**Review Article**

**Plant defense mechanisms against plant parasitic nematodes**

**Abstract**

Plant-parasitic nematodes (PPNs) feed on plant tissues and cause yield losses in agricultural crops. PPNs enter the host plant by penetrating host cells and suppress plant defense response. The defense mechanisms in plant may involve a physical barrier, synthesis of anti-nematode compounds, and transcriptional regulation of defense-related genes. Through strengthening their cell walls plants make physical barriers to PPNs attack. PPN attack produces reactive oxygen species (ROS), anti-nematode compounds or secondary metabolites which are lethal to PPNs. Moreover, some metabolites impede hatching; reduce movement or host searching ability; restrict completion of life-cycle. This review highlights the recent findings of plant defense mechanism against PPNs which will be helpful in developing innovative nematode management strategies in Agriculture.

*Key words:* Plant-parasitic nematodes, Plant defense system, Reactive oxygen species, Plant defense-related genes, Plant anti-nematode compounds, Plant physical barrier, PRRs, MAMPs or PAMPs

1. **INTRODUCTION**

Plant-parasitic nematodes (PPNs) invasion and feeding causes plant damage leading to a considerable yield loss in agricultural crop throughout the world. There are about 4100 nematode species that parasitize hundreds of crop plants and cause significant damage. They are widely distributed and highly adaptable parasite (Jones et al,2013); causing estimated global economic losses of USD 173 billion annually (Elling, 2013). According to their mode of parasitism, PPNs can be classified as endoparasitic, semi-endoparasitic and ectoparasitic (Palomares-Rius et al. 2017). *Trichodorus* spp., Longidorusspp., and  *Xiphinema*spp. are ectoparasitic nematodes, they penetrate their stylet for feeding and remain outside the plant, whereas endoparasitic nematodes , both migratory (*Pratylenchus* spp., and *Anguina* spp.) and sedentary enter the host plant and feed on the plant tissue. Sedentary endoparasites nematodes like root-knot nematodes (RKN), *Meloidogyne* spp., and the cyst nematodes (CN) *Globodera*spp., and *Heterodera*spp., create their feeding sites near to vascular region and feed on the cell cytoplasm. Successful host-parasite interaction needs entering nematodes inside the host plant by invading physical barriers. To aid penetration, nematode juveniles thrusting their stylet and breakdown of the cell wall by various enzymes. A balance is maintained between plants and PPNs, including various signal exchanges (Siddique et al., 2022). As the plants are stationary, therefore they utilize a variety of host defenses concerning both structural components as well as chemical compounds. Both constitutive or pre-formed and inducible defense responses occur after invasion of nematodes. Signaling of these actions is essential for showing defense responses against nematode attack and their spread. This review summarizes the recent findings on how plants recognize PPNs and defend themselves in reaction for penetration and feeding by PPNs. Knowledge about the plant defense mechanism against PPN attack is of crucial importance in developing successful control strategies (Giebel, 2003; Abd-Elgawad, 2022).

1. **PLANTS DEFEND THEMSELVES FROM NEMATODE INFECTION BY SEVERAL WAYS**

Plants adapt their defense strategies to defend nematode attack, while nematodes frequently try to avoid plant defenses. Chemical signals are generated and transmitted in different parts of plant to defend against nematodes (Kaplan & Keen,1980; Gillet et al,2017; Zou et al.2024).

**2.1. Ability of the host plants to identify the invading pathogen:** Host plants are able to sense, recognize and respond accordingly to the invading pathogen (Desmedt et al., 2020). In order to identify the invading pathogens there are two types of recognition system at the molecular level in host plants. These include pattern recognition receptors (PRR) or specified protein receptors and microbial or pathogen-associated molecular patterns (MAMPs or PAMPs) (Chen et al.2024). PAMPs are often present at low concentrations and induce specific defense responses (Manosalva et al.2015). PPN infection may cause damage to the plant tissues which can activate wounding-related plant defense responses through release of damage associated molecular patterns (DAMPs) (Siddique et al.2022). Mendy et al., (2017) demonstrated that J2 of *H. schachtii* also induce general PAMP recognition reactions. Release of PAMPs or DAMPs, after that activation of complex signaling pathways commences pattern-triggered immunity (PTI) against an invading pathogen (Przybylska and Obrępalska-Stęplowska, 2020). Sato et al., (2019) described that Ascarosides (evolutionary conserved group of pheromones derivatives of dideoxysugar ascarylose) and oligogalacturonides released by different PPNs are nematode-associated molecular patterns (NAMPs) are detected by the host plant. De Lorenzo and Cervone, (2022) reported Ascaroside (Asc#18) as a NAMP of *Heterodera glycines* .The MAMP/ NAMPs activate a series of immune responses (PTI) such as the production of reactive oxygen species (ROS) and secondary metabolites, cell death around the PPN-migratory tract, reinforcement of cell walls and activation of the salicylic acid (SA) and jasmonic acid (JA) signaling pathways (Manosalva et al. 2015; Desmedt et al., 2020; Yuan et al., 2021). Nematodes can manipulate plant anatomy and physiology through the secretion of effector molecules that impede plant defenses and promote nematode feeding site establishment. Thus, plants activate their immune response by recognizing the nematode invasion leading to effector triggered immunity (ETI) (Diaz-Granados et al., 2016; Khan and Khan, 2021). Both the ETI and PTI are activated by the extracellular receptor-like kinases/proteins comprising nucleotide-binding domain leucine-rich repeat proteins (NB-LRRs) and/or cytoplasmic kinase (Teixeira et al., 2016; Meresa et al.2024 ; Zhiqi et al.,2025). For example, host plants recognize effectors from RKNs and CNs through NB-LRR receptors (Jones and Dangl 2006;Diaz-Granados et al., 2016). Gr-Vap1 and GrEXPB2 are effector allergen-like protein from *Globodera rostochiensis* and 10A06 from *Heterodera glycines* (Lee et al.,2018; Lozano-Torres et al.2014; De Lorenzo and Cervone, 2022) that can activate ETI in plants.

**2.2.Cell wall as physical barriers in plant:**

A plant cell wall is consisting of various polysaccharides, proteins, and polyphenols that may be change during growth and development in different tissues. The cell wall acts as a physical barrier and can detect any instability caused by biotic and abiotic stresses through specific immune signaling pathways (Swaminathan et al.,2022). Structure and composition of cell wall, waxes or hairs are physical barriers in plant (Malinovsky et al.,2014; Molina et al.2024). Through the process of callose deposition and lignification, plants strengthen their cell walls to make it difficult for nematode to penetrate and feeding. Biosynthesis and deposition of lignin in plant tissues increases its resistance to degradation by enzymes ([Menden et al., 2007](https://www.scielo.org.mx/scielo.php?pid=S0185-33092016000100043&script=sci_arttext&tlng=en#B36)), reduces the transmission of toxins secreted by the pathogen and of nutrients from the host to the pathogen, and also a cause for the production of toxic precursors and free radicals. In peripheral tissues lignin are syntheses through either mechanical wounding, or infection by pests and pathogens .Similarly, plants may deposit phenolic substances that are toxic substances formed in cell walls or epidermal tissues ([Nicholson and Hammerschmit, 1992](https://www.scielo.org.mx/scielo.php?pid=S0185-33092016000100043&script=sci_arttext&tlng=en#B38)). Certain plants have glandular trichomes that secrete compounds, act as repellents or even trap nematodes. Suberin and lignin-based Casparian strips have been observed in roots during RKN infection. Thus, different types of cell wall compositions form a part of inducible defense mechanism in infested plant (Holbein et al. 2019).

**2.3. Reactive oxygen species:** Reactive oxygen species (ROS) are important regulating factors in metabolic activity pathways in plants for the growth and development as well as disease resistance (Zheng et al.,2025). The aerobic respiration of many organisms including higher plants is strongly inhibited by negative ions such as cyanide, azide or sometimes CO. The inhibitory effect of these ions affects the electron transport system. The pathway allows the organism to continue cellular respiration in the presence of cyanide where electrons flow through an alternative oxidase. This type of respiration is said to be cyanide resistant respiration (Feng et al.2013). During the early stages of nematode invasion, the plant cells generates an unfavourable oxidative environment by formation of hydrogen peroxide with superoxides as intermediate and activate local and systemic defense responses (Torres, 2010; Melillo et al., 2011; Lin et al.,2016). The rapid release of superoxide and hydrogen peroxide, is a key mechanism to damage nematode tissues and inhibit their development (Goverse and Smant, 2014). The early detection response to nematode attack aims to detain nematode to their entry sites through on-site programmed cell death, known as hypersensitivity (HR) (Balint-Kurti, 2019). Marinho et al.,(2014) observed that HR is mostly toxic for microbial invaders, as H2O2 reaches an estimated concentration of 5–10 mM. HR cells shows the symptoms of an accumulation of cytoplasm around the invading pathogen, decrease in flaccid membrane potential, loss of electrolytes from cells, or loss of plasmolysing ability of cells (Morel & Dangl, 1997). Some of the tomato, cowpea, potato and pea cultivars shows hypersensitively to *M.incognita*, *G.rostochiensis* and *H.goettingiana* infection by showing the symptoms of extremely vacuolated cytoplasmic contents of syncytial component with a thin layer around cell walls (Leone et al., 2001; Oliveira et al., 2012; Price et al.2021). However, Siddique et al. (2014) observed a promoting role by ROS (H2O2) in cyst nematodes infection. In order to neutralize the excess ROS, plants start initiating antioxidant defense mechanisms in the later stages of infection (Goverse and Smant, 2014; Holbein et al., 2016). The toxic effect of superoxides is diluted by the action of superoxide dismutase (SOD) making disproportionate to H2O2 and O2 . H2O2 is further diluted by catalase or by peroxidase within the cell (Sies, 2014). Antioxidant defense mechanism enables the expression of some other genes encoding ROS-scavenging enzymes (thioredoxins, glutathione peroxidase, xanthineoxidases, oxalate oxidases, amine oxidases) (Delaunois et al., 2014; Iberkleid et al., 2015; Alhoraibi et al.,2019). The 10A06-homologue effector secreted by *H. schachtii* interacts with *Arabidopsis* spermidine synthase 2(SPDS2), which increases the activity of polyamine oxidase (PAO) and consequently enhances the antioxidant pathway in host plants (Hewezi et al., 2010).Strong oxidative bursts have been identified through up-regulation of several peroxidases in early responses of resistant wheat cultivars against *Heterodera avenae*, (Kong et al., 2015). Non-expressor of pathogenesis-related 1(NPR1), a key regulator is associated with a decrease in the number of galls and egg masses in response to *M. incognita* infection (Priya et al., 2011; Siddique et al., 2014). In *Arabidopsis*, ten homologs of respiratory burst oxidase homologs (RBOHs) are prominent during nematode invasion (Mittler et al., 2011). Vicente et al., (2015) observed a correlation between nematode virulence and resistance to oxidative stress in pinewood nematode, *Bursaphelenchus xylophilus*. However, Das et al., (2010) observed that RKN infection is not sufficient to trigger ROS in resistant cowpea and leads to a delayed defense response. Nitric oxide (NO), also another ROS affecting nematode motility, feeding, and development plays a role in plant defense (Khan et al.2023). NO can control cell death in plants by S-nitrosylating nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (Yun et al.,2011). Moreover, ROS involved in reinforcing plant cell walls and inducing callus deposition to limit pathogen entry. In resistant tomato roots infected with *M.incognita,* found to increase the callus deposition in cells surrounding the necrotic areas (Camejo et al.,2016).

**2.4.** **Transcriptional regulation of defense-related genes**: In plants the defense related genes are activated by transcription factors (regulatory proteins) in response to stress (Ling et al. 2017). The regulatory proteins from different families such as WRKY, MYB, AP2 and Bzip are induced in response to various biotic and abiotic stresses (Jiang et al.,2017). These regulatory proteins bind to the promoters of plant genes for establishment of nematode feeding sites PPNs or bind to the suppressors of host genes involved in resistance responses. In case of ETI, pathogen effectors are recognized by plant resistance proteins known as R proteins (Andersen et al., 2018). Generally R proteins induce a rapid and powerful immune response leading to a hypersensitive response. After J2 invasion, the expression of various genes with many different functions is induced, which is associated with defense responses (Gheysen and Fenoll, 2002). In an incompatible host-parasite interaction the single dominant resistance genes from plants interact exclusively with analogous virulence protein (Avr proteins) in the nematode (Ali et al.2017). Kaloshian and Teixeira (2019) identified nucleotide-binding LRR (NB-LRR) proteins that induce resistance to nematodes. Postnikova et al. (2015) found that the genes encoding LRR and TIR-NBS class proteins were up-regulated in a resistant variety of alfalfa against *M. incognita*. Similarly, cyst nematode (*Globodera pallida*) secreted effector like SP1a and RYanodine receptor (SPRY) domain (SPRYSEC) proteins Gp RBP-1, that triggers with Ran GTPase-activating protein 2 (RanGAP2) and GPA2 leading to cell death in potato (Sacco et al., 2009). R-genes Ma, CaMi, Mi-1.2, Mi-9 and Mex-1 gene offer resistance against RKN (Claverie et al., 2011) whereas Hero-A, Grol-4, and Gpa2 gene gives resistance against CN (Sato et al., 2019). Although a single gene in the cluster may offer resistance, multi-gene families are also found in plant R-genes (Friedman & Baker, 2007). Although ten genes are recognized for resistance against *Meloidogyne* spp. in tomatoes, only seven genes (Mi-2, Mi-3, Mi-4, Mi-5, Mi-6, Mi-9, and MI-HT) can activated at high temperatures, e.g., above 32oC (El-Sappah et al.,2019; Devran et al.,2023). Likewise, H1 locus harbors a cluster of intracellular nucleotide-binding (NB)-LRR proteins (NLR) candidate genes which is a single dominant R-gene recognized for resistance against cyst nematode (*Globodera rostochiensis*) (Abd-Elgawad, 2022). In resistant varieties of peanut, the number of induced proteins like putative PR proteins, patatin-like proteins and other stress-related proteins is also found as defense responses (Huang et al.2023). Resistant tomato genes encoding the defense in protein and subtilisin-like protease, through the production of phytoalexins and stress-induced proteolysis (Figueiredo et al.,2018). Thus the plant defense system depends on activating many known and unknown R-genes or Quantitative Trait Loci (QTLs). Resistance genes could be cloned and transferred from some plant cultivars to others. Recognition and cloning of such genes with desirable traits in a plant species can allow the transfer of resistance into other susceptible cultivar(s) of the same species, or even into cultivars of different species. for examples, the Mi-1.2 from tomato against RKN (*Meloidogyne incognita*), Gpa-2 from potato against potato CN (*Globodera pallida*) and Hero A from tomato against potato CNs (*G. pallida* and *G. rostochiensis*) and Cre loci from *Aegilops* spp. against cereal CN (*H. avenae*) in wheat (Kong et al.2015). Therefore, plant genes responsible for PPN resistance are very useful in lowering PPN population levels and enhancing crop yields. **2.5.Phytohormone biosynthesis and signaling:** Hormones synthesized by plants like jasmonate (JA), salicylic acid (SA), ethylene (ET), and auxin involve in defense strategies against PPNs (Hewezi,2015; Gheysen & Mitchum ,2019; Sikder et al.2021). Salicylic acid (SA) and jasmonic acid (JA) can play an important role in the expression of innate and R-gene-associated defense responses (Zhang et al, 2025). SA regulates many genes encoding pathogenesis-related (PR) proteins, mainly PR1 and PR proteins, while the JA regulates the expression of genes encoding defense in thionin, PR3 and PR8 proteins ([Kumar](https://onlinelibrary.wiley.com/authored-by/Kumar/Paramdeep) et al.,2025). Molinari and Loffredo (2006) speculated that *G. rostochiensis* may trigger early but temporary rise of SA to cause early and profuse necrosis. Nguyen et al. (2016) did not find increased susceptibility to *H. schachtii* in *Arabidopsis* SA signaling mutants. Wang et al.(2018) demonstrated that temporary expression of chorismate mutase by *M. incognita* in *Nicotiana benthamiana* cause a lower SA levels. Over expression of chorismate mutase or an isochorismatase by *Hirschmanniella oryzae* in rice also enhances susceptibility to this nematode (Bauters et al.,2020). Kempster et al., (2001) demonstrated resistance in tomato against *M. incognita* (Molinari, 2015), *M. javanica* (Moslemi et al.,2016) and *M. chitwoodi* (Vieira dos Santos et al., 2013), and also in white clover against *Heterodera trifolii* through application of SA.

JA may play a major role in plant-nematode interaction in the roots of both monocotyledons and dicotyledonous plants (Mendy et al. 2017). JA acts as a signal molecule, triggering the production of plant defenses like proteinase inhibitors, terpenoids, and oxylipins. These plant metabolites contribute to the overall defense against nematodes (Gheysen & Mitchum, 2019). Przybylska & Spychalski, (2021) suggested the role of JA- and SA-mediated pathways in maize infected by *M. arenaria* that showed changes in the expression levels of genes encoding the PR3, PR4 and PR5 proteins.Several workers have demonstrated that JA signaling is implicated in plant defense against *M. incognita* , *M. chitwoodi* ,*M. graminicola* ,*H. glycines* and *H. schachtii* (Cooper et al.,2005;Kammerhofer et al., 2015; Zhou et al., 2015;Lin et al., 2017). Zhao et al., (2015) observed that highly susceptible tomato plants to *M. incognita* over expressing miR319and to *M.javanica* over expressing MjFAR showed lower JA levels. This observation is associated with lower expression of the JA-responsive proteinase inhibitor 2; however, some genes in the JA pathway are expressed at higher levels in these roots (Iberkleid et al., 2015).

Ethylene (ET) is a gaseous hormone interacting synergistically with auxin to control root cell development and root hair growth (Strader et al., 2010 ;Kyndt et al.2016)). Arabidopsis roots exposed to an ethylene (ET)-synthesis inhibitor attracted more *M.hapla* J2 than in control roots and that ET-overproducing mutants were less attractive to the zone of elongation of roots (Fudali et al.,2013). Nahar et al., (2011) observed that ET and JA synergistically induced a systemic defense response against *M. graminicola* in rice.

Strigolactones (SL) are carotenoid-derived hormones that regulate various plant developmental and adaptation processes as well as many biotic and abiotic stress responses (Kun-Peng et al.2018). In tomato, silencing of SL biosynthesis genes resulted in higher infection rates of *M.incognita* (Xu et al., 2019).

Gibberellic acid (GA) is well known hormone that stimulate plant growth by the degradation of DELLAs, a class of growth-repressing nuclear proteins. Studies in *Arabidopsis* revealed that GA decreases JA action and increases SA signaling and perception (Navarro et al., 2008; De Vleesschauwer et al., 2016).

Abscisic acid (ABA) may intervene both positive and negative defense responses by cross-talking with JA, SA and ET signaling pathways (Yang et al.2019). Nahar et al.(2012) speculated that the rice plant response against root lesion nematodes depends on the balance between ABA,SA,JA and ET.

**2.6. Production of anti-nematode phytochemicals:** Both constitutive and inducible secondary metabolites are synthesized in plants against nematodes which is known as ANPs. Plants obtain constitutive forms of metabolites that do not require any pest or pathogen invasion. Some of the secondary metabolites, like terpenoids, saponins, benzoxazinoids, organosulfur compounds, alkaloids, chlorogenic acid, and glucosinolates have been shown to inhibit nematode activity (Desmedt et al.2020). In *Brassica* , glucosinolates offer resistance to plant-parasitic nematodes (Potter et al., 2000). In Marigold roots, α-terthienyl compounds exhibits high nematicidal activity against several plant parasitic nematodes (Hamaguchi et al.2019). Glyceollin and terpenoid aldehyde inhibits oxygen uptake and function as nematistatic to the J2 of *M.incognita* (Veech & McClure,1977). Resistant cultivar ‘Centinniel’ of soybean showed an increase level of glyceollin1 nearby the head of the nematode, *H.glycines* race 1 (Huang & Barker, 1991). These secondary metabolites can affect the biology of PPNs in the root system (Lee et al., 2017). Another secondary metabolite phytoalexins are synthesized and accumulated in plants. These are low molecular weight compounds accumulated with the formation of root border cells when the root tips are exposed to PPNs elicitor (esophageal gland secretion, excretory products and various surface compounds of nematodes) (Desmedt et al.,2020). The elicitor may lead to *de novo* synthesis of enzymes like Phenylalanine ammonia lyase (PAL) and chalcone synthatase ([Veech](https://pubmed.ncbi.nlm.nih.gov/?term=%22Veech%20JA%22%5BAuthor%5D) 1982). Ho¨lscher et al.,(2014) revealed synthesize and accumulation of phenalenone-type phytoalexins after root infection in banana by *Radopholus similis*. Over expression of the *Arabidopsis* phytoalexin-deficient 4 gene (AtPAD4), a lipase-like protein activated by salicylic acid (SA) and phytoalexins, enhances resistance in soybean roots in response to *Meloidogyne incognita* and *H.glycines* (Youssef et al., 2013).

Phytoecdysteroids may also provide an important plant defense against nematodes (Schmelz et al., 2000). Plant defense have been observed through 20-hydroxyecdysone (20E), a molting hormone-ecdysteroid inducible in spinach on *Pratylenchus, Heterodera* and *Meloidogyne* spp. Since spinach also contains a minor phytoecdysteroid, polypodine B, was applied exogenously Abnormal molting, immobility, reduced invasion, impaired development, and death in nematodes, *H. avenae* was observed after exposure to 20E either directly at concentration above 4.2 × 10-7 M or in plants (Soriano et al.2004).

**3. CONCLUSION AND FUTURE PERSPECTIVES**

With the advancement of high throughput molecular technologies especially the use of untargeted metabolomics techniques based on nuclear magnetic resonance (NMR) and mass spectrometry (MS), plant defense mechanisms against PPN attacks can be better understandable. Moreover, understanding of the host-parasite interaction and activities like PPN recognition, species-specific responses and immune signaling networks in plant system will benefit the development of nematode disease control strategies. Strategies should be taken into consideration to identify anti-nematode metabolites and their use in agriculture and crop protection.

**DISCLAIMER (ARTIFICIAL INTELLIGENCE)**

Author hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of this manuscript.

**REFERENCES**

Abd-Elgawad, M.M.M. (2022). Understanding molecular Plant-Nematode interactions to develop alternative approaches for nematode control. Plants 11, 2141. <https://doi.org/10.3390/plants11162141>.

Alhoraibi, H., Bigeard, J., Rayapuram, N., Colcombet, J.,& Hirt, H. (2019). Plant Immunity: The MTI-ETI Model and Beyond. Current Issues in Molecular Biology. 30:39-58. doi: 10.21775/cimb.030.039.

Ali, M.A., Azeem, F., Abbas, A., Joyia, F.A., Li, H.& Dababat, A.A. (2017).Transgenic strategies for enhancement of nematode resistance in plants. Frontiers in Plant Science. 8:750.doi: 10.3389/fpls.2017.00750

Andersen, E.J., Ali, S., Byamukama, E., Yen, Y., & Nepal, M.P. (2018). Disease resistance mechanisms in plants. Genes. 9:339. 10.3390/genes9070339

Balint-Kurti, P. (2019). The plant hypersensitive response: concepts, control and consequences. Molecular Plant Pathology. 20(8):1163-1178. doi: 10.1111/mpp.12821.

Bauters, L., Kyndt, T., De Meyer, T., Morreel, K., Boerjan, W., Lefevere, H.,& Gheysen, G. (2020) .Chorismate mutase and isochorismatase, two potential effectors of the migratory nematode *Hirschmanniella oryzae*, increase host susceptibility by manipulating secondary metabolite content of rice. Molecular Plant Pathology. 21(12):1634-1646. doi: 10.1111/mpp.13003.

Camejo, D., Guzman-Cedeno, A., & Moreno, A.(2016). Reactive oxygen species, essential molecules, during plant-pathogen interactions. Plant Physiology and Biochemistry.103:10-23. DOI: [10.1016/j.plaphy.2016.02.035](https://doi.org/10.1016/j.plaphy.2016.02.035).

Chen, X., Li, F., Wang, D., & Cai, L. (2024). Insights into the plant response to nematode invasion and modulation of host defense by plant parasitic nematode. Frontiers in Microbiology.15:1482789. doi: 10.3389/fmicb.2024.1482789.

Claverie, M., Dirlewanger, E., Bosselut, N., Van Ghelder, C., Voisin, R., Kleinhentz, M., et al. (2011). The Ma gene for complete-spectrum resistance to *Meloidogyne* species in *Prunus* is a TNL with a huge repeated C-terminal post-LRR region. Plant Physiology.156, 779–792. doi: 10.1104/pp.111.176230.

Cooper,W.R., Jia, L.& Goggin, L. (2005). Effects of jasmonate-induced defenses on root-knot nematode infection of resistant and susceptible tomato cultivars. Journal of Chemical Ecology. 31: 1953–1967.  DOI: [10.1007/s10886-005-6070-y](https://doi.org/10.1007/s10886-005-6070-y).

Das, S., DeMason, D.A., Ehlers, J.D., Close, T.J. & Roberts, P.A. (2008). Histological characterization of root-knot nematode resistance in cowpea and its relation to reactive oxygen species modulation. Journal of Experimental Botany. 59(6): 1305–1313. <https://doi.org/10.1093/jxb/ern036>

De Lorenzo, G., & Cervone, F. (2022). Plant immunity by damage-associated molecular patterns (DAMPs). Essays in Biochemistry. 66, 459-469. doi: 10.1042/EBC20210087.

De Vleesschauwer, D., Seifi, H.S., Filipe, O., Haeck, A., Huu, S.N., Demeestere, K.,& Höfte, M. (2016). The DELLA Protein SLR1 Integrates and amplifies Salicylic Acid- and Jasmonic Acid-dependent innate immunity in Rice. Plant Physiology. 170(3):1831-47. doi: 10.1104/pp.15.01515.

Delaunois, B., Jeandet, P., Clément, C., Baillieu, F., Dorey, S. & Cordelier, S.(2014). Uncovering plant-pathogen crosstalk through apoplastic proteomic studies. Frontiers in Plant Science.5,249. doi: 10.3389/fpls.2014.00249.

Desmedt, W., Mangelinckx, S., Kyndt, T., & Vanholme, B.(2020). A phytochemical perspective on plant defense against nematodes. Frontiers in Plant Science 11:602079.doi: 10.3389/fpls.2020.602079.

Devran, Z., Özalp, T., Studholme, D.J.,& Tör, M. (2023). Mapping of the gene in tomato conferring resistance to root-knot nematodes at high soil temperature. Frontiers in Plant Science. 14, 1267399. doi: 10.3389/fpls.2023

Diaz-Granados, A., Petrescu, A.J., Goverse, A., & Smant, G. (2016). SPRYSEC effectors: A versatile protein-binding platform to disrupt plant innate immunity. Frontiers in Plant Science 7:1575. <https://doi.org/10.3389/fpls.2016.01575>

Elling, A.A. (2013). Major emerging problems with minor *Meloidogyne* species. Phytopathology. 103(11), 1092–1102. https://doi.org/10.1094/PHYTO-01-13-0019-RVW.

El-Sappah, A.H., M, M.I, El-Awady, H., Yan, S., Qi, S., Liu, J., Cheng, G.T., & Liang, Y. (2019).Tomato natural resistance genes in controlling the root-knot nematode. Genes (Basel). 10(11):925. doi: 10.3390/genes10110925.

Feng, H.Q., Sun, K., Wei, Y. ,Wang, R.F., Jia, L.Y., Zhang, J.P. et al.(2013). Role of cyanide-resistant respiration during light-induced attraction of predators to herbivore-infested leaves. Photosynthetica 51, 583–592 https://doi.org/10.1007/s11099-013-0057-7

Figueiredo, J., Sousa Silva, M., & Figueiredo, A. (2018). Subtilisin-like proteases in plant defence: the past, the present and beyond. Molecular Plant Pathology. 19(4):1017-1028. doi: 10.1111/mpp.12567.

Friedman, A.R., & Baker, B.J. (2007). The evolution of resistance genes in multi-protein plant resistance systems. Current Opinion in Genetics & Development. 17(6):493-499, <https://doi.org/10.1016/j.gde.2007.08.014>

Fudali, S.L.,Wang, C., & Williamson, V.M.(2013). Ethylene signaling pathway modulates attractiveness of host roots to the root-knot nematode *Meloidogyne hapla*. Molecular Plant-Microbe Interactions. 26(1):75-86. DOI: [10.1094/MPMI-05-12-0107-R](https://doi.org/10.1094/mpmi-05-12-0107-r).

Gheysen, G., & Fenoll, C. (2002) Gene expression in nematode feeding sites. Annual Review of Phytopathology. 40:191-219. doi: 10.1146/annurev.phyto.40.121201.093719.

Gheysen, G., & Mitchum, M.G. (2019). Phytoparasitic nematode control of plant hormone pathways. Plant Physiology. 179: 1212–1226. [www.plantphysiol.org/](http://www.plantphysiol.org/) cgi/doi/10.1104/pp.18.01067.

Giebel, J. (2003). Mechanism of resistance to plant nematodes. Annual Review of Phytopathology. 20. 257-279. 10.1146/annurev.py.20.090182.001353.

Gillet,F.X., Bournaud, C., de Souza Junior , J.D.A., & Grossi-de-Sa, M.F.(2017). Plant-parasitic nematodes: towards understanding molecular players in stress responses. Annals of Botany. 119: 775-789. doi:10.1093/aob/mcw260.

Goverse, A., & Smant, G. (2014). The activation and suppression of plant innate immunity by parasitic nematodes. Annual Review of Phytopathology. 52: 243–265. DOI: [10.1146/annurev-phyto-102313-050118](https://doi.org/10.1146/annurev-phyto-102313-050118).

Hamaguchi, T., Sato, K., Vicente, C.S.L.,& Hasegawa, K. (2019). Nematicidal actions of the marigold exudate α-terthienyl: oxidative stress-inducing compound penetrates nematode hypodermis. Biology Open. 8(4):bio038646. doi: 10.1242/bio.038646.

Hewezi, T. (2015). Cellular signaling pathways and posttranslational modifications mediated by nematode effector proteins. Plant Physiology. 169, 1018–1026. doi: 10.1104/pp.15.00923.

Hewezi, T., Peter, J. H., Tom, R. M., Richard, S. H., Melissa, G. M, Eric L. D., et al.(2010). *Arabidopsis* Spermidine synthase is targeted by an effector protein of the Cyst Nematode *Heterodera schachtii*.  Plant Physiology. 152(2): 968-984. <https://doi.org/10.1104/pp.109.150557>

Holbein, J., Franke, R.B. , Marhavy, P. , Fujita, S., Gorecka, M., Sobczak, M., et al.(2019). Root endodermal barrier system contributes to defence against plant-parasitic cyst and root-knot nematodes. The Plant Journal . 100, 221–236. doi: 10.1111/tpj.14459.

Holbein, J., Grundler,F.M.W., & Siddique, S. (2016). Plant basal resistance to nematodes: an update. Journal of Experimental Botany. 67(7): 2049–2061. <https://doi.org/10.1093/jxb/erw005>

Hölscher, D., Dhakshinamoorthy, S., Alexandrov, T., Becker, M., Bretschneider, T., Buerkert, A., et al. (2014). Phenalenone-type phytoalexins mediate resistance of banana plants (Musa spp.) to the burrowing nematode *Radopholus similis*. Proceedings of the National Academy of Sciences of the United States of America. 111 (1): 105-110. doi: 10.1073/pnas.1314168110

Huang, J. S., & Barker, K. R. (1991). Glyceollin I in soybean-cyst nematode interactions: spatial and temporal distribution in roots of resistant and susceptible soybeans. Plant Physiology. 96 (4), 1302–1307. doi: 10.1104/ pp.96.4.1302

Huang, R., Li, H., Gao, C., Yu, W., & Zhang, S.(2023). Advances in omics research onpeanut response to biotic stresses. Frontiers in Plant Science. 14:1101994.doi: 10.3389/fpls.2023.1101994

Iberkleid, I., Sela, N., & Brown, M. S. (2015). *Meloidogyne javanica* fatty acid and retinol-binding protein (Mj-FAR-1) regulates expression of lipid, cell wall, stress and phenylpropanoid-related genes during nematode infection of tomato. BMC Genomics. 16: 272. doi:10.1186/s12864-015-1426-3.

Jiang, J., Ma, S., Ye, N., Jiang, M., Cao, J.,& Zhang, J.(2017). WRKY tranascription factors in plant responses to stresses. Journal of Integrative Plant Biology.59:86-101.doi: 10.1111/jipb.12513.

Jones, J. & Dangl, J. (2006). The plant immune system. Nature. 444: 323–329 <https://doi.org/10.1038/nature05286>.

Jones, J.T., Haegeman, A., Danchin, E.G.J., Gaur, H.S., Helder, J., Jones, M.G.K., et al. (2013). Top 10 plant-parasitic nematodes in molecular plant pathology. Molecular Plant Pathology. 14: 946-961. <https://doi.org/10.1111/mpp.12057>.

Kaloshian, I., & Teixeira, M. (2019). Advances in plant- nematode interactions with emphasis on the notorious nematode genus *Meloidogyne*. Phytopathology. 109:1988-1996. doi: 10.1094/PHYTO-05-19-0163-IA.

Kammerhofer, N., Radakovic, Z., Regis, J. M., Dobrev, P., Vankova, R., Grundler, F. M., et al. (2015). Role of stress-related hormones in plant defence during early infection of the cyst nematode *Heterodera schachtii* in Arabidopsis. New Phytologist. 207:778-789. doi: 10.1111/nph.13395

Kaplan,D.T. & Keen,N.T.(1980). Mechanisms conferring plant incompatibility to nematodes. Revue Nématol. 3 (1) : 123-134 .

Kempster, V.N., Davies, K.A., & Scott, E.S. (2001). Chemical and biological induction of resistance to the clover cyst nematode (*Heterodera trifolii*) in white clover (*Trifolium repens*). Nematology. 3(1), 35-43. <https://doi.org/10.1163/156854101300106874>

Khan, M., & Khan, A.U. (2021).Plant parasitic nematodes effectors and their crosstalk with defense response of host plants: A battle underground, Rhizosphere. 17,100288, https:// doi.org/ 10.1016/j.rhisph.2020.100288.

Khan, M., Ali, S., Al Azzawi, T.N.I., & Yun, B.W. (2023). Nitric Oxide acts as a key signaling molecule in plant development under stressful conditions. International Journal of Molecular Science.  24, 4782. <https://doi.org/10.3390/ijms24054782>

Kong, L., Wu, D., Huang, W., Peng, H., Wang, G., Cui, J., et al.(2015). Large-scale identification of wheat genes resistant to cereal cyst nematode *Heterodera avenae* using comparative transcriptomic analysis. BMC Genomics. 16:801.DOI 10.1186/s12864-015-2037-8.

[Kumar](https://onlinelibrary.wiley.com/authored-by/Kumar/Paramdeep), P., [Pandey](https://onlinelibrary.wiley.com/authored-by/Pandey/Saurabh), S., &  [Pati](https://onlinelibrary.wiley.com/authored-by/Pati/Pratap%2BKumar), P.K.(2025). Interaction between pathogenesis-related (PR) proteins and phytohormone signaling pathways in conferring disease tolerance in plants. Physiologia Plantarum.177(2), e70174, <https://doi.org/10.1111/ppl.70174>.

Kun-Peng, J, Lina, B., & Al-Babili, S. 2018. From carotenoids to strigolactones, Journal of Experimental Botany. 69(9):2189-2204. <https://doi.org/10.1093/jxb/erx476>.

Kyndt, T., Goverse, A., Haegeman, A., Warmerdam,S., Wanjau, C., Jahani, M., et al.2016. Redirection of auxin flow in Arabidopsis thaliana roots after infection by root-knot nematodes. Journal of Experimental Botany. 67: 4559-4570. DOI: [10.1093/jxb/erw230](https://doi.org/10.1093/jxb/erw230).

Lee, H. A., Lee, H. Y., Seo, E., Lee, J., Kim, S. B., Oh, S., et al. (2017). Current understandings of plant nonhost resistance. Molecular Plant-Microbe Interactions. 30, 5–15. doi: 10.1094/MPMI-10-16-0213-CR.

Lee, M.W.,Huffaker, A., Crippen, D., Robbins, R.T., & Goggin, F.L.(2018). Plant elicitor peptides promote plant defences against nematodes in soybean. Molecular Plant Pathology. 19:858-69. <https://doi.org/10.1111/mpp.12570>

Leone,A., Melillo,M.T.,& Bleve-Zacheo,T. (2001).Lipoxygenase in pea roots subjected to biotic stress. Plant Science. 161(4): 703-717, https://doi.org/10.1016/S0168-9452(01)00458-7

Lin, B., Zhuo, K., Chen, S., Hu, L., Sun, L.,Wang, X., & Zhang, L.H. (2016). A novel nematode effector suppresses plant immunity by activating host reactive oxygen species-scavenging system. New Phytologist. 209:1159-73. <https://doi.org/10.1111/nph.13701>.

Lin, J., Wang, D., Chen, X., Köllner, T.G., Mazarei, M., Guo, H., et al. (2017). An (E,E)-α-farnesene synthase gene of soybean has a role in defence against nematodes and is involved in synthesizing insect-induced volatiles. Plant Biotechnology Journal. 15(4):510-519. doi: 10.1111/pbi.12649.

Ling, J., Mao, Z., Zhai, M., [Zeng](https://www.nature.com/articles/s41598-017-03563-6#auth-Feng-Zeng-Aff1), F.[, & Yang](https://www.nature.com/articles/s41598-017-03563-6#auth-Yuhong-Yang-Aff1),  Y, B. (2017).Transcriptome profiling of *Cucumis metuliferus* infected by *Meloidogyne incognita* provides new insights into putative defense regulatory network in Cucurbitaceae. Scientific Reports. 7:3544https://doi.org/10.1038/s41598-017-03563-6

Lozano-Torres, J.L, Wilbers, R.H., Warmerdam, S., Finkers-Tomczak, A., Diaz-Granados, A., van Schaik, C.C., et al. (2014). Apoplastic venom allergen-like proteins of cyst nematodes modulate the activation of basal plant innate immunity by cell surface receptors. PLOS Pathogens.10(12):e1004569. doi: 10.1371/journal.ppat.1004569.

Malinovsky, F.G., Fangel, J.U., & Willats, W.G. (2014). The role of the cell wall in plant immunity. Frontiers in Plant Science. 6(5):178. doi: 10.3389/fpls.2014.00178.

Manosalva, P., Manohar, M., von Reuss, S.,  [Chen](https://www.nature.com/articles/ncomms8795#auth-Shiyan-Chen-Aff3), S.,   [Koch](https://www.nature.com/articles/ncomms8795#auth-Aline-Koch-Aff4), A., [Kaplan](https://www.nature.com/articles/ncomms8795#auth-Fatma-Kaplan-Aff5), F.,  et al. (2015).Conserved nematode signalling molecules elicit plant defenses and pathogen resistance. Nature Communications. 6, 7795. <https://doi.org/10.1038/ncomms8795>.

Marinho, H.S., Real, C., Cyrne, L., Soares, H., & Antunes, F. (2014). Hydrogen peroxide sensing, signaling and regulation of transcription factors. Redox Biology. 2:535-62. doi: 10.1016/j.redox.2014.02.006.

Melillo, M.T., Leonetti, P., Leone, A., Veronico, P., & Bleve-Zacheo, T. (2011). ROS and NO production in compatible and incompatible tomato–*Meloidogyne incognita* interactions. European Journal of Plant Pathology. 130: 489–502. https://doi.org/10.1007/s10658-011-9768-4

Menden, B., Kohlhoff, M., & Moerschbacher, B.M. (2007). Wheat cells accumulate a syringyl-rich lignin during the hypersensitive resistance response. Phytochemistry. 68:513–520. doi: 10.1016/j.phytochem.2006.11.011

Mendy, B., Wang’ombe, M.W., Radakovic, Z.S., Holbein, J., Ilyas, M., Chopra, D., et al. (2017) Arabidopsis leucine-rich repeat receptor–like kinase NILR1 is required for induction of innate immunity to parasitic nematodes. PLOS Pathogens. 13(4): e1006284. https://doi.org/10.1371/journal.ppat.1006284

Meresa, B.K., Matthys, J., & Kyndt, T. (2024).Biochemical defence of plants against parasitic nematodes. Plants.; 13(19):2813. <https://doi.org/10.3390/plants13192813>.

Mittler, R., Vanderauwera, S., Suzuki, N., Miller, G.,
Tognetti, V.B., Vandepoele, K. et al. (2011). ROS signaling: the new wave? Trends in Plant Science. 16: 300–309. <https://doi.org/10.1016/j.tplants.2011.03.007>

Molina, A., Jordá, L., Ángel, T. M., Martín-Dacal, M., José Berlanga, D., Fernández-Calvo, P., et al. (2024). Plant cell wall-mediated disease resistance: Current understanding and future perspectives, Molecular Plant. 17(5): 699-724. https:// doi.org/ 10.1016/ j.molp. 2024.04.003.

Molinari, S.(2015). Systemic acquired resistance activation in solanaceous crops as a management strategy against root-knot nematodes. Pest Management Science.72(5):888-896. <https://doi.org/10.1002/ps.4063>.

Molinari, S., &  Loffredo, E. (2006). The role of salicylic acid in defense response of tomato to root-knot nematodes. [Physiological and Molecular Plant Pathology](https://www.sciencedirect.com/journal/physiological-and-molecular-plant-pathology).  [68 (1–3](%2068%20%281%E2%80%933)):  69-78. <https://doi.org/10.1016/j.pmpp.2006.07.001>.

Morel,.J.B. & Dangl, J.L.(1997). The hypersensitive response and the induction of cell death in plants. Cell Death and Differentiation . 4: 671 -683.

Moslemi, F., Fatemy, S., & Bernard, F. (2016). Inhibitory effects of salicylic acid on *Meloidogyne javanica* reproduction in tomato plants. Spanish Journal of Agricultural Research. 14,1, e1001. <http://dx.doi.org/10.5424/sjar/2016141-8706>.

Nahar, K., Kyndt, T., De Vleesschauwer, D., Höfte, M.,& Gheysen, G. 2011. The jasmonate pathway is a key player in systemically induced defense against root knot nematodes in rice. Plant Physiology. 157(1):305-16. doi: 10.1104/pp.111.177576

Nahar, K., Kyndt, T., Nzogela, Y.B.,& Gheysen, G. (2012). Abscisic acid interacts antagonistically with classical defense pathways in rice-migratory nematode interaction. New Phytologist. 196(3):901-913. doi: 10.1111/j.1469-8137.2012.04310.x.

Navarro, L., Bari, R., Achard, P., Lisón, P., Nemri, A., Harberd, N.P., et al.(2008). DELLAs control plant immune responses by modulating the balance of jasmonic acid and salicylic acid signaling. Current Biology. 18(9):650-5. doi: 10.1016/j.cub.2008.03.060.

Nguyen, P.D.T., Pike, S., Wang, J., Poudel, A.N., Heinz, R. , Schultz, J.C et al. (2016). The Arabidopsis immune regulator SRFR1 dampens defences against herbivory by *Spodoptera exigua* and parasitism by *Heterodera schachtii*. Molecular Plant Pathology. 17(4): 588-600 .DOI: 10.1111/mpp.12304.

Nicholson, R.L., & Hammerschmidt, R. (1992). Phenolic compounds and their role in disease resistance. Annual Review of Phytopathology. 30, 369–389. doi: 10.1146/annurev.py.30.090192.002101

Oliveira, J.T.A, Andrade, N.C., Martins-Miranda, A.S., Soares, A.A., Gondim, D.M.F., Araújo-Filho, J.H. et al. (2012). Differential expression of antioxidant enzymes and PR-proteins in compatible and incompatible interactions of cowpea (*Vigna* *unguiculata*) and the root-knot nematode *Meloidogyne incognita*. Plant Physiology and Biochemistry.51:145-152, <https://doi.org/10.1016/j.plaphy.2011.10.008>.

Palomares-Rius, J.E., Escobar, C., Cabrera, J., Vovlas, A. & Castillo, P.(2017) Anatomical alterations in plant tissues induced by plant-parasitic nematodes. Frontiers in Plant Science. 8:1987.doi: 10.3389/fpls.2017.01987

Postnikova, O.A., Hult, M., Shao, J., Skantar, A., & Nemchinov, L.G. (2015). Transcriptome analysis of resistant and susceptible alfalfa cultivars infected with root-knot nematode *Meloidogyne incognita*. Public Library of Science ONE. 10(3): e0123157. <https://doi.org/10.1371/journal.pone.0123157>

Potter, M.J., Vanstone, V.A., Davies, K.A. et al. (2000). Breeding to Increase the Concentration of 2-phenylethyl Glucosinolate in the Roots of *Brassica napus*. Journal of Chemical Ecology. 26: 1811-1820 https://doi.org/10.1023/A:1005588405774

Price, J.A., Coyne, D., Blok, V.C.,& Jones, J.T.(2021).Potato cyst nematodes *Globodera rostochiensis* and *G. pallida*. Molecular Plant Pathology. 22(5):495-507. doi: 10.1111/mpp.13047.

Priya, D.B., Somasekhar, N., Prasad, J., & Kirti, P. (2011). Transgenic tobacco plants constitutively expressing Arabidopsis NPR1 show enhanced resistance to root-knot nematode, *Meloidogyne incognita*. BMC Research Notes. 4:231. doi: 10.1186/1756-0500-4-231.

Przybylska, A. & Obrępalska-Stęplowska, A.(2020). Plant defense responses in monocotyledonous and dicotyledonous host plants during root-knot nematode infection. Plant Soil. 451:239-260. https://doi.org /10.1007/s11104-020-04533-0.

Przybylska, A.,& Spychalski, M. 2021. Changes in the expression level of genes encoding transcription factors and cell wall-related proteins during *Meloidogyne arenaria* infection of maize (*Zea mays*). Molecular Biology Reports. 48(10):6779-6786. doi: 10.1007/s11033-021-06677-3

Sacco, M.A., Koropacka, K., Grenier, E., Jaubert, M.J., Blanchard, A., Goverse, A., et al. (2009). The cyst nematode SPRYSEC protein RBP-1 elicits Gpa2-and RanGAP2-dependent plant cell death. PLOS Pathogens. 5:e1000564. doi: 10.1371/journal.ppat.1000564

Sato, K., Kadota, Y., & Shirasu, K. (2019). Plant immune responses to parasitic nematodes. Frontiers in Plant Science. 10:1165. doi: 10.3389/fpls.2019.01165.

Schmelz, E.A., Grebenok, R.J., Ohnmeiss, T.E. & Bowers, W.S. (2000). Phytoecdysteroid Turnover in Spinach: Long-term Stability Supports a Plant Defense Hypothesis. Journal of Chemical Ecology. 26:2883-2896 [https://doi.org/10.1023/A:1026454213510](https://doi.org/10.1023/A%3A1026454213510).

Siddique, S., Matera, C., Radakovic, Z.S, Shamim, H.M, Gutbrod, P., Rozanska E, et al. (2014). Parasitic worms stimulate host NADPH oxidases to produce reactive oxygen species that limit plant cell death and promote infection. Science Signaling.  8;7(320):ra33. doi: 10.1126/scisignal.2004777.

Siddique,S., Coomer,A., Baum, T., & Williamson,V.M.(2022). Recognition and response in plant-nematode interactions. Annual Review of Phytopathology. 60:143-62. <https://doi.org/10.1146/annurev-phyto-020620-102355>.

Sies, H. (2014). Role of metabolic H2O2 generation: redox signaling and oxidative stress. J Biol Chem. 289(13):8735-41. doi: 10.1074/jbc.R113.544635.

Sikder, M.M., Vestergard,M., Kyndt, T.T., Kudjordjie, E.N., & Nicolaisen,M..(2021). Phytohormones selectively affect plant parasitic nematodes associated with *Arabidopsis* roots. New Phytologist,.232: 1272-1285.doi: 10.1111/nph.17549.

Soriano,I.R., Riley,I.T., Potter, M.J.,& Bowers,W.S. (2004). Phytoecdysteroids: A novel defense against plant-parasitic nematodes. Journal of Chemical Ecology. 30(10):1885-1899.doi:0098-0331/04/1000-1885/0.

Strader, L.C., Chen, G.L.,& Bartel, B.(2010). Ethylene directs auxin to control root cell expansion.The Plant Journal. 64(5): 874-884.doi: 10.1111/j.1365-313x.2010.04373.x

Swaminathan, S., Lionetti,V. & Zabotina, O.A. (2022). Plant cell wall integrity perturbations and priming for defense. Plants. 11, 3539.https://doi.org/10.3390/plants11243539.

Teixeira, M.A., Wei, L., & Kaloshian, I.(2016). Root-knot nematodes induce pattern-triggered immunity in *Arabidopsis thaliana* roots. New Phytologist. :211(1):276-287. <https://doi.org/10.1111/nph.13893>.

Torres, M.A. (2010). ROS in biotic interactions. Physiologia Plantarum. 138(4):414-429. doi:10.1111/j.1399-3054.2009.01326.x

[Veech](https://pubmed.ncbi.nlm.nih.gov/?term=%22Veech%20JA%22%5BAuthor%5D), J.A.(1982). Phytoalexins and their role in the resistance of plants to nematodes. Journal of Nematology. 14(1):2-9. PMCID: PMC2618140

Veech, J.A.,& McClure, M.A. (1977). Terpenoid aldehydes in cotton roots susceptible and resistant to the root-knot nematode, *Meloidogyne incognita*. Journal of Nematology. 9(3):225-9. PMID: 19305600

Vicente, C.S.L., Ikuyo, Y., Mota, M. &  [Hasegawa](https://bmcmicrobiol.biomedcentral.com/articles/10.1186/1471-2180-13-299#auth-Koichi-Hasegawa-Aff1), K.(2013).Pinewood nematode-associated bacteria contribute to oxidative stress resistance of *Bursaphelenchus xylophilus*. BMC Microbiology. 13, 299 <https://doi.org/10.1186/1471-2180-13-299>.

Vieira dos Santos, M.C., Curtis, R.H.C. & Abrantes, I. (2013). Effect of plant elicitors on the reproduction of the root-knot nematode *Meloidogyne chitwoodi* on susceptible hosts. European Journal of Plant Pathology. 136: 193–202 https://doi.org/10.1007/s10658-012-0155-6

[Wang](https://bsppjournals.onlinelibrary.wiley.com/authored-by/Wang/X.), X., [Xue](https://bsppjournals.onlinelibrary.wiley.com/authored-by/Xue/B.), B.,  [Dai](https://bsppjournals.onlinelibrary.wiley.com/authored-by/Dai/J.), J, [X., Qin](https://bsppjournals.onlinelibrary.wiley.com/authored-by/Qin/X.), [L., Liu](https://bsppjournals.onlinelibrary.wiley.com/authored-by/Liu/L.), [Y.,& Chi](https://bsppjournals.onlinelibrary.wiley.com/authored-by/Chi/Y.), [J.T.](https://bsppjournals.onlinelibrary.wiley.com/authored-by/Jones/J.%2BT.) (2018). A novel *Meloidogyne incognita*chorismate mutase effector suppresses plant immunity by manipulating the salicylic acid pathway and functions mainly during the early stages of nematode parasitism.Plant Pathology.67(6):1436-1448. <https://doi.org/10.1111/ppa.12841>.

Xu, X., Fang, P., Zhang, H., Chi, C., Song, L., Xia, X., et al.(.2019). Strigolactones positively regulate defense against root-knot nematodes in tomato. Journal of Experimental Botany 4:1325-1337. <https://doi.org/10.1093/jxb/ery439>

Yang, J., Duan, G., Li, C., Liu, L., Han, G., Zhang, Y.et al.(2019).The crosstalks between jasmonic acid and other plant hormone signaling highlight the involvement of jasmonic acid as a core component in plant response to biotic and abiotic stresses. Frontiers in Plant Science. 10:1349. doi: 10.3389/fpls.2019.01349.

Youssef, R.M., MacDonald, M.H., Brewer, E.P., Bauchan, G.R., Kim, K.H.,& Matthews, B.F. (2013). Ectopic expression of AtPAD4 broadens resistance of soybean to soybean cyst and root-knot nematodes. BMC Plant Biology. 13:67. doi:10.1186/1471-2229-13-67.

Yuan, M., Jiang, Z., Guozhi, Bi., Menghui, L. K.N. ,Wang, Y., Cai, B., et al. (2021). Pattern-recognition receptors are required for NLR-mediated plant immunity. Nature. 592(7852): 105-109. doi:10.1038/s41586-021-03316-6.

Yun, B.W., Feechan, A., Yin, M., Saidi, N.B., Le Bihan, T., Yu, M. (2011). S-nitrosylation of NADPH oxidase regulates cell death in plant immunity. Nature. 478(7368):264-268. doi: 10.1038/nature10427

Zhang, P., Jackson, E., Li, X., & Zhang,Y. (2025). Salicylic acid and jasmonic acid in plant immunity, Horticulture Research. 12(7), uhaf082, <https://doi.org/10.1093/hr/uhaf082>.

Zhao,W., Li, Z., Fan, J., Hu, C., Yang, R., Qi, X.,et al. (2015).Identification of jasmonic acid-associated micro RNAs and characterization of the regulatory roles of the miR319/TCP4 module under root-knot nematode stress in tomato. Journal of Experimental Botany. 66(15): 4653–4667. <https://doi.org/10.1093/jxb/erv238>

Zheng, C., Chen, J.P., Wang, X.W., Li, P. (2025). Reactive Oxygen Species in Plants: Metabolism, Signaling, and Oxidative Modifications. Antioxidants. 14, 617. https://doi.org/10.3390/antiox14060617

Zhiqi, H., Tingyi, W., Dongdong, C., Lan, S., Guangheng, Z., Qian, Q, et al.(2025). Leucine-Rich Repeat Protein family regulates stress tolerance and development in plants. Rice Science. 32(1): 32-43, <https://doi.org/10.1016/j.rsci.2024.12.003>.

Zou, J. , Kyndt, T.,  Yu, J.,  & Zhou, J. (2024). Plant–nematode battle: engagement of complex signaling network. Trends in Parasitology. 40(9):846-857. <https://doi.org/10.1016/j.pt.2024.07.010>.