**Effect of alkaloid-rich fraction of *Dacryodes edulis* leaves on the microstructure of the hippocampus and blood glucose levels of STZ-induced diabetic Wistar rats**

**Abstract**

**Aim:** the aim of this study was to elucidate the effect of alkaloid-rich fraction of *Dacryodes edulis* on body weight, blood glucose level and the microanatomy of the hippocampus of STZ-induced Diabetic Wistar rats.

**Methodology:** 25 adult male Wistar rats were used for the experiment. These rats were randomly assigned into 5 groups, each containing 5 rats designated as A, B, C, D and E. A group served as the normal control and were allowed access to drinking water and food *adlibitum.* Diabetes was induced in groups B to E through intraperitoneal administration of 60 mg/kg of Streptozotocin (STZ) for 7 days. Group B served as the untreated group, while groups C and D received 1000mg/kg and 500 mg/kg of alkaloid-rich fraction of *Dacryodes edulis*. Group E was administered 50 mg/kg of metformin.

**Results:** the results revealed increase body weight, elevated glucose levels and atrophied pyramidal cells in the pyramidal cell layer of the hippocampus, along with numerous vacuoles containing lipid deposits in the untreated group B compared to the control group A, groups C, D and E exhibited potential ameliorative effects from the alkaloid-rich fraction of *Dacryodes edulis* and metformin, though the effect was dose dependent.

**Conclusion:** Alkaloid-rich fraction of *Dacryodes edulis* is highly effective in reducing glucose levels, regulating body weight and protecting hippocampal neurons from neurodegeneration in STZ-induced diabetic adult male Wistar rats.

**Keywords**: Alkaloid, *Dacryodes edulis*, Diabetes, Neurodegeneration, pyramidal cells

**1. INTRODUCTION**

Diabetes mellitus is a progressive and debilitating condition characterized by prolonged elevated glucose level and disruption in the metabolism of carbohydrate, lipids and amino acids due to excessive production or inadequate utilization of glucose. This arises due to inadequate production or functioning of insulin in the pancreas (Inbaraj and Inbaraj, 2014; Gwarzo et al., 2014). The elevated blood glucose level in individuals results in polyuria, polydipsia as well as polyphagia (Martin et al., 2012; wild et al., 2004). Patients with diabetes havedeficiencies in their antioxidant defense system, which may increase their susceptibility to free radical-induced damage and the progression of diabetic sequelae (Gwarzo et al., 2014). Several corroborating factors imply that oxidative stress-induced harm is implicated in the etiology of a wide range of human disorders. Free radical-induced damage occurs when the generation of oxidative stressors surpasses the existing radical scavenging capacity. The stressors are bioactive compounds containing oxygen such as hydrogen peroxide, HCl and radical species like superoxide anion and hydroxyl radicals (Alhassan et al., 2009).

Global prevalence of diabetes among adults over 18 years has increased from 4.7% in 1980 to 8.5% in 2014. In 2016, WHO published 422 million adults living with diabetes and 1.5 million deaths yearly (Global Report on Diabetes Geneva, 2016). The international Diabetes mellitus is expected to rise from 387 million people in 2014 to 592 million within the next 20 years. 316 million with impaired glucose tolerance are at increases risk from the disease with projection showing that more than 1 billion people will be living with diabetes in 2035 (daRocha et al., 2016). High blood sugar is a common effect of uncontrolled diabetes and may lead to serious damage to many body systems such as the nervous system and increase the risk of untimely death (Lallukka et al., 2016).

Nervous system transmits signals between the brain and the rest of the body including internal organs (Medlineplus, 2016). The basic unit of the nervous system is a nerve. The human brain contains about 100 billion nerves. The nervous system also contains non-neuronal cells called glia. These glial cells perform many important functions that help maintain proper nervous system. The brain is made up of many pathways of communicating neurons and glia. These pathways allow different parts of the brain to communicate with each other and work together to control body functions, emotions, thinking, behavior and other activities (Medlineplus, 2016; Society for Neuroscience, 2012; National Institute of Neurological Disorders and Stroke, 2018). Neuroprotection refers to strategies of defending the central nervous system against neuronal injury caused by acute stroke, trauma or neurodegenerative disorders (Kummar et al., 2006).. In the past 20 years, the nutritional neuroscience emerged as a recognized discipline with the potential to make significant contributions to our understanding of the relationship between nutrition and cognitive functions (Gillete et al., 2007). The use of medicinal plants as an alternative prevention of neurodegenerative disease became paramount to many Scientist. Recently, the use of herbal plants such as *Telfairia occidentalis, Talinium triangulare, Averrhoa carambola* and *Dacryodes edulis* has gained attention for its potential to ameliorate neurodegenerative diseases such as Alzheimer’s-type cognitive dysfunction in Wistar rats. (Eru *et al*., 2024; 2022; 2021; 2020; Ifiok *et al*.,2024; Paulinus *et al*., 2024; Udoh *et al*., 2020).

*Dacryodes edulis* is highly nutritious, containing lipids, proteins and provitamins. Many health-enhancing chemical constituents such as alkaloids, tannins, flavonoids and saponin are present in parts the plant including its leaves. The bioactive compounds obtained from the leaves are reported to possess antioxidants, anti-sickle cell and antimicrobial activities (Ononamadu et al., 2019). Plant-derived antioxidants have been used in the treatment and management of various diseases, including neurological disorders (Sadhwani et al., 2021). This study was therefore conducted to investigate the effects of alkaloids from *Dacryodes edulis* on the microstructure of the hippocampus, blood glucose levels and insulin status of STZ-induced diabetic Wistar rats.

**2. METHODOLOGY**

**2.1. Experimental design and animals**

With ethical approval number: FAREC-FBMS 150-2/2023, twenty-five adult male Wistar rats weighing140-190g were bought from the animal farm and kept in the animal room. Before the experiment, the experimental animals were kept for acclimatization under standard conditions of temperature (27oC – 30oC) for two weeks; they were given rat chow and water *ad libitum*. After two weeks of adjustment, the rats were arbitrarily arrayed into five groups; each having five rats designated A, B, C, D and E.

**2.2 Plant extract preparation**

The mature fresh *Dacryodes edulis* leaves were plucked and washed in running tap water and thereafter rinsed properly in distilled water. It was air dried, powdered with a grinder and stored in an air tight plastic container till required for analysis. Crude alkaloid from *Dacryodes edulis* leaves was extracted by heating the powdered sample (500g) for 4 hours at 55°C with 1200 ml of ethanol (20%). The extract was filtered and residue was re-extracted with 200 ml of ethanol (20%). The extract was concentrated on water bath till the volume reduced to 200 ml, which was mixed with 100 ml diethyl ether in a separating funnel. The mixture was vigorously shaken and then the separating funnel was fixed in a stand till the development of aqueous and diethyl layer. Aqueous portion was collected while the diethyl ether portion was discarded. To the aqueous layer n-butanol (80 ml) was added and properly mixed by vigorous shaking. The n-butanol extract was treated with 10 ml of 5% NaCl solution. The resultant solution was concentrated on a water bath and the crude alkaloid extract (Zeb *et al*., 2014).

**2.3 Induction of Hyperglycaemia**

Diabetes was induced after 14 hours of fasting by intraperitoneal injection of 60 mg/kg body weight of STZ for seven days. Following the induction, the tail vein blood was collected to determine fasting blood glucose level using Accu-Chek glucometer. Rats with fasting blood glucose over 250 mg/dL were considered diabetic and included in the experiments.

**2.4 LD50 determination**

Using Lorke’s method (1983), LD50 of **alkaloids from *Dacryodes edulis***leaves were both established to be >5000 mg/kg and doses were determined using 10% and 5% of the established LD50.

**2.5 Administration of alkaloids from *Dacryodes edulis* leaves and metformin**

Group A served as the negative control and received animal feed and water *ad libitum*; group B served as the positive control and received 60 mg/kg body weight of STZ only for seven days; group C received 60 mg/kg body weight of STZ and 1000mg/kg body weight of alkaloids from *Dacryodes edulis* leaves; group D received 60mg/kg body weight of STZ and 500mg/kg body weight of alkaloids from *Dacryodes edulis* leaves; and group E received 60mg/kg body weight of STZ and 50 mg/kg body weight of metformin (bought from Bez Pharmacy, Etta Agbo Calabar-Nigeria). These extracts and drugs were administered for fourteen days.

**2.6 Histological tissue processing**

The animals were immolated with their brain tissues perfused and treated24 hours after the last administration. The whole brain was eviscerated, weighed and fixed in 10% buffered formal saline. This was done to maintain the morphological integrity of the tissue (Williams et al., 2006). The hippocampus of the rats was dissected out and used for histological studies using the haematoxylin and eosin (H & E) staining method and observed under light microscope (OMAX: 40X-2500X). During tissue processing, the hippocampal tissue was dehydrated through ascending percentage of alcohol for an hour each. The tissue was cleared in two changes of xylene for an hour each, infiltrated and embedded in molten paraffin wax. The solid tissue blocks in paraffin were mounted in the rotator microtome and sections were cut at six µm. The cut sections were floated in warm water bath, then picked and mount with an albumenized slide. Paraffin slides of the hippocampal tissue were de-waxed of paraffin through two changes of xylene for 5 minutes each, rehydrated through ascending percentage of alcohol and rinsed under ceaseless tap water. For 15 minutes, cut tissues were later dyed with haematoxylin and rinsed under ceaseless tap water for 5 minutes. For a minute, cut tissues were distinguished in acid alcohol, blued and counter stain with Eosin for another 1 minute. Sections were rinsed in tap water, dehydrate and cleared in xylene, allowed to dry with few drops of distyrene plasticizer xylene (DPX) kept on the slide and cover slipped (Avwioro, 2010).

**2.7 Statistical analysis**

Data were analyzed using a statistical package for social science version 21.0 The student t-test was used with data represented as mean ± standard error of the mean (SEM) and statistically significant at p < 0.05.

**3. RESULTS**

**3.1 Morphological observation**

The initial and final body weights of rats were monitored during the experimental period. Group A revealed 154.79±7.16 and 210.05±10.04, respectively; group B are 244.71±13.51 and 211.73±16.39, respectively; group C results are results are 184.24±1.82 and 197.24±6.01, respectively; group D results are 156.39±8.64 and 215.82±1.92, respectively while group E results are 161.05±5.55 and 183.32±4.55, respectively (figure 1). It was observed that there was significant decrease in body weight compared to the control group (p<0.05) as well as other groups. The result also showed a significant (p<0.05) increase in body weight of the rats in group A compared to the rats in Group C and D. The initial weight, final weight, change in body weight and percentage change in body weight for all the groups are shown in figure 1.

**3.2 Blood sugar level**

The STZ-induced diabetic rats exhibited a significant increase in blood sugar level (P<0.05), similar to the treatment groups C to E before the initial treatment of alkaloids from *Dacryode edulis* and metformin. After administration of treatment, the results showed a significant (p>0.05) reduction in blood sugar level in the treatment groups (C-E) compared to the normal group A and diabetic control group B. There was also a significant (p<0.05) difference in the change in the blood sugar level of the rats in the high dose treatment group C compared to the drug treatment group E (figure 2).

**3.3 Histological observations**

Sections of the hippocampus stained with H & E in the control group a normal pyramidal cells in the pyramidal layer with numerous neuropils. Neuroglial cells and blood capillaries are also present in other layers (granular and molecular). Group B revealed atrophied pyramidal cells, numerous neuropils and capillaries are also present (plate 2). Group C revealed distorted pyramidal cells with numerous vacuoles compared to the control group (plate 3). Groups D and E revealed normal pyramidal cells compared to the control groups, neuroglial cells and neuropils are also present (plates 4 & 5).

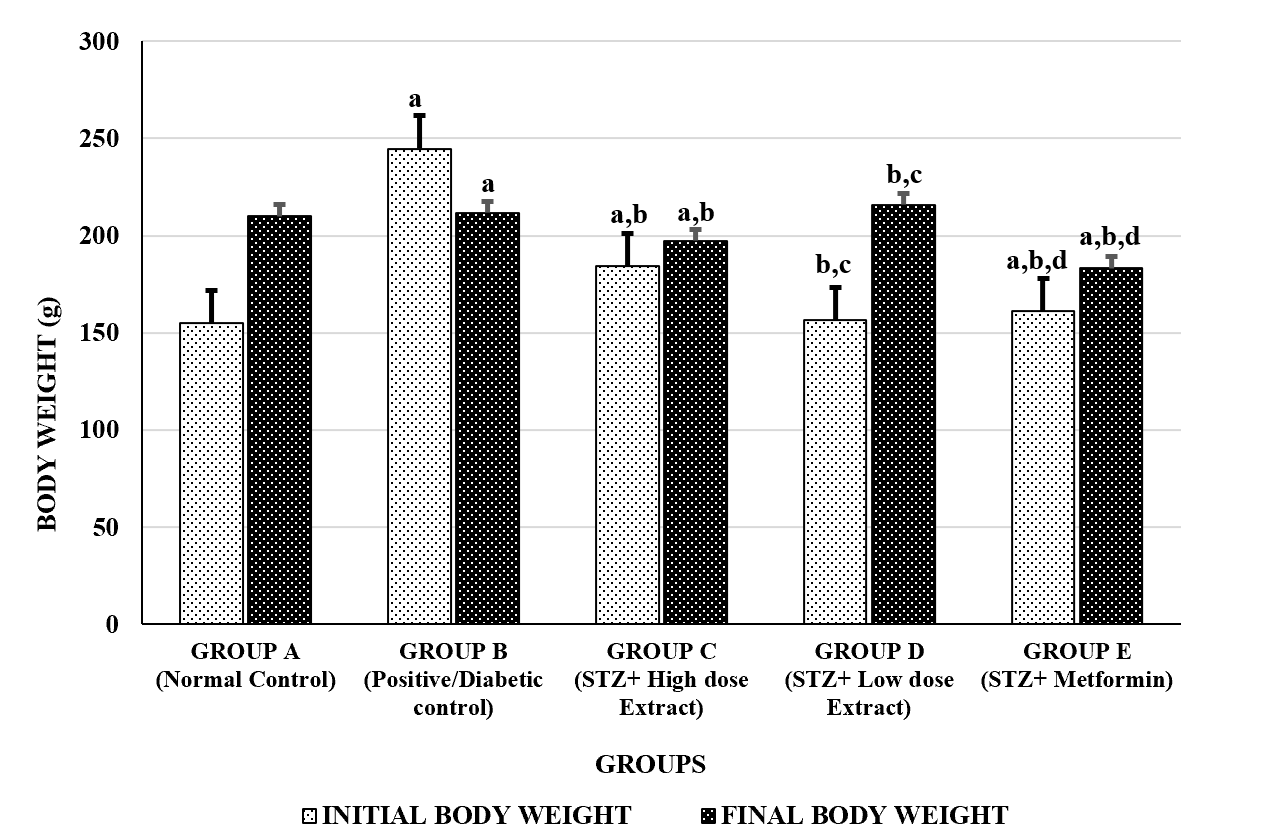


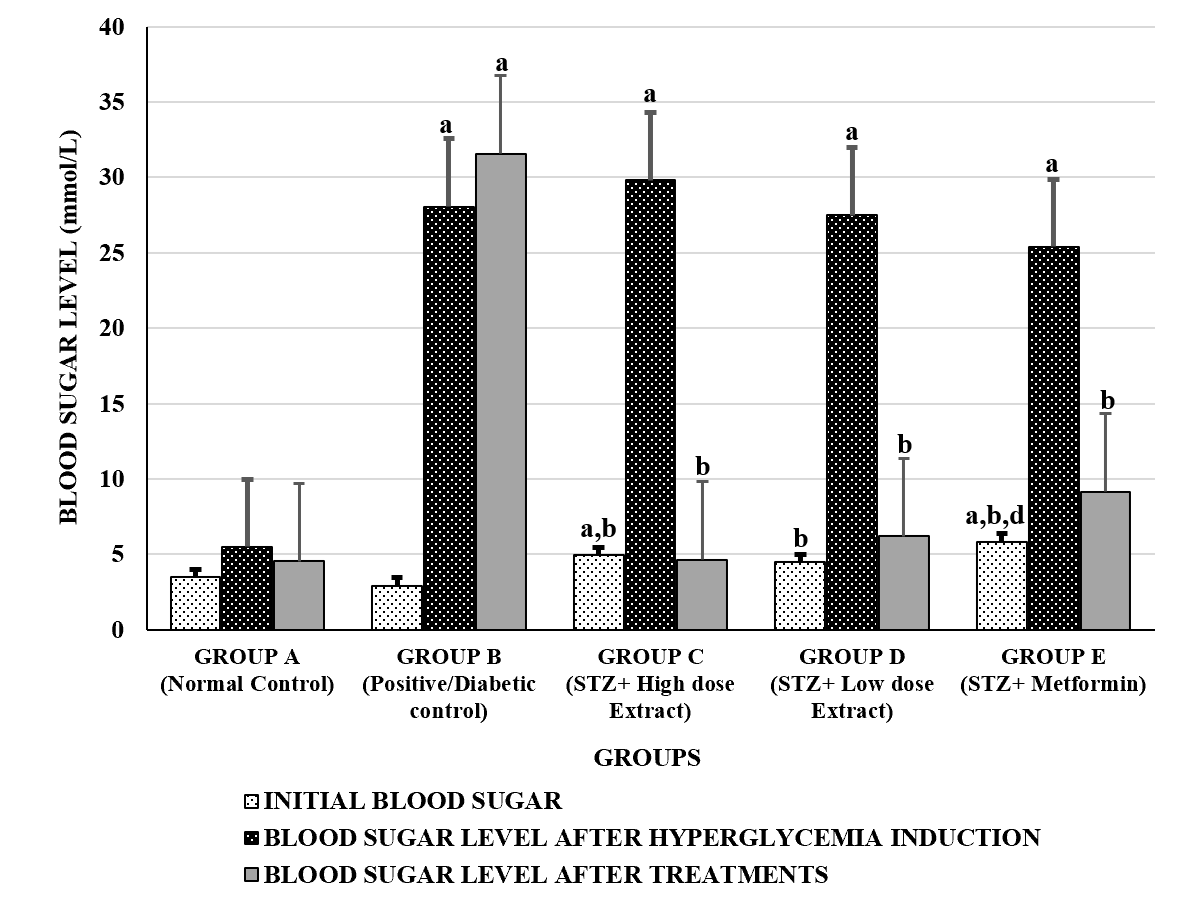
Figure 1: Bar graph showing the initial and final body weight of the experimental animals

a = significantly different from Control group at p<0.05.

b = significantly different from Diabetic group at p<0.05.

c = significantly different from High dose treatment group at p<0.05.

= significantly different from Low dose treatment group at p<0.05.



**Figure 2: Comparison of the blood sugar levels (mmol/L) in the different experimental groups.**

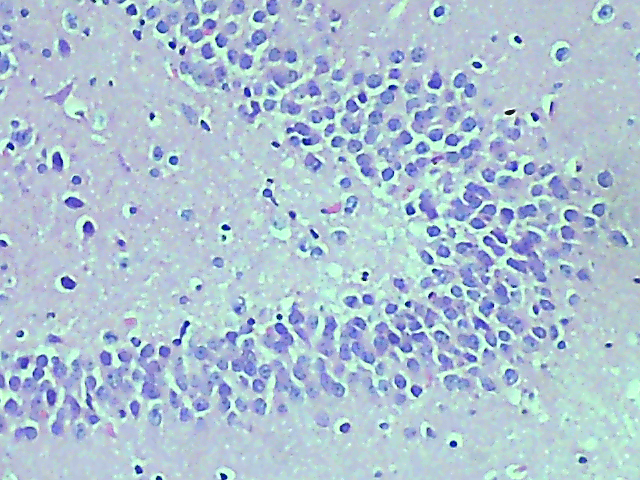
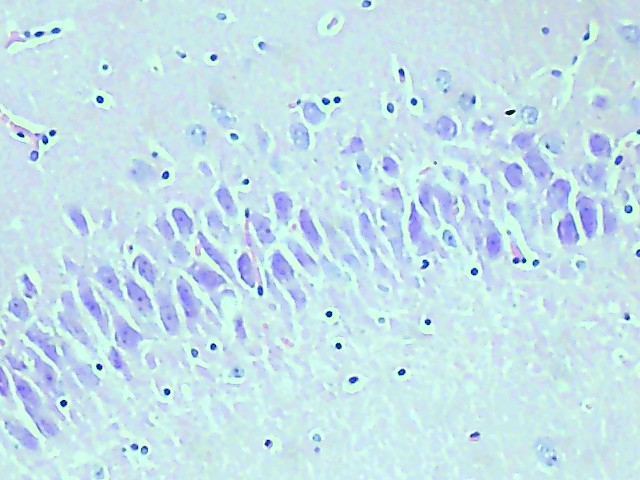
Values are expressed as mean + SEM, n =5.

a = significantly different from Control group at p<0.05.

b = significantly different from Diabetic group at p<0.05.

c = significantly different from High dose treatment group at p<0.05.

d = significantly different from Low dose treatment group at p<0.05



PCL

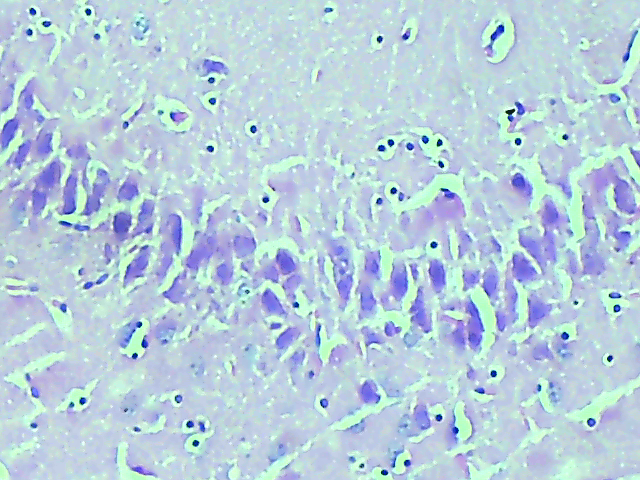
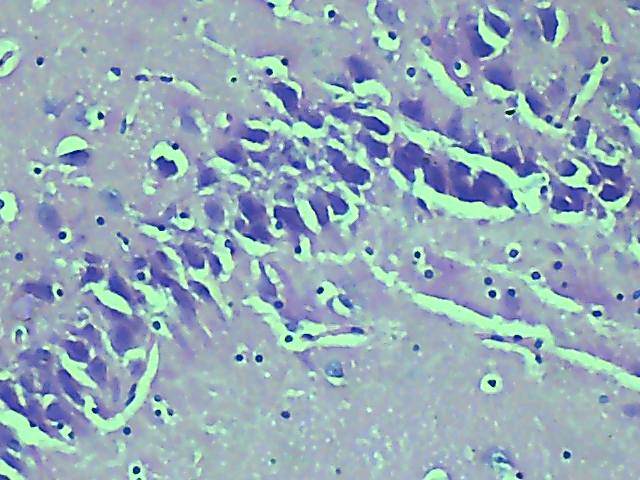
PCL

Plate 2 section of the hippocampus showing atrophied pyramidal cells in the pyramidal cell layer with numerous neuropils, neuroglial cells and vacuoles

Plate 1 section of the hippocampus showing the three layers with normal pyramidal cells in the pyramidal cell layer (PCL).

B

A



PCL

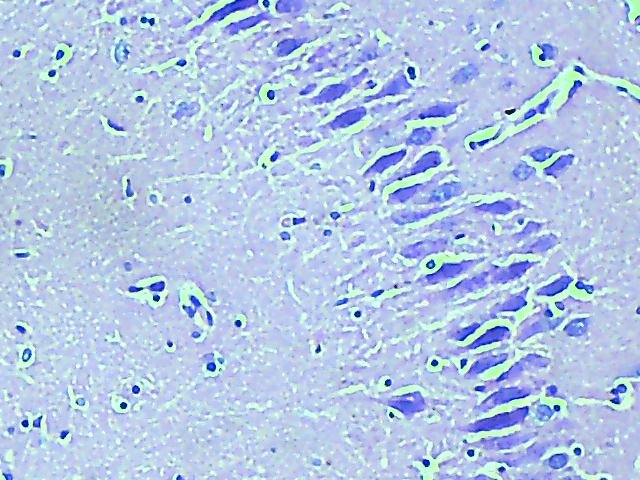
PCL

Plate 4 section of the hippocampus showing normal pyramidal cells with neuroglial cells and less vacuoles.

Plate 3 section of the hippocampus showing hypertrophied pyramidal cells in the pyramidal cell layer with numerous vacuoles.

D

C



E

PCL

Plate 5 section of the hippocampus showing normal pyramidal cells, neuroglial cells, blood capillaries, neuropils and less vacuoles.

**4. Discussion**

Diabetes is a chronic disorder associated with many debilitating multi-systemic complications and are of many causes. The spectrum of the diabetic pathology is complex and thus would require beyond single therapy for management of such complications. The more acceptable approach should embrace alternatives to orthodox care, hence diabetes remains a public health concern with estimated 463 million cases worldwide according to Tuomi et al (2014). In 2017, diabetes resulted in approximately 4.2 million deaths. It is the 7th leading cause of death globally according to Boadu et al., (2017). The pooled prevalence of 5.77% in Nigeria suggests that about 11.2 million people are living with diabetes (Gabriel et al., 2015).

There are two main types of diabetes (type 1 and 2). Studies have it that type 2 diabetes can be a risk factor for Alzheimer’s disease (AD), vascular dementia and others. Studies have proved that similar to diabetes, glucose is not used properly in the brain of people with AD which may be caused by nerve death which reduces the brain’s ability to interpret messages in the case of vascular dementia, brain cells due to lack of oxygen preventing brain from communicating with each other. According to Alzheimer Society of Canada, recent studies suggests that the brains of people with AD are in a diabetic state partly due to the decrease in or insensitivity to insulin (Alzheimer Society of Canada, 2023).

In this study we investigated alkaloids rich fraction of *Dacryodes edulis* leaves on the microstructure of the hippocampus and blood glucose levels of STZ-induced diabetic Wistar rats. Morphologically, there was significant weight loss in group B that was given STZ alone compared to the negative control and the experimental group (figure 1). This effect is as a result of increased lipolysis within the adipose tissue and increased breakdown of protein (Buse et al., 2011). A study by Michelle (2013) also revealed that loss of glucose in urine may cause severe weight loss associated with lipolysis and protein breakdown. Significant increase in body weight was observed in the experimental groups mostly in group D with 38% increase. The increase in body weight in the treated group may be resulted from the reduced blood sugar statuses and normoglycemia stimulated by the alkaloids fraction of *Dacryodes edulis* administered post STZ induction. According to Holmann (2007), insulin influences increase cellular update of glucose resulting in weight gain by reducing metabolic rate and preventing glycosuria (Russel-Jones et al 2007). This may be a result of reduced glycosuria and increased caloric intake from ingestion of diets with high caloric contents.

Haematological observation of the rats in diabetic control group revealed that there was significant increase in the blood glucose compared to the normal control and the post treated groups with alkaloids from *Dacryodes edulis* and metformin (figure 2). This suggests the destruction of insulin producing cells in the diabetic control group B. This result is in line with the work of Lenzen (2008) who reported that STZ damage pancreatic beta cells causing hyperglycemia and hypoinsulinemia. The groups treated with alkaloid rich fraction of *Dacryodes edulis* revealed significant decrease compared to the diabetic control groups at the end of the study. Herbal plants are mainly used to treat different kinds of ailments such as AD and epilepsy where Eru et al. (2024; 2022; 2020) demonstrated how *Talinum triangulare* and *Telfairia occidentalis* reduce the effect of neurotoxicity caused by scopolamine-induced Alzheimer’s type cognitive dysfunction rats due to its antioxidant components and guava leaf has anticonvulsant and neuroprotective properties on pentylenetetrazole (PTZ)-induced epileptic Wistar rats (Williams et al., 2025). Studies have it that fractionated components in plants such as flavonoids, saponin and glucoside of *Vernonia amydalina* reduces blood glucose level and ameliorates pancreatic damage (Ugoanyanwu et al., 2015) as a result of their antioxidant free radical scavenging activities. The microanatomical study stained with haematoxylin and eosin using light microscope revealed atrophied pyramidal cells in the pyramidal cell layer of the hippocampus as well as numerous vacuoles filled with lipid depositions as compared to the negative control group and the posttreated groups. One study has shown that normal cell death, gliosis, swollen or damage axons and myelin sheath are characteristics of chemically induced neurodegeneration (Cavanagh, 1984) This is true because the diabetic group caused by STZ in group B revealed atrophied pyramidal cells and numerous vacuoles with lipid deposits (plate 2). Normal nerves perform normal physiology but when abnormal or exposed to certain toxic substance which may result to injury on any part of the neurons, different degenerative changes may occur due to either obstruction in blood flow or causing the brain cells from not utilizing the glucose level in the brain by reducing metabolic rate and preventing glycosuria according to Russel-Jones et al., (2007).

The CA1 and CA3 subfields of the hippocampus are vulnerable to cell injury (George and Schneider*,* 1998) which is in line with the group that was given STZ alone and group C that received 1000mk/kg body weight of alkaloid-rich fraction of *Dacryodes edulis*. This study also confirm the involvement of pyramidal cells found in the pyramidal layer as degenerative changes observed in the hippocampus were predominantly evident in mostly the experimental groups treated with STZ. These degenerative changes may lead to dysfunction of the hippocampus characterized by inability to establish new long term memory. However, the distorted microanatomy of the hippocampus observed was mild in group D that received 500 mg/kg body weight (plate 4) compared to the diabetic group B (plate 2). The observed microstructure showed dose related pattern of cellular repair with group D that received 500 mg/kg alkaloid-rich fraction exhibiting the most ameliorative potentials. The above results are in line with the work of Anani et al (2024) who reported that *Dacryodes edulis* ameliorates neurodegeneration in the hippocampus, reduce lipid peroxidation and increase glutathione level in rats caused by ketamine overdose. This alkaloid-rich fraction may had neutralized the excess free radicals, protect the cells against their toxic effect as well as providing enabling environment for cells and tissue survival.

**Conclusion**

This study deduced that alkaloid-rich fraction of *Dacryode* *edulis* ameliorates cellular alterations, body weight as well as normalizing blood glucose level in STZ-induced diabetic Wistar rats.

**Disclaimer (Artificial intelligence)**

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

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