Phytochemical Analysis Of Leaves And Flowers Of *Cassia Auriculata* Linn.

**ABSTRACT**

The present study reveals the morphological characters, organoleptic and fluorescence analysis by using various chemicals and reagents to examine the presence of phytochemicals visually in powdered samples of leaves and flowers of *Cassia auriculata*. The qualitative analysis represents the presence of various phytochemicals *viz*., steroids, reducing sugars, sugars, alkaloids, phenols, flavonoids, saponins, tannins, anthroquinine and aminoacids in crude extracts (aqueous, ethanol and petroleum ether) of leaves and flowers of *Cassia auriculata*. The presence of minerals *i.e.,* potassium, phosphorous, sulphur, calcium, were observed in leaves and flowers of *Cassia auriculata* by qualitatively. Quantitative analysis deals on maximum amount of flavonoids in flowers of *Cassia auriculata* and lesser amounts of alkaloids were recorded in leaves and flowers of *Cassia*  respectively.

***Key words:*** *Cassia auriculata,* Fluorescence analysis, Phytochemicals,

**INTRODUCTION**

Phytochemicals are a large group of plant-derived compounds that are hypothesis to be responsible for much of the disease production conferred by diet high in fruits vegetables, beans, cereal and plant-based beverages such as tea and wine. Based on their chemical structure phytochemicals can be grouped into such groups as tannins, flavonoids, glycosides, saponins, alkaloids, triterpeniods and sterols [1].

For a long period of time, plants have been a valuable source of natural products for maintaining human health, especially in the last decade, with more intensive studies for natural therapies. About 80% of individuals from developed countries use traditional medicine, which has compounds derived from medicine plants. Sources for new safe, biodegradable and renewable drugs. The use of plants as therapeutic agents in addition to being used as food is age long [2].

Though the therapeutic uses of plants by the primitive people lack scientific explanations [3] there is a great awareness in the use and significance of these medicinal floras by the World Health Organization in several resource-poor nations [4]. This has led to intensified efforts on the documentation of medicinal plants [5].

Therefore, such plants should be investigated to better understand their properties, safety and efficiency [6]. The cost of drugs in use today is too expensive for the majority of the population in the third world countries and therefore the search for some cheap sources of antimicrobial substances in nature become inevitable plants are good.

*Cassia auriculata* Linn (Family : Caesalpinaceae) distributed throughout hot deciduous forests of India and holds a very prestigious position in Ayurveda and Siddha system of medicine. The plant has been reported to possess antipyretic [7]; hepato protective [8]; anti diabetic, anti peroxidative and antihyperglyceamic [9] and microbicidal activity [10]. The flowers are used to treat urinary discharges, nocturnal emissions, diabetes and throat irritation [11]. Hence, the objectives of the present study is focused on to evaluate the phytochemicals qualitatively and quantitatively of the *Cassia auriculata* using various extracts.

**MATERIALS AND METHODS**

**Collection Of Plant Materials**

Leaves and flowers of *Cassia auriculata* (*Caesalpinaceae*)were collected from Karuppanathi Dam near Chokampatti, Tenkasi (TK.) Tamil Nadu.

**Organoleptic Study**

The plant powder characteristics such as color, odor, taste and nature were evaluated.

**Preparation of Crude extracts**

 The collected plant samples were thoroughly washed under running tap water and shade dried. The samples were pulverised with the help of a blender / mixer and soaked in aqueous, ethanol and petroleum ether were prepared by macerating one gram of powder with 10ml of solvents taken in flasks wrapped separately in Erlenmeyer flasks. The preparation were allowed to stand for 4 hrs. at room temperature. Then the extracts were filtered using Whatmann filter No. 1 and stored for further use. The crude extracts were analysed qualitatively and quantitatively and yield percentage of the extract was determined by using the equation [12].

 Yield (%) = W2 – W1 / W0 x 100

Where W2 - weight of the extract and container

 W1 – weight of the empty container and W0  is the weight of the initial dried sample.

**Fluorescence Analysis Of The Powder**

The fluorescence analysis of powdered samples *i.e*., leaves and flower mixed with different solvents and reagents were carried out using long ultraviolet (UV) lamps (365nm) and visible wavelengths [13 – 15].

**Preliminary Phytochemical Analysis**

The qualitative tests for extracts to detect the presence of phytochemicals such as alkaloid, tannin, saponin, flavonoid and phenol were carried out using standard procedures [16] .

**Quantitative analysis of Phytochemicals**

**Phenol Determination**

100 mg of the extract of the sample was weighed accurately and dissolved in 100 ml of triple distilled water (TDW). 1 ml of this solution was transferred to a test tube, then 0.5 ml 2N of the Folin Ciocalteu reagent and 1.5ml 20% of Na2CO3 solution was added and ultimately the volume was made up to 8 ml with TDW followed by vigorous shaking and finally allowed to stand for 2 hours after which the absorbance was taken at 765 nm. These data were used to estimate the total phenolic content using a standard calibration curve obtained from various diluted concentrations of gallic acid.

**Alkaloid Determination**

Five gram of the powered plant samples were weighed into 200 ml of 20% acetic acid in ethanol was added and covered to stand for 4 h. This was filtered and the extract was concentrated using a water bath to one-quarter of the original volume. Concentrated ammonium hydroxide was added drop wise to the extract until the precipitation was complete. The whole solution was allowed to settle and the precipitate was collected by filtration and weighed.

**Flavonoid Determination**

To estimate flavonoids quantitatively, 10 g of powdered sample of each plant material was extracted twice with 10 ml of 80% aqueous methanol at room temperature. The whole solution was filtered through Whatmann filter paper No.1, the filtrate was later transferred into crucibles, evaporated to dryness on a water bath to a constant weight [17].

**Tannin Determination**

Distilled water (50 ml) was added to 500 mg of the sample taken in a 500 ml flask and kept in shaker for 1 h. It was filtered into a 50 ml volumetric flask and made up to the mark. Then 5 ml of the filtrate was pippetted out into a test tube and mixed with 2 ml (10 fold diluted) of 0.1 M FeCl3 in 0.1 N HCl and 0.008 M potassium ferrocyanide. The absorbance was measured at 605 nm within 10min [18].

**RESULTS AND DISCUSSION**

Plant based antibacterial preparations are known to have enormous therapeutic potential due to the presence of several antibacterial substances [19] . In order to identity the antibacterial active compounds of the herbs or medicinal plants, such factor should be taken into consideration including the extraction and bio assay techniques employed. Generally, the type of solvent used for the extraction plays a significant role in the solubility of the active principles of plant material that not only affected the amount of representative compounds where consequently will influence the antibacterial activity of the extract [20]. Flowers of *Cassia auriculata* are showy with dense bunches, brightly yellow color corolla, leaf obovate with tapering apex, dark or prominent vein, waxy coated and coriaceous (Table 1).

**Organoleptic studies**

 It is an important parameter of powder analysis which is technique for the qualitative detection of morphological and sensory profile of drugs [[21]. The study revealed the characteristic color, odor, taste and nature of powdered medicinal species (Table 2). The results of pharmacognostical and phytochemical studies conducted in the bark and leaves of *Terminalia travancorensis* Wight & Arn. (Combretaceae), a tree, endemic to the Western Ghats and their pharmacognostical studies included the organoleptic, physico-chemical and fluorescence analysis of the bark and leaf powder [22].

**Percentage yield of crude extracts**

Table 3. represents the percentage yield of crude extracts in medicinal plants in which high yield occurred in ethanolic extract of leaves (24%) and low yield was observed in flowers of *Cassia auriculata* (6%). Highest yield percentage accounted for 1.70% was obtained in maceration with methanol followed by ethyl acetate at 1.28% and n-hexane at 0.93% in a study done on *Psidium guajava* leaves extract, suggested that methanol was the best solvent for solubility of several compounds [23]. Never the less, the preferred extraction method should be simple, fast, economical and importantly able to retain the important phyto constituents [24].

**Fluorescence analysis**

In fluorescence analysis study shows specific colour appeared with specific reagents (Table.4). The powdered leaves of *Cassia* recorded as light green to brown colour under ordinary white light and fluorescent green to dark brown appeared in long UV light (365 nm). The ***Cassia*** flower powder appeared as light yellow to dark black in ordinary light, while under long UV light (365 nm) exposed showed fluorescence green to dark red in colour.

Herbal drug which are used in various traditional medicine needs detailed investigation with ethano pharmacological approach. The present study provides information in respect of identification, standardization of herbal drug of *Cassia auriculata* of Ayurvedic compendia. Correct identification and quality assurance of the starting materials is an essential pre requiste to ensure reproducible quality of herbal medicine which will contribute to its safety and efficacy [25].

The fluorescence analysis is adequately sensitive and enables the precise and accurate determination over a satisfactory concentration range without several time consuming dilution steps prior to analysis of pharmaceutical samples [26].The fluroscence colour is specific for each compound. A non fluorescent compound may fluorescence it mixed with impurities that are fluorescent.

**Qualitative analysis**

As shown in the Table 5. maximum number of phytochemicals were observed in water extract of *Cassia* leaf followed by ethanol extracts of *Cassia* flower. Lesser number of phytochemicals was seen in ethanol extract of *Cassia* leafs.

Our results are positive correlated with ethanolic extracts revealed the presence of the high concentration of tannins, reducing sugars and steroids in the stem, bark and roots [27]. Flavonoids, phenolics and protein prevent in high concentration in the stem, bark while anthroquenone, glycosides and alkaloids were present in the leaves and roots of ***Cassia abbreviate*** respectively.

Bio activity properties of herbs are where closely related to their phytochemicals consitutents which are classified into various major groups [28]. Ethanolic extract of *Cassia* flower possess reducing sugars, phenol, tannins, steroids can summarized in Table 3. However, it is important to highlight that the type of diluent used was the main factor that could influence in variation of phytoconstituents being extracted.

While in aqueous extract of *Cassia* flower showed the absence of phytoconstituents namely alkaloids, saponins, antheroquinone other study that evaluated the existence of phytochemicals of petroleum ether, ethanol and aqeous extracts also revealed the difference in the solubility of active compouds [29].

Phenolic compounds were the most common secondary metabolites implicated with microbial growth inhibitory action in herbs [30, 31]. Plants are rich in wide variety of secondary metabolites such as tannin, terpenoids, alkaloids and flavonoids etc., which have been *in vitro* to have antibacterial and antifungal properties.

**Quantitative estimation of major phyto components**

Table . 6 reveals the amount of phytochemicals quantitatively, in which more amount of flavonoids was recorded in flowers of *Cassia auriculata* (28 %) followed by leaves of *Cassia*  (14 %). Lowest amount of alkaloids was observed in leaves and flowers of *Cassia* (6% and 5% of alkaloids) respectively, while tannins were more in leaves of *Cassia* (0.65 mg/g) followed by *Cassia*  flowers (0.47 mg/g).

Plant based compounds have several biological applications. An alkaloid compound has been reported to exhibit lethal effects against colon and breast cancer cells and has been used for antimicrobial, antiviral, antiprotozoal and anti tumor applications [32]. Flavonoids have been used for anti diabetic, anti microbial activities, anti -inflammatory and anti aging preparations [33]. Previous researchers have shown that the plant phenolic compounds offer the role of potential natural antioxidants [34, 35].

Flavonoids and phenols have raised particular interest because of their potential biological characteristics as antioxidant, antiestrogenic, anti inflammatory, immune modulatory, cardio protective and anti carcinogenic compounds [36]. Tannins play an essential role in many biological applications because of their anti inflammatory, cardio protective and anti microbial properties [37] . Presence of alkaloids, tannins, total phenols, carbohydrates, total tannins, saponins, terpenoids and total glycosides in varying content using various solvents *viz.*, ethanol, methanol, acetone, choloroform, petroleum ether and also in water, clearly shown that the more number of photochemical compounds are maximum soluble in ethanol solvent [38].

**Qualitative analysis of minerals**

 The presence of minerals *i.e.,* potassium, phosphorous, sulphur, calcium, were observed in leaves and flowers of *Cassia auriculata* by qualitatively. Similarly reports that different plant species, elemental accumulation depends on various factors such as the type of soil, fertilization method, plant species and environmental conditions [39].

Proximate and mineral nutrient analysis validates the signigicance of the extracts with a high amount of carbohydrates and proteins along with siginificantly high amount of zinc, iron, manganese, calcium, magnesium and potassium involved in various metabilic reactions of *Calligomum crinitum* [40].

**REFERENCES**

1. Tiwari .P., Kumar B. Kaur M., Karu G and Karu H. 2011. Phytochemical screening and extraction: A review. *International Pharmacerica Sciencia.* **1**:1.
2. Sachin Chaudhary and Anit Kumar, 2014. Phytochemical analysis and assessment of *in vitro* antihelminthinic activity of *Cassia auriculata* Linn. Leaves.
3. Dutta, 1994. A.C. Botany for degree students. Oxford University Press, London, 73.
4. WHO, 2002. WHO traditional medicine strategy 2002- 2005. WHO, Geneva.
5. Perumal S.R and Ignacimuthu S. 2000. Antibacterial activity of some folklore medicinal plants used by tribes in Western Ghats of India. *J. Ethanopharmacol*. 69 : 63-71.
6. Muthukumaran .P., Elayarani .M., Shanmuganathan.P and Cholarajan .A. 2001. Antimicrobial Activities of *Cassia auriculata* L. and *Morinda tinctoria* Roxb. *International Journal of Research in Pure and Applied Microbiology,* 1(2): 9-12.
7. Wealth of India, 1950. Raw materials, Vol. II. Publications and Information Directorate, *Council of Scientific and Industrial Research*, New Delhi, 95.
8. Rao K.N. and Vedavathy S. 1991. Antipyretic activity of six indigenous medicinal plants of Tirmula hills. *J. Ethnopharmacol*., 33: 193-196.
9. Manickam .P., Namasivaqyam .N., Periyasamy .V and Rajagopal .S. 2002. Effect of *Cassia auriculata* leaf extract on lipids in rats with alcoholic liver injury. *Asia Pacific J. Cli. Nut*., 11: 57-163.
10. Pari L and Latha M, 2003. Antihyperglycaemic effect of *Cassia auriculata* in experimental diabetes and its effects on key metabolic enzymes involved in carbohydrate metabolism. *Cli. Exp. Pharmacol. Physiol*.; 30: 38-43.
11. Prakash S.K., 2006. Effects of Herbal extracts towards microbicidal activity against pathogenic *Escherichia coli* in Poultry. *Int. J. Poultry Sci*., 5: 259-261.
12. Anokwuru, C.P., G.N. Anyasor, O.Ajibaye, O. Fakoya and P. Okebugwu, 2011. Effect of extraction solvents on phenolic, flavonoid and antioxidant activities of three Nigerian Medicinal plants. *Nat. Sci.,* **9**: 53 - 61.
13. Chase C.R. and Pratt R.S. 1949. Fluorescence of Powdered Vegetable drugs with particular reference to Development of a System of Identification. *J. Am. Pharmacol. Assoc.,* **38**, p 32.
14. Kokoshi C.J., Kokoshi R.J. and Sharma F.T. 1958. Fluorescence of powdered vegetable drugs under Ultraviolet radiation. *J. Pharm. Asses*., **47** : 715 - 717
15. Wilson Color Chart – Horticultural Color Chart,. Vol. 1,2, Henry Stone and Son Ltd. Banbury, Great Britain, 1938, 1941, 1 -100, 10 - 200.
16. Harborne J.B. 1998. Phytochemical Methods, A guide to modern technique of plant analysis, Chapman and Hall, London, pp : 108 - 148.

### Kumaran, A. and Karunakaran, J. R. 2006. Antioxidant Activities of the Methanol Extract of  *Cardiospermum halicacabum*. *Pharmaceutical Biology*, 44(2), 146 – 151.

1. van-Burden, T.P. and T Robinson, 1981. The biochemistry of alkaloids. Springer, Heidelberg, New York.
2. Srinivasan, D., S. Nathan, T. Suresh and P.L. Perumalsamy, 2001. Antimicrobial activity of certain Indian medicinal plants used in folkloric medicine. *J. Ethnopharmacol*., 74: 217-220.
3. Nair, R., T. Kalariya and S. Chanda, 2005. Antibacterial activity of some selected Indian medicinal flora. *Turk. J. Biol*., 29: 41- 47.
4. Kokate C.K., Purohit A.P. and Gokhale S.B. 2002. Text book of Pharmacognosy. 18th ed. Pune: Nirali Prakashan.
5. Lakshmi. M., Bindu, R.. N and Chandrasekhara Pillai P. K. 2012. Pharmacognostic evaluation and phytochemical analysis of bark and leaves of *Terminalia travancorensis* Wight and Arn. (Combretaceae). *Journal of Pharmacy Research,* **5** (4) : 1988 – 1991.
6. Shafiei, S.N.S., 2012. *In-vitro* antibacterial activity and phytochemical screening of bioactive compounds from Guava (*Psidium guajava* L.) crude leaf extracts. M.Sc. Thesis, Universiti Putra Malaysia, Malaysia.
7. Annegowda, H.V., P.V. Tan, M.N.Mordi, S. Ramanathan, M.R. Hamdan, M.H.Sulaiman and S.M. Mansor, 2013. TLC – bioautography-guided isolation, HPTLC and GC-MS assisted analysis of bioactives of *Piper betle* leaves extract obtained from various extraction techniques: *In vitro* evaluation of phenolic content, antioxidant and antimicrobial activities. *Food Anal. Methods*, **6** : 715 - 726.
8. Ghildiyal. S., Gautam MK., Joshi, V.K and Goel R.K, 2012. Pharmacognostical study of *Hedychium spicatum* ( Ham – Ex-n Smith ) rhizome. *Asian Journal of Tropical Biomedicine ,* ( In press).
9. Pimenta A.M., Montenegro M.C., Ara-Ujo A.N and Martinez J.C 2006. Application of sequential injections analysis to pharmaceutical analysis. *Journal of Pharmaceutical Biomedical Analysis,* **40**: 16-34.

### Huang, Q., Liu, X., Zhao, G., Hu, T. and Wang, Y., 2018. Potential and challenges of tannins as an alternative to in-feed antibiotics for farm animal production. *Anim. Nutr*. 4 (2) : 137–150.

1. Al-Daihan, S., M. Al-Faham, N. Al-Shawi, R. Almayman, A. Brnawi, S. Zargar and R.S. Bhat, 2013. Antibacterial activity and phytochemical screening of some medicinal plants commonly used in Saudi Arabia against selected pathogenic microorganisms. *J. King Saud Univ. Sci.,* 25: 115-120.
2. Syahidah, A., C.R. Saad, H.M. Daud and Y.M. Abdelhadi, 2015. Status and potential of herbal applications in aquaculture: A review. *Iran. J. Fish. Sci*., 14: 27-44.
3. Burt, S. 2004. Essential oils: Their antibacterial properties and potential applications in foods: A review. *Int. J. Food Microbiol*., 94: 223-253.
4. Witkowska, A.M., D.K. Hickey, M. Alonso-Gomez and M. Wilkinson, 2013. Evaluation of antimicrobial activities of commercial herb and spice extracts against selected food-borne bacteria. *J. Food Res*., 2: 37-54.
5. Rinaldi, M.V.N., I.E.C. Diaz, I.B. Suffredini and P.R.H. Moreno, 2017. Alklaoids and biological activity of beriba (*Annonan hypoglauca*). *Rev. Bras. Pharmacogn*.,**27** (1) : 77 - 83.

### Wang, T.-Y., Li, Q., Bi, K.-S., 2018. Bioactive flavonoids in medicinal plants: structure, activity and biological fate. *Asian J. Pharm. Sci.* 13 (1), 12–23.

1. Dhiman, A., S. Nanda and S. Ahmad 2016. A quest for staunch effects of flavonoids, Utopian protection against hepatic ailments. *Arab.J. Chem*., **9**:1813-23.

### Takaidza, S., Mtunzi, F., Pillay, M., 2018. Analysis of the phytochemical contents and antioxidant activities of crude extracts from Tulbaghia species. *J. Tradit. Chin. Med.* 38 (2): 272 – 279.

1. Kumar and Baskar, 2015. Screening and quantification of phytochemicals in the leaves and flowers of *Tabernaemontana heyneana* wall-a near threatened medicinal plant. *Indian Journal of Natural Product Resources*, **5**:237 - 243.

### Abu Zarin, M., Wan, H.Y., Isha, A., Armania, N., 2016. Antioxidant, antimicrobial and cytotoxic potential of condensed tannins from *Leucaena leucocephala* hybrid-Rendang. *Food Sci. Human Wellness* 5 (2): 65 – 75.

1. Suman Kumar Ratnampally and Venkatshwar Chinna, 2017. Quantitative analysis of phytochemicals in the Bark extracts of medicinally important plant *Cassia fistula* Linn. *Int. J. Curr. Microbiol App, Sci.*, **6** (4) : 1073 - 1079.
2. Bengtsson, H., Oborn, I., Jonsson, S., Nilsson, I., Andersson, A. 2003. Field balance of some mineral nutrients and trace elements in organic and conventional dairy farming a case study at Objebyn, Sweden. *Eur. J. Agron*., **20**:101-116.
3. Naqbi , K.M.A.A., Karthiswaran , K., Kurup, S.S., Abdul MuhsenAlyafei, M. and Jaleel, A. 2022. Phytochemcials, proximate composition, mineral analysis and *in vitro* antioxidant activity of *Calligomum crinitum*  Boiss. *Horticulture*, **81**(156):1-14.

 **Table . 1. Morphological features *of Cassia auriculata***

|  |  |  |  |
| --- | --- | --- | --- |
| **S. No** | **Parameters** | **Leaf** | **Flower** |
| 1. | Habit | Shrub | Shrub |
| 2. | Root system | Tap root | Tap root |
| 3. | Stem | Branches | Branches |
| 4. | Leaf | Lanceolate | Lanceolate |
| 5. | Flower | Large and showy | Large and showy |
| 6. | Colour | Green | Yellow |
| 7. | Inflorescence | Racemose | Racemose |

**Table. 2. Oraganoleptic character *of Cassia auriculata***

|  |  |  |
| --- | --- | --- |
| **Characters** | **Leaf** | **Flower** |
| Odour | No smell | No smell |
| Taste | Bitter | Bitter |
| Colour | Normal green | Yellow |
| Texture | Fine | Fine |

 **Table 3: Percentage yield of** *Cassia auriculata*

|  |
| --- |
|  **Yield percentage** |
| **Ethanolic Extract** | **Wt. of initial dried sample (W0)** | **Wt. of empty container (W1)** | **Wt. of extract & container (W2)** | **Yield (%)** |
| Leaves  | 1.0 | 65.40 | 65.64 | 24 |
| Flowers  | 1.0 | 61.37 | 61.43 | 6 |

**Table 4. Fluorescence analysis of** *Cassia**auriculata*

|  |  |  |  |
| --- | --- | --- | --- |
| S. No | Reagent Used | *Cassia* leaf | *Cassia* flower |
| Long UV light(365 nm) | Visiblelight | Long UV light(365 nm) | Visiblelight |
| 1. | Concentrated HNO3 | Dark brown | Light brown | Dark red  | Dark red |
| 2. | Concentrated HCL | Greenish brown | Pale green  | Dark green  | Light green |
| 3. | Acetone | Dark brown | Brown | Light yellow | Light yellow |
| 4. | NH3 + Ammonia | Brownish green | Brown | Flourescence green | Light yellow |
| 5. | Chloroform | Dark green | Light green | Dark yellow | Light yellow |
| 6. | Benzene | Dark green  | Light green | Greenish yellow | Light yellow |
| 7. | Ethanol | Dark brown  | Light brown | Greenish yellow | Light yellow |
| 8. | Petroleum ether | Dark green | Light green | Dark yellow | Light yellow |
| 9. | Glacial acidic acid | Dark brown | Dark brown | Greenish yellow | Light yellow |
| 10. | HNO3 + NH3 | Greenish brown | Brown  | Dark black  | Dark red |
| 11. | H2SO4 | Dark brown | Dark brown | Dark black | Dark black |
| 12. | 50% HNO3 | Greenish brown | Brown | Dark black | Dark red |
| 13. | 50% HCL | Dark green  | Light green | Greenish yellow | Light yellow |
| 14. | 1N Aqueous NaOH | Dark green | Light green | Dark red  | Dark black |
| 15. | 1N Alcoholic NaOH | Dark brown  | Dark brown | Dark black  | Light black |
| 16. | 50% H2SO4 | Dark brown | Light brown | Brownish green | Light yellow |
| 17. | Ferric chloride | Flourescent green | Brown | Dark red | Light red |
| 18. | 40% of NaOH + 10% Lead acetate | Dark green | Brown | Greenish yellow | Light red |

**Table 5. Phytochemical analysis of** *Cassia**auriculata* **in various extracts**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| S.No | Phytoconsitituents | *Aqueous* | Ethanolic | Petroleum ether |
|  |  |  |
| Leaf | Flowers | Leaf | Flowers | Leaf | Flower |
| 1 | Steriods | - | - | + | - | + | - |
| 2 | Reducing sugar | - | + | - | + | - | - |
| 3 | Sugar | + | + | - | + | + | + |
| 4 | Alkaloids | + | - | - | - | + | - |
| 5 | Phenol | + | + | - | + | + | + |
| 6 | Flavonoids | + | - | + | + | - | - |
| 7 | Saponin | + | - | + | + | - | + |
| 8 | Tannin | + | + | - | + | + | + |
| 9 | Anthroquenine | - | - | - | - | - | - |
| 10 | Amino acids | + | - | - | + | - | - |

**Table :6. Quantitative analysis of phytochemicals**

|  |  |  |  |
| --- | --- | --- | --- |
| **S.NO** | **Phytoconsituents** | ***Cassia* leaf** | ***Cassia* flower** |
| 1. | Flavonoids (%) | 14 | 28 |
| 2. | Phenols (mg / g) | 0.16 | 0.20 |
| 3. | Tannins (mg / g) | 0.65 | 0.47 |
| 4. | Alkaloids (%) | 6.0 | 5.0 |