**Efficiency of *Ocimum gratisimum* as an anesthetic in the transportation of *Clariasgariepinus* fingerlings**

**Abstract**

High fish mortality caused by stressors during transportation hurts the aquaculture sector specifically fish farmers. Synthetic anesthetics made from chemical materials are banned because of safety and residual effects. This study was conducted to determine the effectiveness of *Ocimumgratissimum* (Scent leaf) powder as a natural anesthetic in transporting*Clariasgariepinus* fingerlings. A total of 300 *Clariasgariepinus*fingerlings were exposed to 0.00, 10.00, 20.00, and 30.00g/6L inclusion levels of *Ocimumgratissimum*leaf powder in 12 kegs (10litres) filled with 6L of water each. Twenty-five (25) *C. gariepinus* fingerlings were introduced into each treatment and control in a complete randomized design (four treatments and three replicates). The experimental fish were transported from Minna to Suleja at 204 km/h (to and fro). Anaesthetized fish exhibit decreased induction time with an increase in concentration of the *Ocimumgratissimum*powder while recovery time increased with an increase in concentration. The longest induction and shortest recovery times were recorded in 10g/6l (T2), while the shortest induction and longest recovery times were recorded in 30g/6l (T4). The survival rate of fingerlings decreased with an increase in the concentration of the *Ocimumgratissimum*. A high mortality rate was recorded in treatment 4 (30g/6l) due to increased concentration of experimental plants. The result obtained from the analysis of physical and chemical characteristics of water before, during, and after the transportation showed variation in most of the parameters. There was a significant difference (p<0.05) in dissolved oxygen, pH, and sulfate but no significant difference (p>0.05) was observed in Temperature and Ammonia. Due to its effectiveness and availability; *Ocimumgratissimum* is close to an ideal anesthetic. Farmers could therefore use 10.00g/6L concentration of the powder for transportation of *Clariasgariepinus*.

Keywords: Anesthetic, *Occimumgratisimum,* induction, recovery, survival, mortality

1. **Introduction**

Anesthetic is any substance that courses reversible loss of sensation or reduces sensitivity to pain and may cause unconsciousness which could be used to immobilize fish so they can be easily managed during aquaculture practices such as harvesting or capturing, transportation, sorting or grading, tagging, sampling, artificial reproduction procedures and surgery (Matin, *et al.*, 2009; Neiffer and Stamper, 2009; Javahery and Moradlu, 2012)

The handling of fish out of their natural environment always creates stress which affects their physiology and anatomy. Fishes cultured undergo a multi-phase of stress from several stressors such as transportation, grading, weighing, and stocking which are unavoidable in aquaculture (Gabriel and Akinrotimi 2011) consequently resulting in poor performance, increased vulnerability to disease, and mortality in extreme cases. To minimize the mortality induced by stress, the fish are usually sedated and immobilized by using sedatives/anesthetics before handling (King *et al.,* 2005; Ross and Ross 2008). Over the years, a large number of chemicals and agents have been applied to fish in the hopes of inducing anesthesia, with varying degrees of success, however, only a few of them have found in widespread usage sufficient to allow even a basic understanding of their effects and optimal application (Aliakbar and Hadideh, 2015).

*Ocimumgratissimum(Lamiaceae*), commonly known as “alfavaca” also known as “Efirin” in Yoruba language is native to Africa (Bayoub*et al.,* 2010). In Nigeria, the plant grows virtually in all regions. It could be found in many farms and residential and industrial areas. It grows and survives well in southwest Nigeria and can be found in backyards where it is not intentionally planted. The plant has been use for many purposes ranging from human consumption to its application in traditional medicine in Nigeria. It has a good aroma and its leaves has become a delicacy and serve as spicy for fish and meat products such as kilishi, dambu, yaji. (El-Hawarry and Correa 2012). However, many authors have reported that the plant has some chemical compounds with antimicrobial properties such as eugenol, linaol, methyl cinnamate, camphor, and thymol. It has been demonstrated in previous studies that eugenol (75-98%) isolated from *Ocimumgratissimum*is the major antimicrobial component that inhibits and kills gram-positive bacteria and fungi (Gafaar*el al*., 2010 and Kumaravel, Sivasubramanian 2010).

*Clariasgariepinus* also known as the African sharp-tooth catfish is a species of catfish of the family *clariidae*, the air-breathing catfish. The African catfish is a large eel-like fish, usually dark grey or black coloration at the back, fading to a white belly. The African catfish is second only in size to the Vundu of Zambesian waters. (Froese, Rainer, Pouly, Daniel (eds. 2014). It has an average length of 1.5 meters and weighs up to 60kg. The fish has a flat bony head, a slender body, and a broad terminal mouth with four pairs of barbells (sensory organs). Only the pectoral fins have spines. African sharp-tooth catfish have a scale-less slimy skin and accessory air-breathing organs limps which enables them to stay out of water for many hours provided a good condition prevails.

# **Materials and Methods**

# **2.1 Experimental Site**

The experiment was conducted in two phases. The first phase was the transportation of the treated fingerling from Minna to Suleja. The second phase (post-transportation phase) was an analysis of physicochemical parameters

# **2.2 Experimental Plant**

Scent leaf (*Ocimumgratissimum*) leaves was collected from farmlands and confirmed with (Agbaje 2008) plant key and conveyed to the wet and dry laboratory. Upon arrival, it was washed thoroughly to remove dirt, insect or worm. The leaves were shed dried for 72 hours, grinded and sieved into fine particles (<250µm). Sensitive weighing balance (model 20001) was used to measure (10g, 20g and 30g respectively).

**2.3 Experimental Fish**

Three hundred pieces of *Clariasgariepinus* fingerlings (a high valued aqua cultural fish species) comprised of both sexes with a mean weight of 3g were acquired and acclimatized for three days. Before the transportation, the fish were starved for 12 hours. A Complete randomized design comprising of four treatments and three replicates were used for the experiment and designated as; control (without anesthetic), 10g, 20g, and 30g of *Ocimumgratisimmum,* powder /liter respectively. 25 pieces of *Clariasgariepinus* fingerlings each were introduced in plastic gallons devoid of the control. The fish were transported to and fro Minna and Suleja at a distance of (204km). The induction and recovery time were noted.

# **2.4 Water Quality Parameter**

DO, pH, temperature, sulfate, and ammonia were analyzed before, during, and after transportation using a thermometer, dissolved oxygen meter, pH meter, ammonia using the nitrogen distillation method, and sulfate using the volumetric method.

**2.5 Statistical Analysis**

Data obtained from the experiment were subjected to one-way analysis of variance (ANOVA) using SPSS version 23 and the mean difference was determined using DUNCAN multiple range test

# **3 Results**

# **3.1 Induction and recovery time of *Clariasgariepinus* fingerlings after administration of the**

Table 1 the induction and recovery time of *C. gariepinus* fingerlings exposed to various concentrations of *Ocimumgratissimum* powder (scent leaf) presented in Table 1. The fish exhibit normal behavioral activities such as swimming, balance, opercula movement, and general movement of the tail and fins at 0.00 g/6l concentration (control). The last induction time of 148 minutes was recorded in treatment 2 (10g/6l) concentration, where there was a total loss of movement and the fish sank motionless at the bottom of the gallon.

The fastest induction time of the anesthetic agent was 25 minutes recorded in treatment 4 (30g/6l) concentration, where there was total loss of movement in the fish, while the induction and recovery time in control was 0.00 and 0.00 respectively. No mortality of the fish species was recorded during the induction time. The recovery time of *C. gariepinus*fingerlings was faster at lower concentrations 10g/6l using *Ocimumgratissimum* powder at 160 minutes and longer as the concentrations increased.

**Table 1: Induction time and recovery time in *Clariasgariepinus* fingerlings treated with varying inclusion levels of *Ocimumgratissimum* powder.**

|  |  |  |
| --- | --- | --- |
| **Concentration**  **(g/6l)** | **Overall anesthesia time**  **(min)** | **Overall recovery time**  **(min)** |
| 0.0 | 0:00 | 0:00 |
| 10 | 10:28 am (148min) | 1:08 pm(160min) |
| 20 | 9:31 am (91min) | 1:20 pm(229min) |
| 30 | 8:25 am (25min) | 1:35 pm(310min) |
|  |  |  |

**3.2 Mortality Rate of *Clariasgariepinus* Fingerlings Exposed to Various Inclusion Levels of *Ocimumgratisimum***

Table 2 depicts the mortality rate of fingerlings subjected to different inclusion levels of *Ocimumgratisimum*. The trends of mortality of the fingerlings exposed to various inclusion levels (0, 10, 20, 30g) increase with the concentration. At 30 g/6l concentration, the percentage of mortality increased significantly (P<0.05) with an increase in the *Ocimumgratisimum*.

**Table 2: Mortality Rate of *Clariasgariepinus* Fingerlings Exposed to Various Inclusion level of *Ocimumgratisimum***

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Mortality rate %** | | | | |  |
| **Time (mins)** | **T1(0g)** | **T2 (10g)** | **T3 (20g)** | **T4 (30g)** | |
| 0 | 0 | 0 | 0 | 0 | |
| 30 | 0 | 0 | 0 | 0 | |
| 60 | 0 | 0 | 0 | 0 | |
| 90 | 0 | 0 | 0 | 0 | |
| 120 | 0 | 0 | 0 | 0 | |
| 150 | 0 | 0 | 0 | 0 | |
| 180 | 0 | 0 | 0 | 11.67 |  |
| 210 | 0 | 0 | 26.67 | 16.67 |  |
| 240 | 0 | 0 | 26.67 | 48.33 |  |
| 270 | 0 | 0 | 41.67 | 66.67 |  |
| 300 | 0 | 0 | 58.33 | 66.67 |  |
| 330 | 6.67 | 1.67 | 68.33 | 73.33 |  |
| 360 | 8.33 | 1.67 | 68.33 | 100 |  |
| 390 | 8.33 | 1.67 | 68.33 | 100 |  |

# **3.3 Survival Rate of *Clariasgariepinus* Fingerlings Exposed to Various Inclusion level of *Ocimumgratisimum***

Table 3 shows the survival rate of fingerlings treated with different inclusion levels of *Ocimumgratisimum*as an anesthetic. The survival rate of fingerling exposed to various inclusion levels (0, 10, 20, 30g/L) became high at 10g/6l and the percentage of survival decreased significantly (p>0.05) with an increase in the *Ocimumgatisimum*concentration.

**Table 3:Survival Rate of *Clariasgariepinus* Fingerlings Exposed to Various Inclusion level of *Ocimumgatisimum* (0g, 10g, 20g and 30g)**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Survival rate %** | | | | | |
| **Time (min)** | **T1(0g)** | **T2 (10g)** | **T3 (20g)** | **T4 (30g)** | |
| 0 | 100 | 100 | 100 | 100 | |
| 30 | 100 | 100 | 100 | 100 | |
| 60 | 100 | 100 | 100 | 100 | |
| 90 | 100 | 100 | 100 | 100 | |
| 120 | 100 | 100 | 100 | 100 | |
| 150 | 100 | 100 | 100 | 100 | |
| 180 | 100 | 100 | 100 | | 88.33 |
| 210 | 100 | 100 | 73.33 | | 88.33 |
| 240 | 100 | 100 | 73.33 | | 51.67 |
| 270 | 100 | 100 | 58.33 | | 33.33 |
| 300 | 100 | 100 | 41.67 | | 33.33 |
| 330 | 93.33 | 98.33 | 31.67 | | 26.67 |
| 360 | 91.67 | 98.33 | 31.67 | | 0 |
| 390 | 91.67 | 98.33 | 31.67 | | 0 |

# **3.4 Variation of Physicochemical Parameter of Water for before, during, and after Transportation with Different Inclusion Levels of *Ocimumgratisimum* (0 g, 10 g, 20 g, 30 g)**

Figure 1 shows the physico-chemical parameters of time variation of water with different inclusion levels of *Ocimumgratisimum* (0g, 10g, 20g, 30g). There was no significant difference in water temperature (p>0.05) across all treatments. However, the highest mean value was recorded during transportation (31.22±0.87) while the lowest was recorded before transportation (30.29±0.52). Significant difference (p<0.05) was recorded in pH level among all the treatments during transportation. The highest mean value was recorded before transportation (6.44±0.12) while the lowest was recorded during transportation (5.96±0.29). There was a significant difference (p<0.05) in dissolved oxygen (mg/l). However, the highest mean was recorded before transportation (5.59±1.56) while the lowest was recorded after transportation (2.71±2.34). The Ammonia level also shows a significant difference (p<0.05) with the highest recorded during transportation (0.04±0.02) and the lowest after transportation (0.02±0.01). However, the variation differs significantly. For sulfate, the result does not differ significantly (p>0.05). However, the highest value was recorded during transportation (14.11±10.27) while the lowest was recorded after transportation (11.08±2.80).

**3.4 Physico-chemical parameters of Water with Different Inclusion Levels of *Ocimumgratisimum***

Table 4 depicts the Mean Value of Physical and chemical characteristics of Water with -Different Inclusion Levels of *Ocimumgratisimum* (0g, 10g, 20g, and 30g). Throughout the experiment, the water temperature (°C), did not show any variation with increased concentration of *Ocimumgratisimum* in all the treatments. However, the highest mean value was recorded in T1 (31.96±1.19) which serves as the control, and the lowest mean value was recorded in T4 (28.75±5.93) with 30g of *Ocimumgratisimum*. There were significant differences (p<0.05) in the values obtained for pH. The highest mean value was recorded in T1 (6.38±0.18) and the lowest in T4 (5.93±0.47). DO decreases as the concentration of *Ocimumgratisimum*increases. The values obtained for dissolved oxygen was significantly higher (p<0.05) in T1 (6.55±1.40 mg/l) while the lowest was recorded in T4 (1.62±2.30). Thus, the values differ significantly (P<0.05) compared with the control. For Ammonia, there was no significant difference (p>0.05) in all the treatments, but the highest mean value was recorded in T1 (0.04±0.03) and the lowest mean was recorded in T4 (0.02±0.01). Significant differences was observed in the values of sulphate (P<0.05) with the highest mean score recorded in T3 (16.84±10.78) and the lowest recorded in T1 (6.62±1.05).

**Table 4: Mean Value of Physicochemical parameters of Treatment Variation of Water with Different Inclusion Level of *Ocimumgratisimum* (0g, 10g, 20g, and 30g)**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Parameters** | **TI** | **T2** | **T3** | **T4** |
| Temperature (°C) | 31.96±1.19a | 31.21±1.03a | 30.93±0.80a | 28.75±5.93a |
| pH | 6.38±0.18a | 6.22±0.20ab | 6.02±0.34b | 5.93±0.47b |
| D O(mg/L) | 6.55±1.40a | 4.24±1.03b | 2.38±1.90c | 1.62±2.30c |
| Ammonia(mg/L) | 0.04±0.03a | 0.04±0.01a | 0.03±0.01a | 0.02±0.01a |
| Sulphate(mg/L) | 6.62±1.05b | 12.27±1.95a | 16.84±10.78a | 12.93±0.77a |

Mean with different superscript on the same row are significantly different (P<0.05)

# **4 Discussion**

In aquaculture, transporting live fish is a challenge. Many factors influence the success of fish transport among which includes the duration of transport, water parameters, the size, density, and physical condition of the fish and the duration of the depuration before fish transport (Golombieski*et al*., 2003; Carneiro *et al*., 2009; Becker *et al.,* 2012). This study revealed the influence of *Ocimumgratisimum* as an anesthetic on the induction, recovery, survival, and mortality of fish and the physicochemical characteristics of water. Cooke *et al*. (2004) reported highly sedated fish, lose perception, balance, and normal swimming. This conforms to this study which the fingerlings exhibit loss of balance, opercula beat, and normal swimming in treatments three and four due to an increase in concentration of *Ocimumgratisimum*.

The sedation time decreases with increasing concentration of *Ocimumgratisimum* powder. Also, there was a direct relationship between recovery time and concentrations of *Ocimumgratisimum*. As the concentration increases, the transition time to induction decreases, and recovery time also increases. . This result is in line with the findings of Kamble, *et al*. (2014) on reduction in induction time with increased concentration of clove oil on *Cyprinus carpio*. A similar result was recorded by Agokei & Adebisi (2010) when Nile tilapia (*O. niloticus*) was exposed to different concentrations of aqueous and alcoholic extract of Tobacco. Results from several studies indicate that the induction times under anesthetics may increase, decrease or remain unchanged with the concentrations of the anesthetics, species and size of exposed fish (Walsh and Pease, 2002). In a study using juvenile angel fish *Pterophyllumscalare*, Mitjana*et al*. (2014) observed that at the highest doses 80mg/l for clove oil and 140mg/l for MS 222, the induction time remained unchanged. Transportation involves capture, loading, transport, unloading and stocking and so can induce large stress responses that can affect fish over a prolonged recovery period (Ashley, 2007; Sneddon *et al*., 2016; Souza *et al.,* 2018; Cogliati*et al*., 2019). The trend between the survival and the mortality rate is directly related. At the initial stage, the survival rate among all treatments was 100%. Mortality was recorded after 150 minutes in T3 and T4. Subsequently, at 330 minutes, mortality was recorded in T1 and T2, High number of mortality was recorded in T4 while the lowest mortality was recorded in T2. This may be due to the increase in concentration of *Ocimumgratisimum* and metabolic activities that took place during the transportation of fish. This is in consonant with the report of (Soto and Burhanuddin, 1995; Munday and Wilson, 1997; Iversen *et al*., 2003; Hisano*et al.,* 2008; Inoue *et al*., 2011; Javahery*et al.,* 2012). The survival rates of experimental fish decreased sharply at higher concentrations of anesthetics. Light sedation is more appropriate for transport procedures because it allows fish to maintain balance, swimming activity, and respiration (Cooke *et al*., 2004). Measuring the water quality parameters is vital for the efficiency of fish transportation (Sampaio and Freire, 2016). The temperature of the water remained within the comfortable value range for the species, according to Silva and Fujimoto (2015). Water temperature was similar across treatments within the 4hrs transportation periods. But the value was reduced in T4. There is a direct relationship between water-dissolved oxygen and *Ocimumgratisimum*. DO value reduces significantly with an increase in concentration of *Ocimumgratisimum.* The decrease in DO with an increase in concentration of *Ocimumgratisimum* in this study confirms the report of Audu*et al*. (2013) that DO increases with an increase in concentration of *C. sativa*. The dissolved oxygen level decreased below the optimum range, the PH value was acidic and sulfate also increased beyond the optimum range, this made the mortality of experimental fish to be high in T3 and T4.

In turn, pH exhibited different behaviors, with pH falling during transportation. At the end of the transportation experiment, pH was significantly higher in the control group as compared to the fish anesthetized with 30 g/l *Ocimumgratisimum*. INOUE *et al.* (2005) reported that pH decreased after 4 h of transportation of juvenile matrinxã (*Bryconcephalus, Gunther*) anesthetized with 5 mg L-1 clove oil, probably due to the increase in CO2 values. The reduction in water pH after transport indicates an increase in CO2. Ammonia levels decrease significantly with an increase in concentration of*Ocimumgratisimum*. This result confirms the report of Singh *et al.* (2004) that the Indian carp fry *C. catla, L. rohita, and C. mrigala* exposed to 2-phenoxyethanol (0.09 mg L-1) exhibited decreased NH3 excretion. The sulfate level in T1 was lower than those in the other treatment. There is a direct relationship between sulfate level and *Ocimumgratissimum*, as the concentration of *Ocimumgratissimum* increases the sulfate level also increases. An increase in deterioration of water quality promotes more stress to fish and, consequently, mortality (Luz *et al.,* 2013; Moreira *et al*., 2015). Anesthetics are beneficial for decreasing fish arousal and avoiding physical injuries during transport. Light sedation is more appropriate for transport procedures because it allows fish to maintain balance, swimming activity, and respiration (Cooke *et al.,* 2004).

# **5 Conclusion**

The finding in this study revealed that the use of *Ocimumgratissimum*at light concentration has sedation potential on transportation of *Clariasgariepinus*fingerlings, particularly on short distance. However; high concentration of this plant extract has effects on the survival of *Clariasgariepinus*fingerlings. Further research should be conducted to evaluate the metabolic, hematology and histology impact of *Ocimumgratissimum*powder on fish transportation

# **References**

Agokei OE, Adebisi AA. (2010). Tobacco as an anesthetic for fish handling procedures. *Journal of Medicinal Plants Research.* Vol 4 (14) pg. 1396-1399

Aliakbar, Y. and Hadideh, N.J. (2015). Determination of MS222 Anesthetic Dose and Its Effect on Liver and Kidney Tissues of Barbusgrypus. *2nd Int'l Conference on Advances in Environment, Agriculture and Medical Sciences Antalya Turkey* (ICAEAM'15),

Audu BS, Adamu KM, Ufodike EBC. (2013) Behavioural response and opercula ventilation rate of Nile Tilapia (Oreochromis niloticus) fingerlings after anaesthesia with aqueous crude leaf extract of marijuana (Cannabis sativa). *Applied Science Research Journal*. Vol 1(2) pg. 66-77. 22

Bayoub K, Baibai T, Mountassif T, Retmane A, Soukri A (2010) Antibacterial activities of the crude ethanol extracts of medicinal plants against Listeria monocytogenes & some other pathogenic strains. *Afr J Biotechnol*. Vol 9. Pg. 4251-4258.

Becker, AG., TV. Parodi, CG. Heldwein, CC. Zeppenfeld, B. M. Heinzmann& B. Baldisserotto. (2012). Transportation of silver catfish, Rhamdiaquelen, in water with eugenol and the essential oil of Lippia alba. *Fish Physiology and Biochemistry*. Vol 38. pg. 789-796.

Carneiro, PC. F., P. H. S. Kaiseler, E. A. C. Swarofsky& B. Baldisserotto. (2009). Transport of jundiáRhamdiaquelen juveniles at different loading densities: water quality and blood parameters. *Neotropical Ichthyology.* Vol 7 pp. 283-288.

[Cooke,](http://refhub.elsevier.com/S0044-8486(19)31452-8/rf0075) SJ., Suski, CD., Ostrand, KG., Tufts, BL., Wahl, DH., (2004). Behavioural an[d](http://refhub.elsevier.com/S0044-8486(19)31452-8/rf0075) physiological assesment of low

concentrations of clove oil anaesthetic for handlin[g](http://refhub.elsevier.com/S0044-8486(19)31452-8/rf0075) and transporting largemouth bass [(](http://refhub.elsevier.com/S0044-8486(19)31452-8/rf0075)Micropterus salmoide[s](http://refhub.elsevier.com/S0044-8486(19)31452-8/rf0075)). Aquaculture 239, 50[9–529 1016/j.aquaculture.2004.06.028](http://refhub.elsevier.com/S0044-8486(19)31452-8/rf0075).

El-Hawarry WH (2012) Biochemical & non-specific immune parameters of healthy Nile tilapia (Oreochromis niloticus), Blue tilapia (Oreochromis aureus) & their interspecific hybrid (male O. aureus × female O. niloticus) maintained in semiintensive culture system. *OJAFR* Vol 2. Pg.84-88.

Froese, Rainer,Pouly, Daniel (eds.) (2014) “ Clariasgariepinus” in fish Base. version

Gabriel UU, Akinrotimi OA. (2011) Management of stress in fish for sustainable aquaculture development. *Researcher*. Vol 3. Pg.28-38.

Gafaar AY, El-manakhly EM, Soliman M.K., Soufy H, Monas Z, et al. (2010) Some pathological, biochemical &Haematological investigation on Nile tilapia (Oreochromis niloticus) following chronic exposure to edifenphos pesticide. *J Am Science* Vol 6. Pg. 542-551. Hema 12 R, Kumaravel S, Sivasubramanian C GC-MS study on the potentials of Syzygiumaromaticum. *Researcher*. Vol 2 pg. 1-4.

Golombieski, J.I., LVF. Silva, B. Baldisserotto& J. H. S. da Silva. (2003). Transport of silver catfish (Rhamdiaquelen) fingerlings at different times, load densities, and temperatures. *Aquacultur*. Vol 216. Pg. 95-102.

Hisano, H., Ishikawa, MM., Ferreira, RÁ., Bulgarelli, A.L., Costa, T.R. and Padua, S.B. (2008). Tempo de indução e de recuperação de douradosSalminusbrasiliensis (Cuvier, 1816), submetidos a diferentesconcentrações de óleo de cravo Eugenia sp. *Acta Scientiarum. Biological Science. vol.* 30, no. 3, pp. 303-307.

Inoue, LAKA., Afonso, LOB., Iwama, G.K. and Moraes, G. (2005). Effects of clove oil on the stress response of matrinxã (Bryconcephalus) subjected to transport. *Acta Amazonica.* vol. 35, no. 2, pp. 289-295.

Inoue, LA.KA., Boijink, C.L., Ribeiro, P.T., Silva, A.D. and Affonso, E.G. (2011). Avaliação de respostasmetabólicas do tambaquiexpostoao eugenol embanhosanestésicos. *Acta Amazonica*vol. 41, no. 2, pp. 327-332.

Iversen, M., Finstad, B., Mckinley, R.S. and Eliassen, R.A., (2003). The efficacy of metomidate, clove oil, Aqui-STM and Benzoak® as anaesthetics in Atlantic salmon (Salmo salar L.) smolts, and their potential stress-reducing capacity. *Aquaculture,* vol. 221, no. 1-4, pp. 549-566.

Javahery, S., Nekoubin, H. and Moradlu, A.H. (2012). Effect of anaesthesia with clove oil in fish. *Fish Physiology and Biochemistry*. Vol. 38, no. 6, pp. 1545-1552.

Kamble AD. Saini VP, Ojha ML. (2014). The efficacy of clove oil as anaesthetic in Nile tilapia (Oreochromis niloticus) and its potential metabolism reducing capacity. *International Journal of Fauna and Biological Studies.* Vol 1(6) pg. 01-06

King WV, Hooper B, Hillsgrove S, Benton C, Berlinsky D (2005.) The use of clove oil, metomidate, tricaine methanesulphonate and 2-phenoxyethanol for inducing anaesthesia and their effect on the cortisol stress response in black sea bass (Centropristisstriata L.). *Aquaculture Research.* Vol 36 pg. 1442-1449.

Luz, R.K., Costa, LS., Ribeiro, P.A.P., Silva, R.F., Rosa, P.V., (2013). Influência do tempo de transporte para juvenis de

pacamã (Lophiosilurusalexandri). Arq. Bras. Med. Vet. Zootec. 65, 1895–1898. [https://doi.org/10.1590/S0102-09352013000600044.](https://doi.org/10.1590/S0102-09352013000600044)

Matin SMA, Hossain MA, Hashim MA. (2009) Clove oil anaesthesia in singhi (Heteropneustesfossilis) and lata (*Channa punctatus*) fish. *The Bangladesh Veterinarian.* Vol 26 pp 68-73

Mitjana, O., C. Bonastre, D. M. V. Falceto, J. Esteban, A. Josa, E. Espinosa (2014). The efficacy and effect of repeated exposure to 2phenoxyethanol, clove oil and tricaine methanesulphonate as anesthetic agents on juvenile Angelfish (*Pterophyllumscalare*). *Acta Vet Brno* Vol 74. pg. 139-146

Moreira, AG., Coelho, AA., Albuquerque, LF., Moreira, RT., Farias, WR., (2015). Eugenol effect as a mitigated agent of

stress in the transport of Nile tilapia juveniles. Pesqui. Vet. Bras. 35 (11), 893–898. [https://doi.org/10.1590/S0100-736X2015001100004.](https://doi.org/10.1590/S0100-736X2015001100004)

Munday, PL. and Wilson, SK., (1997). Comparative efficacy of clove oil and other chemicals in anaesthetization of*Pomacentrusamboinensis*, a coral reef *fish. Journal of Fish Biology,* vol. 51, no. 5, pp. 931-938.

Neiffer, DL. and Stamper, MA. (2009). Fish sedation, anesthesia, analgesia, and euthanasia: considerations, methods, and types of drugs. *Ilar Journal.* Vol 50 (4) pg. 343-360.

Ross, LG. and B. Ross (2008). Anesthetic and sedative techniques for aquatic animals. *3rd ed. Blackwell Publishers Oxford,* UK. 222pp.\

Sampaio, F.D., Freire, C.A., (2016). An overview of stress physiology of fish transport: changes in water quality as a function of transport duration. Fish Fish. 17 (4), 1055–1072. [https://doi.org/10.1111/faf.12158.](https://doi.org/10.1111/faf.12158)

Singh, RK., VR. Vartak, AK. Balange& M. M. Ghughuskar. (2004). Water quality management during transportation of fry of Indian major carps, Catlacatla (Hamilton), Labeorohita (Hamilton), and *Cirrhinusmrigala*(Hamilton). *Aquaculture.* Vol 235 pp. 297-302.

Soto, CG. and Burhanuddin, G. (1995). Clove oil as a fish anaesthetic for measuring length and weight of rabbitfish (Siganuslineatus). *Aquaculture.* Vol. 136, no. pp. 149-152.

Walsh CT. and BC. Pease (2002). The use of clove oil as an anesthetic for the long-finned eel, Anguilla reinhardtii (Steindachner). *Aquaculture Research.* Vol 33. Pp. 627-635.