). The dehydrogenase activity was found to be higher in surface soils than sub-surface soils. Higher dehydrogenase activity in plots of local farmers may be due to high application of organic manure. Thus, the addition of organic manures enhances overall biological activity and is generally measured in terms of dehydrogenase (Martens *et al.,* 1992). Whereas, the enzymatic activity is low in migrated farmers who applied the highest fertiliser inputs might be due to the reason that mineral fertilisation had weaker effects on dehydrogenase activity as compared to organic manuring (Shen *et al.,* 2010). The organic manure additions, soil organic carbon contents and dehydrogenase activities are in concurrence with the above observations. Decreased enzymatic activity in the sub-surface layer with low soil organic matter may be attributed to an increase in soil depth, and lower organic matter content further strengthens the role of soil organic matter. Similar reports are made by several authors (Nagaraja, 1997; Shivakumar, 2010; Sharan *et al.,* 2020; and Dinesh *et al.,* 2010).

**Acid phosphatase enzyme activity**

 The phosphatase activity in soils of ginger different growers are presented table 4 The activity was found significantly higher in local farmers-1 (22.38 ± 2.23 µg PNP g-1 of soil hr-1 and 11.45 ± 1.70 µg PNP g-1 of soil hr-1 in surface and sub surface soils respectively) while least activity was found in migrated farmers (15.45 ± 3.91 µg PNP g-1 of soil hr-1 and 8.83 ± 0.49 µg PNP g-1 of soil hr-1). However, phosphatase activity was higher in surface soils compared to sub-surface soils. Higher enzymatic activity in local farmer-1 fields could be due to the fact that the phosphatase activity is generally higher in organically amended soils. The phosphatase activity and organic carbon were positively correlated in the present study also suggesting that that organic matter significantly increases the phosphatase activity in soil and the results are in accordance with that of Dick (1994) and Tabatabai (1984). Least activity was noticed in soils of migrated farmers which could be attributed to marked suppression in phosphatase activity due to P fertilization (Wang *et al.,* 2008). However, the decrease in enzymatic activity in sub-surface soils may be attributed to lower organic matter content (Dinesh *et al.,* 2010)

### Table 1. Categorization of ginger farmers based on the nutrients applications

|  |  |  |
| --- | --- | --- |
| **Group of Ginger farmers** | **Organic manures used****(t ha-1)** | **Nutrients through fertilizers (kg ha-1)** |
| **N** | **P2O5** | **K2O** |
| G1: Local Farmers -1(High OM + High Fertilizer) | 27.41 ± 2.78 | 101.52 ± 32.92 | 75.17 ± 15.44 | 80.19± 8.02 |
| G2: Local Farmers -2(Low OM + Mod. High Fertilizer) | 15.65 ± 2.55 | 134.45± 21.67 | 122.76± 21.57 | 151.65 ± 18.06 |
| G3: Migrated Farmers(Very Low OM + Very High Fertilizer) | 11.65 ± 1.77 | 198.80± 9.34 | 162.22± 8.24 | 195.25 ±13.87 |

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**NOTE**: The recommended dose of N, P2O5, K2O nutrients for ginger is-100:50:50 kg ha-1 and 25 t ha-1 FYM

**Table 2. Soil organic carbon and Bulk density in soils of different groups of ginger fields**

|  |  |  |
| --- | --- | --- |
| **Groups of ginger farmers** | **Soil organic carbon****(%)** | **Bulk density****(Mg m-3** |
| **0-15 cm** | **15-30 cm** | **0-15 cm** | **15-30 cm** |
| G1: Local Farmers -1(High OM + High Fertilizers) | 1.33 ± 0.08a | 1.08 ± 0.05a | 1.15 ± 0.07b | 1.22 ± 0.04b |
| G2: Local Farmers -2(Low OM + Mod. High Fertilizers) | 0.82 ± 0.06b | 0.50 ± 0.07b | 1.19 ± 0.03b | 1.24 ± 0.04b |
| G3: Migrated Farmers(Very Low OM + Very High Fertilizers) | 0.77 ± 0.10b | 0.46 ± 0.06b | 1.24 ± 0.07a | 1.29 ± 0.06a |
| **S. Em. ±** | 0.02 | 0.03 | 0.019 | 0.016 |
| **CD (p=0.05)** | 0.07 | 0.09 | 0.05 | 0.04 |

**Note:** The extent of significant differences among 3 groups for each parameter is indicated by using alphabets

**Table 3. Soil carbon stock in soils of different groups of ginger farmer fields**

|  |  |  |
| --- | --- | --- |
| **Groups of ginger farmers** | **Carbon stock****(t ha-1)** | **Total carbon stock****(t ha-1)** |
| **0-15 cm** | **15-30 cm** | **0-30 cm** |
| G1: Local Farmers -1(High OM + High Fertilizers) | 23.17 ± 1.60a | 18.14 ± 2.03a | 41.30 ± 3.39a |
| G2: Local Farmers -2(Low OM + Mod. High Fertilizers) | 14.84 ± 1.32b | 8.42 ± 2.01a | 23.26 ± 3.06b |
| G3: Migrated Farmers(Very Low OM + Very High Fertilizers) | 13.62 ± 1.68b | 6.67 ± 1.70b | 20.28 ± 3.21c |
| **S. Em. ±** | 0.48 | 0.60 | 1.02 |
| **CD (p=0.05)** | 1.42 | 1.77 | 2.97 |

**Note:** The extent of significant differences among 3 groups for each parameter is indicated by using alphabets

**Table 4. Soil enzymatic activities under different ginger fields**

|  |  |  |  |
| --- | --- | --- | --- |
| **Groups of ginger farmers** | **Urease****(µg NH4+ released g-1 soil hr-1)** | **Dehydrogenase****(µg TPF g-1 of soil day-1)** | **Acid phosphatase****(µg PNP g-1 of soil hr-1)** |
| **0-15 cm** | **15-30 cm** | **0-15 cm** | **15-30 cm** | **0-15 cm** | **15-30 cm** |
| G1: Local Farmers -1(High OM + High Fertilizers) | 112.97 ± 6.26b | 83.06 ± 4.27a | 15.21 ± 2.92a | 9.30 ± 0.40a | 22.38 ± 2.23a | 11.45 ± 1.70a |
| G2: Local Farmers -2(Low OM + Mod. High Fertilizers) | 129.83 ± 9.53a | 85.49 ± 6.68a | 11.29 ± 2.01b | 6.90 ± 1.78b | 17.52 ± 3.08b | 9.34 ± 0.53b |
| G3: Migrated Farmers(Very Low OM + Very High Fertilizers) | 132.81 ± 8.59a | 90.56 ± 9.67a | 9.12 ± 2.81b | 6.73 ± 1.73b | 15.45 ± 3.91b | 8.83 ± 0.49b |
| **S. Em. ±** | 2.60 | 2.28 | 0.82 | 0.46 | 0.99 | 0.33 |
| **CD (p=0.05)** | 7.61 | NS | 2.41 | 1.34 | 2.91 | 0.98 |

**Note:** The extent of significant differences among 3 groups for each parameter is indicated by using alphabets

**CONCLUSION**

 It was observed from the present study that the soils of the group-1 ginger growing farmers with high organic manure additions recorded higher soil carbon stock compared to low organic manure added ginger fields belonging to group-2 and group-3 farmers. In terms of biological activity, the activity of dehydrogenase, and acid phosphatase enzymes increased with an increase in organic manure applications. Thus, the addition of organic manure plays a crucial role in maintaining soil health and soil biological properties.

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Asfaw, N., Demissew, S., 2009. Aromatic plants in Ethiopia. Shoma Books, Addis Ababa, pp. 27–185.

Balloli, S. S., Sharma, K. L., Kausalya, R., Ramesh, V., Venkateswarlu, B. and Ramakrishna, Y. S., 2007, Impact of land use practices on soil fertility status of dry land Alfisols. *Indian J. Dry Land Agric. Res. Dev.,* 22(2): 163-166.

Batjes, N. H., 1996, Total carbon and nitrogen in soils of the world.*Eur. J. Soil Sci*., 47: 151-163.

Bhavya, V. P., Anil Kumar, S., Ashok Alur., Shivanna, M. and Shivakumar, K. M., 2018, Changes in soil physical properties as a result of different land use systems with depth. *Int. J. Curr. Microbiol. App. Sci.,*7(1): 323-327.

Casida, L. E., Klein, D. A. and Santoro, T., 1964, Soil dehydrogenase activity.*Soil Sci*., 98: 371-376.

Dattaraja H.S., Pulla S, Suresh H.S., Nagaraja M.S., Srinivasamurthy C.A., and Sukumar, R., 2018 Woody plant diversity in relation to environmental factors in a seasonally dry tropical forest land scape. *J. Vegtn. Sci.,* 18: 1-11.

Dick, R. P., Sandor, J. A. and Eash, N. S., 1994, Soil enzyme activities after 1500 years of terrace agriculture in the Colca Valley, Peru. *Agric.Ecosyst. Environ.*, 50(2): 123-131.

Dinesh R, Srinivasan V, Hamza S, Manjusha A. 2012 Shortterm incorporation of organic manures and biofertilizers influences biochemical and microbial characteristics of soils under an annual crop [Turmeric (*CurcumalongaL.*)]. Bioresour Technol 101: 4697-4702.

Dinesh, R., Srinivasan, V., Hamza, S. and Manjusha, A., 2010, Short-term incorporation of organic manures and biofertilizers influences biochemical and microbial characteristics of soils under an annual crop [Turmeric (*Curcuma longa L.*)]. *Bioresource Technol.,*101: 4697-4702.

 Egamberdieva, D., Jabborova, D., 2018. Medicinal plants of Uzbekistan and their traditional uses. Springer Nature Switzerland AG. In: D, Egamberdieva., M, Öztürk., (Eds.), Vegetation of Central Asia and Environs. Springer Nature Switzerland AG, pp. 211–237.

Eivazi, F, Tabatabai M.A., 1977 Phosphatases in soils. *Soil Biol. Biochem.,* 9:167-172.

Grewal, J. S., Sharma, R. C. and Sud, K. C., 1981, Effect of continuous application of PK- fertilizer and FYM on potato yield and some soil properties*. J. Indian Soc. Soil Sci.,* 29:129-131.

Hofmann, E., 1963 Methods of enzymatic analysis. New York, Academic, USA: 913-946

Jabborova, D. and Egamberdieva, D., 2019. Antibacterial, antifungal, and antiviral properties of medicinal plants. In: D. Egamberdieva., A. Tiezzi., (Eds.), Medically Important Plant Biomes: Source of Secondary Metabolites Springer Nature Singapore Pte Ltd, pp. 51–65.

Jabborova, D., Enakiev, Y., Sulaymanov, K., Kadirova, D., Ali, A. and Annapurna, K., 2021. Plant growth-promoting bacteria Bacillus subtilis promote growth and physiological parameters of *Zingiber officinale* Roscoe. Plant Sci. Today. 8, 66–71.

Kaushik. R. D., Verma. K., Dang, Y. P., Sharma, A. P., Verma, S. L. and Pannu, B. S., 1984, Effect of nitrogen and farm yard manure on yield of crops, nutrients uptake and soil fertility in Paddy-Wheat rotation. *Indian J. Agric. Res.,* 18: 73-78.

Lal R (2015) Restoring soil quality to mitigate soil degradation. Sustainability *7*: 5875-5895.

Martens, D. A., Johanson, J. B. and Frankenberger, W. T. 1992, Production and persistence of soil enzyme with repeated addition of organic residues. *Soil Sci.,* 153: 53-61.

Nagaraja, M.S., 1997, Biomass turnover, nutrient status and biological processes among different land use systems. *Ph.D. Thesis,* UAS. Bangalore

Nannipieri P, Kandeler E, Ruggiero P 2002 Enzyme activities and microbiological and biochemical processes in soil, In enzymes in the environment: Activity, ecology, and applications, Edn Burns RG, Richard Dick, pp 1-33

Ning C.C., Gao P.D., Wang B.Q., Lin W.P., Jiang N.H., Cai K.Z., 2017 Impacts of chemical fertilizer reduction and organic amendments supplementation on soil nutrient, enzyme activity and heavy metal content. *J. Integrative Agri.* 16(8):1819-1831.

Niranjana K.S., Yogendra K., and Mahadevan K.M.(2018) Physicochemical characterization and fertility rating of maize growing soils from hilly zone of Shivamogga district, Karnataka. *Indian J. Agric. Res.* 52(1):56-60.

Palm C.A., Giller K.E., Mafongoya P.L., 2001 Management of organic matter in the tropics: Translating theory into practice. *Nutr. Cycl. Agroecosyst.* **61**:63-75.

Parham J.A., Deng S.P., Raun W.R., Johnson G.V., 2002 Longterm cattle manure application in soil. I. Effect on soil phosphorus levels, microbial biomass C and dehydrogenase and phosphatase activities. *Biol. Fertil. Soils*. **35**:328- 337.

Pathak R.K., Ram R.A., 2013 Bio-enhancers: A potential tool to improve soil fertility, plant health in organic production of horticultural crops. Progressive Horticulture 45(2):237- 254

Patil, R.B. and Jagdish, P., 2004, Charzacterization and classification of some sal *(Shorearobusta)* supporting soils in Dindori district of Madhya Pradesh*. J. Indian Soc. Soil Sci.,* 52(2): 119-125.

Rao, D. L. N. and Ghai, S. K. 1989., Urease and dehydrogenase activity of alkali and reclaimed soils. *Australian J. Soil Res.,* 23: 661-665.

Sharan. B., R, Nagaraja, M. S., Mallesha, B. C. and Kadalli, G. G., 2020, Enzyme activities at varied soil organic carbon gradients under different land use systems of Hassan District in Karnataka, India. *Int. J. Curr. Microbiol. Appl. Sci*.,9(3): 1739-1745.

Sharma, M. P., Bali, S. V. and Gupta, D. K., 2001, Crop yield and properties of black soils as influenced by residue management under rice-wheat-maize cropping sequence.*J. Indian Soc. Soil. Sci.,*50: 6-9.

Shen, W., Lin, X., Shi, W., Min, J., Gao, N., Zhang, H., Yin, R. and He, X., 2010, Higher rates of nitrogen fertilization decrease soil enzyme activities, microbial functional diversity and nitrification capacity in a Chinese polytunnel greenhouse vegetable land. *Plant Soil*.,337: 137–150.

Shivakumar, K. M., 2013, Studies on carbon and nutrient dynamics in different land use systems of a watershed in the Western Ghats. *Ph. D. Thesis,* UAS., Bangalore.

Singh, S.P., 2015. Nutrient supplementation through organic manures for growth and yield of ginger (*Zingiber officinale* Rose). J. Eco-friendly Agric. 10, 28–31.

Srikanth, K., Srinivasamurthy, C. A., Siddaramappa, R. and Ramakrishna, V. R., 2000, Direct and residual effect of enriched compost, FYM, Vermicompost and fertilizer on soil properties of an Alfisols. *J. Indian Soc. Soil Sci.,*48(3): 496-499.

Srinivasan V, Thankamani CK, Dinesh R, Kandiannan K, Hamza S, Leela NK, John Zachariah T (2019) Variations in soil properties, rhizome yield and quality as influenced by different nutrient management schedules in rainfed ginger. *Agri. Res.* 8(2):218-230.

Tabatabai, M. A., 1984, Soil Enzymes. In: *Methods of Soil Analysis*, part 2. (Eds.) A. L. Page, R. H. Miller and D. R. Keeney, American Society of Agronomy, Soil Science Society of America, Madison, Wisconsin, USA. pp. 903-947.

Tejada, M., Garcia, C., Gonzalez, J. L., Hernandez, M.T., 2008, Use of organic amendment as a strategy for saline soil remediation: influence on the physical, chemical and biological properties of soil. *Soil Biol. Biochem.,* 38: 1413-1421.

Wang, Q. K., Wang, S. L., Liu and Y. X., 2008, Responses to N and P fertilization in a youngEucalyptus dunnii plantation: microbial properties, enzyme activities and dissolvedorganic matter. *Appl. Soil Ecol*., 40: 484-490.

Weiss, E.A., 1997. Essential oil crops. CAB International, Wallingford, pp. 539–567. Xaziev, F.X., Xapbed, U.X., 2005. Methods of Soil Enzymology. M.: Science, ISBN: 5020339407, 252.

Arcusa, R., Villaño, D., Marhuenda, J., Cano, M., Cerdà, B., & Zafrilla, P. (2022). Potential role of ginger (*Zingiber officinale* Roscoe) in the prevention of neurodegenerative diseases. *Frontiers in Nutrition*, *9*, 809621.

Prasath, D., Srinivasan, V., Sial, P., Leela, N. K., Akshitha, H. J., & Raghuveer, S. (2024). Ginger. *Handbook of Spices in India: 75 Years of Research and Development*, 1661-1792.

Ravi, Y., Narayanpur, V. B., Gangadharappa, P. M., Hiremath, J. S., Shantappa, T., Prashant, A., & Doddamani, M. B. (2022). Performance evaluation of ginger genotypes on their yield and nutrient uptake under the coastal Karnataka, India. *International Journal of Bio-resource and Stress Management*, *13*(6), 630-637.

Ramasamy, M., Ghosh, S., Yadav, K. K., Chitra, M., B, D., S.A, S. kanth, & Karthickraja, A. (2024). The Role of Organic Farming in Enhancing Soil Structure and Crop Performance: A Comprehensive Review. *Journal of Scientific Research and Reports*, *30*(10), 890–904.