**Effect of 6-benzylaminopurine treatments on postharvest quality and storage life of Jamun (*Syzygiumcumini*Skeels) fruit**

**ABSTRACT**

*Syzygiumcumini* (Skeels.), commonly known as Jamun, Java plum, or black plum, is a bioactive-rich fruit recognized for its high concentrations of antioxidants, vitamins, and phytochemicals, including hydrolysable tannins, flavonoids, anthocyanins, gallic acid, and quercetin. Despite its nutritional and pharmacological significance, the fruit exhibits a notably short postharvest shelf life of approximately 3–4 days under ambient conditions. This rapid perishability hinders effective storage, transportation, and commercialization, often resulting in considerable postharvest losses. This study evaluated the impact of 6-benzylaminopurine (BAP) on extending shelf life and preserving the quality of fully ripe fruit under cold storage. Fruits were dipped in BAP solutions (0.5, 1.0, and 1.5 mM), air-dried, and stored at 7 ± 1°C in low-density polyethylene (LDPE) bags. Two control groupsdistilled water-treated fruits in open LDPE bags and untreated fruits in sealed LDPE bagswere also included. The result demonstrated that the 1.5 mM BAP treatment was most effective, significantly reducing spoilage and maintaining key physicochemical attributes. These findings demonstrate the potential of BAP, particularly at 1.5 mM, as a postharvest treatment to enhance storability, reduce losses, and improve the commercial viability of jamun. By extending shelf life and minimizing losses, this approach can improve the availability of jamun to consumers and support its commercial viability. The results underscore the potential of BAP as a postharvest treatment to address storage challenges for perishable tropical fruits like jamun, contributing to reduced food waste and enhanced marketability.

**Keywords:** Jamun, Bio-active compounds, 6-benzylaminopurine, Low-density polyethylene

**Introduction**

The Indian blackberry (*Syzygiumcumini*Skeels), commonly referred to as black plum or jamun, is a subtropical, underutilized fruit species native to the Indian subcontinent and belongs to the family Myrtaceae. The fruit is characterized by a deep purple exocarp and a purplish-pink to white mesocarp, enclosing a single, hard seed (endocarp). Jamun is a rich reservoir of essential nutrients and bioactive phytochemicals, including vitamins (ascorbic acid, retinol, niacin), minerals (calcium, iron, magnesium, phosphorus, potassium, sodium), sugars, amino acids, and a wide range of secondary metabolites (de Carvalho Tavares et al., 2016). Among the key bioactive constituents in jamun are anthocyanins, tannins, carotenoids, phenolic acids, flavonols, and flavanonols, which contribute to its recognized nutraceutical and therapeutic properties. The predominant anthocyanin pigments include malvidin and the 3,5-O-diglucosides of delphinidin, petunidin, and cyanidin (Raza et al., 2015). Additionally, compounds such as gallic acid, ellagic acid, and tannins—mainly present in the seeds—have been reported to exhibit antidiuretic activity. Other phytochemicals including lupeol, stigmasterol, and β-sitosterol are known for their significant anti-inflammatory and antinociceptive properties, suggesting the fruit’s potential for pharmacological applications (Arya et al., 2017). Previous studies have also indicated the beneficial effects of jamun in regulating blood pressure (Bhargava et al., 1968), and mitigating heart, liver, and pulmonary disorders (Raza et al., 2015). Importantly, its hypoglycemic activity has been well documented, making it particularly valuable in the dietary management of diabetes mellitus (Koley et al., 2011; Ayyanar et al., 2013).

Despite its high nutritional and therapeutic value, *S. cumini* suffers from severe postharvest losses. In India, it is estimated that approximately 0.5 million tonnes (MT) of jamun fruit are lost annually due to its extreme perishability and limited shelf life of only 2–3 days under ambient conditions (Patil et al., 2012). The fruit is harvested predominantly during the monsoonal season (mid-June to mid-August), resulting in a short harvesting window of about one month. This temporal concentration of harvest leads to market glut, forcing producers to sell their produce at reduced prices (Rai et al., 2011). The fruit's rapid postharvest deterioration, primarily due to microbial and physiological spoilage, further complicates long-distance transportation and limits market accessibility, ultimately aggravating economic losses.

To reduce postharvest spoilage and enhance shelf life, numerous approaches have been investigated, particularly those involving plant growth regulators. Some of the recent strategies include treatments with calcium chloride and gibberellic acid (Ayar et al., 2011), the use of antioxidant-rich edible coatings (Baraiya et al., 2015), modified atmosphere packaging techniques (Rai et al., 2011), and the application of salicylic acid in combination with chitosan coatings to slow down senescence (Saurabh et al., 2019).

One such compound, **6-benzylaminopurine (BAP)**, is a synthetic cytokinin known for its capacity to delay senescence and improve postharvest quality. BAP has been demonstrated to reduce weight loss, spoilage, and undesirable changes in color, texture, and biochemical composition in a range of horticultural produce (Siddiqui et al., 2015; Zhang et al., 2018). Its effectiveness in preserving physicochemical and functional quality attributes during storage has been substantiated in several fruit and vegetable crops (Palma et al., 2019). Therefore, this study investigates the effectiveness of 6-benzylaminopurine (BAP) in delaying senescence and maintaining the postharvest quality of jamun under low-temperature storage, focusing on its impact on physicochemical and functional attributes.

**Materials and methods**

Jamun fruits at the ripe stage were harvested from approximately 20-year-old trees in July 2019 and promptly transferred to the Postharvest Laboratory, Department of Horticulture, Institute of Agricultural Sciences, Banaras Hindu University. The selection criteria for the fruits included uniformity in size, shape, color, and maturity, as well as the absence of blots, pests, diseases, and mechanical damage, to ensure that only healthy fruits were used for research. The fruits were disinfected with a 2% sodium hypochlorite solution for 2 minutes, air-dried, and then treated with 6-benzylaminopurine at concentrations of 0.5 mM, 1.0 mM, and 1.5 mM for 5 minutes. Control fruits were treated with distilled water under the same conditions as the treated fruits. The fruits were air-dried at room temperature, packaged in LDPE bags, and stored at 7±1°C. Quality attributes were assessed every 5 days during storage.

**Weight loss (WL)**

The weight loss of jamun was quantified using a gravimetric approach, where the percentage weight loss was calculated using the formula: WL (%) = [(IW - FW) / IW] × 100, with IW representing the initial fruit weight and FW representing the fruit weight on the sampling day.

**Decay loss**

This approach enabled an accurate assessment of postharvest decay loss. The incidence of decay was evaluated by quantifying the number of fruits exhibiting symptoms of decay, and the result was expressed as a percentage of the total fruit sample observed, calculated using the formula: Decay Loss (%) = (Spoiled Fruits / Total Fruits) × 100, allowing for a precise determination of decay loss.

**Malondialdehyde content**

MDA content in jamun fruit was determined following the protocol described by Zheng and Tian (2006).Fruit tissue (0.5 g) was homogenized in 5 mL of 5% TCA and centrifuged at 10,000 rpm for 15 min at 4°C. The supernatant (2 mL) was mixed with 2 mL of 5% TCA containing 0.6% TBA, heated at 90°C for 30 min, then rapidly cooled. Absorbance was measured at 450 and 532 nm, and MDA content was expressed as nmol/g FW.

**Total soluble solids**

Total soluble solids (TSS) in fruits during storage were measured using a digital refractometer (Atago, Tokyo, Japan), and the results were expressed in degrees Brix (°Brix). Titratable acidity was determined via titration method, following the standard protocol outlined by the Association of Official Analytical Chemists (AOAC, 2000).

**Total anthocyanins content**

The extraction of anthocyanins from 0.05 g of peel and 0.5 g of pulp was performed using 10 mL of ethanolic HCl, according to the standardized procedure described by Lees and Francis (1972). After overnight storage at a low temperature, the sample was centrifuged at 10,000 rpm for 10 minutes, and the absorbance was recorded at 535 nm. The total anthocyanin content was expressed as mg/100 g FW.

**Ascorbic acid content**

Jones and Hughes (1983) developed a procedure for determining ascorbic acid content in fruits. In this method, 0.2 g of fruit sample was mixed with a 3% w/v metaphosphoric acid solution, and the volume was adjusted to 20 mL. A 10 mL aliquot was taken from the solution and titrated with 2,6-dichlorophenol indophenol dye until a pink endpoint was reached. The ascorbic acid content of the fruit was expressed as milligrams per 100 grams of fresh weight (mg/100 g FW).

**Total phenolics content**

Total phenolic content was quantified following the method of Singleton et al. (1999). A 0.5 g fruit sample was mixed with 10 mL of 80% ethanol. An aliquot of 100 µL of the extract was mixed with 2.9 mL distilled water and 0.5 mL of 1 N Folin–Ciocalteu reagent. After 3 minutes, 2 mL of 20% sodium carbonate solution was added, and the mixture was incubated for 90 minutes at room temperature. Absorbance was measured at 760 nm, and results were expressed as mg gallic acid equivalents per 100 g fresh weight (mg GAE/100 g FW).

**Total flavonoids content**

Zhishen et al. (1999) outlined a procedure for assessing the total flavonoid content in jamun fruit, where 0.5 g of fruit sample was mixed with 10 mL of methanol and then centrifuged at 10,000 rpm for 10 minutes. A precise combination of 1 mL of the supernatant, 4 mL of distilled water, and 0.3 mL of 5% sodium nitrite solution was prepared in a test tube, allowing for further analysis. After 5 minutes, 0.3 mL of 10% aluminum chloride hexahydrate (AlCl3·6H2O) solution was added to the test tube. The solution was then incubated at room temperature for 6 minutes, followed by the addition of 1 N sodium hydroxide (NaOH) solution. The final volume was adjusted to 10 mL with distilled water. The analytical procedure involved measuring the absorbance at 510 nm, enabling the calculation of the total flavonoid content of jamun fruit as mg RE/100 g FW, providing a precise measurement of flavonoid compounds.

**Total antioxidant capacity**

Total antioxidant capacity was assessed using the CUPRAC (Cupric Ion Reducing Antioxidant Capacity) assay, following the protocol described by Apak et al. (2008), which enables reliable quantification of the fruit's antioxidant potential. The reaction mixture consisted of 100 µL of 80% ethanolic extract of fruit sample, 1 mL each of 10 mM copper(II) chloride, 7.5 mM neocuproine, ammonium acetate buffer (pH 7.0), and distilled water. After incubation at room temperature for 30 minutes, absorbance was measured at 450 nm. Results were expressed as micromoles of Trolox equivalents per gram of fresh weight (µmol TE/g FW).

**Analysis of data**

The study was designed as a completely randomized factorial experiment comprising five treatment groups, each replicated three times. Data for all measured parameters are presented as mean values. Statistical differences among treatment means were assessed using Tukey’s Honest Significant Difference (HSD) test. A significance level of p ≤ 0.05 was established to determine statistically significant differences. All statistical analyses were performed using SAS software version 9.2 (SAS Institute Inc., Cary, NC, USA) for data interpretation.

**Results and discussion**

**Weight loss**

In the present study, a progressive increase in fruit weight loss was observed across all treatments throughout the storage duration.Table 1. shows the effect of different treatments on physiological loss in weight (PLW%) of Jamun fruits during 30 days of cold storage. Fruit stored under open conditions (Control Open) exhibited the highest weight loss, reaching 27.32% at 30 DAS. In contrast, sealing (Control Sealed) significantly reduced PLW to 9.10% at 30 DAS. Among the BAP (6-benzylaminopurine) treatments, 1.5 mM BAP was most effective, resulting in the lowest weight loss of 4.67% at 30 DAS, followed by 1.0 mM (5.61%) and 0.5 mM (7.30%). These results indicate that BAP, particularly at 1.5 mM, effectively reduces PLW by delaying senescence and maintaining fruit integrity during storage.Fruits and vegetables often experience weight loss after harvest during storage, which can significantly reduce their consumer appeal and marketability. The primary cause of weight loss in jamun is the rapid loss of water due to its thin skin, resulting in fruit shrinkage during postharvest storage. Additionally, weight loss leads to a loss of texture, compromising visual appeal and ultimately reducing consumer acceptance. The role of BAP in reducing weight loss of fruit can be related to a strengthening of cell wall (Massolo*et al*., 2014) and reduction in respiration rate (An *et al*., 2006). The effectiveness of BAP in reducing weight loss during postharvest storage has also been reported in other fruits such as Mangosteen (Efendi and Hermawati, 2010), cherimoya (Franco-Mora *et al*., 2015) and apricot (Canli *et al*., 2014) fruit.

**Decay loss**

Jamun fruits deteriorate rapidly after harvesting due to spoilage-causing microorganisms that enter through the thin skin and stem-end. Their delicate nature also makes them prone to mechanical damage, increasing susceptibility to infection. Table 1. shows that BAP treatments effectively reduced disease incidence during low-temperature storage. The data reveals a progressive increase in decay loss in Jamun fruits during storage, particularly after 15 days. The Control Open samples exhibited the highest decay loss, reaching 34.30% at 30 DAS, whereas the Control Sealed samples demonstrated a lower yet significant decay of 20.30%. BAP treatments were effective in reducing decay, with 1.5 mM BAP yielding the most favorable results, showing only 9.87% decay at 30 DAS. This was followed by 1.0 mM (14.48%) and 0.5 mM (19.35%). Notably, all treatments exhibited zero decay up to 15 DAS. These findings suggest that BAP, particularly at 1.5 mM, significantly delays the onset of decay and extends shelf life.This study showed that postharvest BAP application significantly reduced spoilage in jamun fruit. BAP may limit pathogen attack by delaying cell wall degradation and softening (Massolo et al., 2014) and enhancing the expression of defense-related proteins and genes (Ge et al., 2011; Kachrooand Robin, 2013). Similar effects were observed in litchi fruit (Zhang et al., 2018).

**Malondialdehyde content**

Malondialdehyde, produced during lipid peroxidation by reactive oxygen species, serves as a common marker of fruit senescence, ripening, and tissue damage.In this study, malondialdehyde content increased progressively with storage duration. Table 1. presents the effects of BAP treatments on malondialdehyde (MDA) content, a marker of lipid peroxidation and membrane damage, in Jamun fruits during storage. The Control Open samples exhibited the highest MDA accumulation, reaching 9.73 nmol/g at 30 DAS, indicative of increased oxidative stress. In contrast, Control Sealed fruits demonstrated slightly lower MDA levels (9.20 nmol/g). BAP-treated fruits exhibited a significant reduction in MDA content, with 1.5 mM BAP proving most effective, limiting MDA to 7.63 nmol/g at 30 DAS. BAP concentrations of 1.0 mM and 0.5 mM also reduced MDA levels to 8.51 and 8.69 nmol/g, respectively. These findings underscore the protective role of BAP, particularly at 1.5 mM, in mitigating oxidative stress and preserving membrane stability during storage.This indicated that the treatment was highly effective in delaying senescence and maintaining bettermembrane integrity of the *jamun* fruit during storage.The results of this study demonstrate that BAP treatments significantly reduce malondialdehyde accumulation in fruits by enhancing ROS scavenging capacity, thereby maintaining membrane integrity during storage.

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
|  | T**reatments** | **Days after storage (DAS)** | | | | | |
| **5 DAS** | **10 DAS** | **15 DAS** | **20 DAS** | **25 DAS** | **30 DAS** |
| **Physiological loss in weight (%)** | **Control Open** | 2.53 ± 0.12 a | 5.07 ± 0.25 a | 11.4 ± 0.4 a | 17.2 ± 0.52 a | 23.3 ± 0.41 a | 27.32 ± 0.80 a |
| **Control Sealed** | 0.59 ± 0.08 bc | 1.35 ± 0.18 b | 2.6 ± 0.10 b | 5.4 ± 0.33 b | 6.64 ± 0.18 b | 9.10 ± 0.26 b |
| **BAP(0.5 mM)** | 0.52 ± 0.06 bc | 0.92 ± 0.01 b | 1.23 ± 0.02 c | 4.32 ± 0.11 bc | 4.88 ± 0.19 c | 7.30 ± 0.19 bc |
| **BAP(1.0 mM)** | 0.83 ± 0.02 b | 1.13 ± 0.09 b | 1.63 ± 0.03 bc | 3.08 ± 0.29 cd | 3.59 ± 0.10 d | 5.61 ± 0.03 cd |
| **BAP(1.5 mM)** | 0.36 ± 0.00 c | 0.94 ± 0.04 b | 1.36 ± 0.17 c | 2.78 ± 0.11 d | 3.49 ± 0.14 d | 4.67 ± 0.12 d |
| **Decay loss (%)** | **Control Open** | 0 | 0 | 0 | 11.18 ± 0.68 a | 17.29 ± 0.44 a | 34.30 ± 2.12 a |
| **Control Sealed** | 0 | 0 | 0 | 7.76 ± 0.20 b | 9.9 ± 0.37 c | 20.30 ± 1.12 b |
| **BAP(0.5 mM)** | 0 | 0 | 0 | 5.45 ± 0.45 c | 13.84 ± 0.63 b | 19.35 ± 0.68 b |
| **BAP(1.0 mM)** | 0 | 0 | 0 | 5.31 ± 0.26 c | 9.77 ± 0.25 c | 14.48 ± 0.75 c |
| **BAP(1.5 mM)** | 0 | 0 | 0 | 2.12 ± 0.32 d | 5.91 ± 0.48 d | 9.87 ± 0.35 d |
| **Malondialdehyde content (nmol/ g)** | **Control Open** | 2.82 ± 0.14 a | 3.93 ± 0.20 a | 5.00 ± 0.09 a | 6.21 ± 0.20 a | 7.44 ± 0.23 a | 9.73 ± 0.08 a |
| **Control Sealed** | 2.53 ± 0.07a | 3.61 ± 0.18a | 3.95 ± 0.14 b | 5.29 ± 0.15 b | 6.69 ± 0.14 ab | 9.20 ± 0.28 ab |
| **BAP(0.5 mM)** | 2.62 ± 0.18 a | 3.37 ± 0.19 a | 4.03 ± 0.69 b | 5.17 ± 0.29 b | 6.61 ± 0.24 ab | 8.69 ± 0.12 b |
| **BAP(1.0 mM)** | 2.54 ± 0.15 a | 2.93 ± 0.08 a | 3.79 ± 0.25 b | 5.11 ± 0.07 b | 5.87 ± 0.22 bc | 8.51 ± 0.22 bc |
| **BAP(1.5 mM)** | 2.58 ± 0.11 a | 3.16 ± 0.03 a | 3.55 ± 0.22 b | 4.59 ± 0.15 b | 5.30 ± 0.14 c | 7.63 ± 0.16 c |

**Table 1.  Effect of 6-benzylaminopurine on malondialdehyde content (nmol/g FW) of *jamun* fruit during storage at low temperature. Vertical bars represent standard error of means (n= 3). Bars with different letters on each sampling day indicate significant difference (p ≤ 0.05) among treatments.**

**Total anthocyanins content in fruit peel and pulp**

The results of this study showed that the total anthocyanin content of jamun fruit decreased gradually across all treatments during storage, likely due to enzymatic and non-enzymatic degradation reactions. Treatment with BAP may have stimulated PAL activity, leading to increased anthocyanin synthesis and retention in jamun fruit. Anthocyanin content, responsible for the characteristic deep purple colour and antioxidant properties of Jamun, gradually decreased in both peel and pulp during cold storage (Table 2). However, fruits treated with BAPparticularly at 1.5 mMshowed significantly better retention of anthocyanins throughout the storage period. At 30 days after storage (DAS), the 1.5 mM BAP treatment preserved the highest levels of both peel (336.10 mg/100g FW) and pulp anthocyanins (6.28 mg/100g FW), whereas the untreated fruits stored under open conditions (Control Open) showed a marked decline, retaining only 273.13 mg/100g FW in peel and 4.87 mg/100g FW in pulp. This suggests that BAP effectively slows down anthocyanin degradation, possibly by reducing oxidative stress and delaying senescence, thereby helping maintain the fruit's visual appeal and nutritional quality during extended storage.Several studies have linked increased anthocyanin accumulation with enhanced PAL activity. Postharvest BAP treatment in jamun may have stimulated PAL activity, boosting anthocyanin synthesis and retention. Additionally, BAP likely suppressed PPO activity, further supporting the preservation of anthocyanin pigments (Zhang et al., 2018).

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameters** | T**reatments** | **Days after storage (DAS)** | | | | | |
| **5 DAS** | **10 DAS** | **15 DAS** | **20 DAS** | **25 DAS** | **30 DAS** |
| **Total peel anthocyanin content (mg/100g FW)** | **Control Open** | 489.11 ± 8.77 a | 489.01 ± 8.28 a | 446.46 ± 15.15 a | 403.49 ± 8.81 a | 339.96 ± 7.91 a | 273.13 ± 14.59 b |
| **Control Sealed** | 493.44 ± 16.59 a | 498.92 ± 12.17 a | 450.13 ± 9.05 a | 416.18 ± 5.99 a | 353.31 ± 11.58 a | 289.53 ± 8.78 ab |
| **BAP(0.5 mM)** | 497.78 ± 11.13 a | 502.71 ± 14.58 a | 472.57 ± 12.92 a | 444.52 ± 18.99 a | 349.37 ± 14.48 a | 302.28 ± 9.03 ab |
| **BAP(1.0 mM)** | 488.47 ± 17.69 a | 488.23 ± 9.63 a | 467.82 ± 16.34 a | 443.55 ± 16.24 a | 376.62 ± 4.81 a | 319.08 ± 19.15 ab |
| **BAP(1.5 mM)** | 530.70 ± 11.27 a | 514.07 ± 11.71 a | 490.18 ± 8.07 a | 460.57 ± 7.75 a | 392.09 ± 14.4 a | 336.10 ± 10.91 a |
| **Total pulp anthocyanin content (mg/100g FW)** | **Control Open** | 12.25 ± 0.41 a | 9.99 ± 0.33 a | 8.45 ± 0.29 a | 7.66 ± 0.33 a | 6.56 ± 0.27 a | 4.87 ± 0.08 b |
| **Control Sealed** | 11.91 ± 0.48 a | 10.02 ± 0.24 a | 9.04 ± 0.28 a | 8.07 ± 0.32 a | 6.98 ± 0.06 a | 4.94 ± 0.43 b |
| **BAP(0.5 mM)** | 12.34 ± 0.25 a | 10.39 ± 0.19 a | 9.09 ± 0.25 a | 7.92 ± 0.40 a | 7.00 ± 0.41 a | 5.02 ± 0.09 b |
| **BAP(1.0 mM)** | 12.65 ± 0.30 a | 11.00 ± 0.34 a | 9.58 ± 0.29 a | 8.32 ± 0.32 a | 6.84 ± 0.45 a | 5.71 ± 0.10 ab |
| **BAP(1.5 mM)** | 12.95 ± 0.25 a | 10.97 ± 0.32 a | 9.68 ± 0.29 a | 8.83 ± 0.12 a | 7.75 ± 0.19 a | 6.28 ± 0.28 a |

**Table 2.  Effect of 6-benzylaminopurine on total peel and pulp anthocyanin content (mg/100g FW) of *jamun* fruit during storage at low temperature. Vertical bars represent standard error of means (n= 3). Bars with different letters on each sampling day indicate significant difference (p ≤ 0.05) among treatments.**

**Total soluble solids**

TSS (Total Soluble Solids) is a key quality indicator in the food industry, reflecting the concentration of soluble compounds in fruits and vegetables. It helps assess ripeness, flavor, and processing suitability. In jamun, TSS includes sugars, acids, and vitamins, with major sugars being sucrose, fructose, maltose, glucose, galactose, and mannose (Noomrio&Dahot, 1996).In this experiment, Table 3, presentdata pertaining to the effect of different 6-benzylaminopurine treatments on total soluble solids content in *jamun* fruit. Total soluble solids (TSS), an important indicator of fruit sweetness and ripeness, initially increased in all treatments during early storage, then gradually declined as storage progressed. The untreated control fruits stored in open conditions showed the highest early spike in TSS (17.63% at 15 DAS) but experienced a sharp decline, thereafter, dropping to 12.86% by 30 DAS. In contrast, fruits treated with BAP, especially at 1.5 mM, maintained more stable TSS levels over time. At 30 DAS, BAP (1.5 mM) retained the highest TSS (14.2%), significantly outperforming the open control, which reflects BAP's effectiveness in preserving fruit quality and delaying metabolic deterioration during storage.BAP treatment probably decreased respiration rate in fruit thereby reducing utilization of the respiratory materials, as a consequence delayed loss of soluble solids (Siddiqui et al., 2015).

**Ascorbic acid content**

Ascorbic acid content is an important quality attribute in fruit, offering nutritional benefits and antioxidant properties (Yahia et al., 2001). The ascorbic acid content, a crucial antioxidant and quality indicator in fruits, generally decreased during storage across all treatments. Nevertheless, fruits treated with BAP, particularly those subjected to 1.5 mM BAP, exhibited significantly better retention of ascorbic acid over time (Table 3). At 30 DAS, fruits treated with 1.5 mM BAP maintained the highest ascorbic acid level (29.31 mg/100 g), which was markedly higher than both the open (21.99 mg/100 g) and sealed control fruits (22.42 mg/100 g). This indicates that BAP at 1.5 mM effectively delays vitamin C degradation, thereby enhancing the nutritional and antioxidant profile of Jamun during extended cold storage.BAP treatment in jamun may help slow the breakdown of ascorbic acid by reducing the respiration rate and modulating senescence-related processes (Siddiqui et al., 2015). Similarly, postharvest application of BAP has been shown to preserve ascorbic acid levels in cherimoya fruit (Franco-Mora et al., 2015).

**Total phenolics content**

 In plants, phenolic compounds are synthesized as secondary metabolites. No significant differences in phenolic content were observed between treated and untreated jamun after 5 days of storage. A similar trend was also observed after 20 days of storage (Table.3). At the commencement of storage, phenolic levels were comparable across treatments. However, by 30 days after storage (DAS), the fruits treated with 1.5 mM BAP retained 274.33 mg GAE/100g FW, whereas the control fruits stored in open bags decreased to 225.33 mg GAE/100g FW, and the sealed control fruits to 229.66 mg GAE/100g FW. A higher retention of phenolic compounds was observed in fruit treated with 1.5 mM BAP, with the maximum effect noted at the highest concentration.This suggests that BAP treatment helped in maintaining higher antioxidant levels during cold storage, possibly by reducing oxidative stress and delaying senescenceJamun fruit is rich in phenolic compounds, including flavonoids, anthocyanins, and tannins, which contribute to its antioxidant and nutraceutical properties (Baraiya et al., 2015). Major phenolics include gallic, ferulic, ellagic, and chlorogenic acids, catechin, and caffeic acid. BAP application has been reported to maintain phenol content, possibly by inhibiting polyphenol oxidase activity and reducing phenolic oxidation (Zhang et al., 2018).

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameters** | T**reatments** | **Days after storage (DAS)** | | | | | |
| **5 DAS** | **10 DAS** | **15 DAS** | **20 DAS** | **25 DAS** | **30 DAS** |
| **Total soluble solids** (%) | **Control Open** | 14.5 ± 0.11 ab | 16.26 ± 0.17 a | 17.63 ± 0.17 a | 16.93 ± 0.18 a | 13.93 ± 0.52 a | 12.86 ± 0.18 b |
| **Control Sealed** | 14.36 ± 0.08 ab | 15.6 ± 0.41ab | 16.03 ± 0.23 b | 15.7 ± 0.32 ab | 14.2 ± 0.20 a | 13.33 ± 0.24 ab |
| **BAP(0.5 mM)** | 13.86 ± 0.16 b | 14.8 ± 0.11 b | 15.5 ± 0.17 bc | 14.53 ± 0.38 b | 14.36 ± 0.20 a | 13.46 ± 0.23 ab |
| **BAP(1.0 mM)** | 14.76 ± 0.18 a | 15.23 ± 0.35 ab | 14.93 ± 0.08 c | 14.76 ± 0.36 b | 14.73 ± 0.32 a | 13.53 ± 0.12 ab |
| **BAP(1.5 mM)** | 14.33 ± 0.18 ab | 15.06 ± 0.29 ab | 15.16 ± 0.23 bc | 14.86 ± 0.29 b | 14.6 ± 0.20 a | 14.2 ± 0.23 a |
| **Ascorbic acid content (mg/ 100 g)** | **Control Open** | 58.80 ± 2.15 a | 48.10 ± 1.00 c | 41.50 ± 1.12 a | 34.63 ± 1.21 a | 27.08 ± 1.12 a | 21.99 ± 1.19 b |
| **Control Sealed** | 60.62 ± 0.83 a | 54.63 ± 0.88 a | 44.44 ± 1.05 a | 35.42 ± 2.54 a | 31.20 ± 2.22 a | 22.42 ± 1.50 b |
| **BAP(0.5 mM)** | 60.46 ± 1.78 a | 48.60 ± 0.93 bc | 45.78 ± 1.90 a | 37.16 ± 1.64 a | 33.26 ± 1.71 a | 23.68 ± 0.73 ab |
| **BAP(1.0 mM)** | 63.19 ± 1.04 a | 53.31 ± 1.16 ab | 41.13 ± 1.87 a | 38.29 ± 1.87 a | 34.25 ± 1.31a | 25.40 ± 1.68 ab |
| **BAP(1.5 mM)** | 62.38 ± 2.07 a | 55.42 ± 1.06 a | 44.28 ± 0.89 a | 40.15 ± 1.73 a | 35.96 ± 1.15a | 29.31 ± 1.75 a |
| **Phenolics content (mg GAE/100g FW)** | **Control Open** | 440.66 ± 7.21 a | 370.66 ± 11.05 a | 351.66 ± 6.71 a | 319.66 ± 11.46 a | 278.66 ± 5.48 a | 225.33 ± 4.09 a |
| **Control Sealed** | 460.33 ± 12.91 a | 404.66 ± 22.39 a | 357.66 ± 14.11 a | 326 ± 5.19 a | 270.66 ± 11.21 a | 229.66 ± 11.97 a |
| **BAP(0.5 mM)** | 456 ± 16.52 a | 399.66 ± 9.26 a | 368.33 ± 9.93 a | 343.66 ± 13.44 a | 283.00 ± 11.59a | 255.66 ± 9.33 a |
| **BAP(1.0 mM)** | 468.33 ± 8.76 a | 406.33 ± 10.26 a | 350.66 ± 3.38 a | 330 ± 7.37 a | 301.66 ± 4.25 a | 257.66 ± 18.88 a |
| **BAP(1.5 mM)** | 472.33 ± 8.00 a | 420 ± 6.08 a | 368.33 ± 13.13 a | 345.66 ± 9.83 a | 305 ± 11.71 a | 274.33 ± 7.62 a |

**Table 3.  Effect of 6-benzylaminopurine on total soluble solids (%), ascorbic acid (mg/ 100g) and phenolics content (mg GAE/100g FW)of *jamun* fruit during storage at low temperature. Vertical bars represent standard error of means (n= 3). Bars with different letters on each sampling day indicate significant difference (p ≤ 0.05) among treatments.**

**Total flavonoids content**

Flavonoids, a group of polyphenolic secondary metabolites with strong antioxidant properties, are present in Jamun fruit in forms such as dihydroquercetindiglucoside, myricetin glucoside, rhamnoside, acetyl-rhamnoside, and pentoside (Jagetia, 2017). In this study, the total flavonoid content in Jamun fruits exhibited a gradual decline during cold storage across all treatments. However, the rate of decline was significantly slower in BAP-treated fruits, particularly at the 1.5 mM concentration (Table 4). Fruits treated with 1.5 mM BAP consistently retained higher flavonoid levels at 5-day intervals compared to controls. Notably, flavonoid content in BAP-treated fruits remained relatively stable up to 20 days of storage, while the control fruits, especially those stored in open packets, showed a more rapid decline. By 30 days after storage (DAS), the highest flavonoid content was recorded in the 1.5 mM BAP-treated fruits (33.33 mg RE/g), in contrast to 25.2 mg RE/g in the open control and 26.73 mg RE/g in the sealed control. These findings suggest that BAP treatment, particularly at higher concentrations, effectively preserved flavonoid compounds by mitigating oxidative degradation and delaying senescence, thereby enhancing the nutritional and functional quality of Jamun during extended cold storage.

**Total antioxidant capacity (CUPRAC)**

Jamun fruit's total antioxidant capacity is attributed to various bioactive compounds, including anthocyanins, phenols, flavonoids, and ascorbic acid. During the initial 10 days of storage, the total antioxidant capacity of both treated and untreated fruits remained relatively stable. However, by the 15th day of storage, fruits treated with BAP at concentrations of 0.5 mM, 1.0 mM, and 1.5 mM demonstrated significantly higher antioxidant capacity compared to the control fruits (Table 4). Among the treatments, BAP at 1.0 mM and 1.5 mM consistently maintained elevated antioxidant levels throughout the storage period. By the final day of storage (30 DAS), fruits treated with 1.5 mM BAP exhibited the highest total antioxidant capacity (6.34 mg CE/100g FW), whereas the lowest value was recorded in fruit stored in open LDPE packets without any treatment (4.86 mg CE/100g FW). Notably, there was no significant difference in antioxidant capacity between fruits treated with 0.5 mM BAP and the sealed control. These findings indicate that higher concentrations of BAP are more effective in preserving antioxidant potential by mitigating oxidative stress and delaying senescence during cold storage.BAP treatment has been shown to reduce ROS accumulation, contributing to enhanced antioxidant capacity. Similarly, Zhang et al. (2018) reported that postharvest BAP application preserves antioxidant capacity in litchi fruit.

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameters** | T**reatments** | **Days after storage (DAS)** | | | | | |
| **5 DAS** | **10 DAS** | **15 DAS** | **20 DAS** | **25 DAS** | **30 DAS** |
| **Total flavonoid content (mg RE/ g)** | **Control Open** | 62.06 ± 2.49 a | 54.46 ± 1.39 a | 48.26 ± 1.07 b | 42 ± 1.61 a | 33.93 ± 1.83 b | 25.2 ± 1.05 b |
| **Control Sealed** | 62.6 ± 3.29 a | 58.93 ± 2.88 a | 51.2 ± 1.05 ab | 45.33 ± 2.09 a | 36.2 ± 1.03 ab | 26.73 ± 0.46 b |
| **BAP(0.5 mM)** | 60.53 ± 2.35 a | 58.6 ± 1.81 a | 51 ± 0.70 ab | 45.06 ± 1.50 a | 39.06 ± 1.04 ab | 28.86 ± 0.63 ab |
| **BAP(1.0 mM)** | 61.6 ± 2.80 a | 59.86 ± 1.27 a | 53.2 ± 1.22 a | 45.33 ± 0.93 a | 40.26 ± 1.57 a | 29.6 ± 1.55 ab |
| **BAP(1.5 mM)** | 62.33 ± 1.33 a | 59.33 ± 2.18 a | 53.13 ± 1.15 ab | 46.73 ± 1.27 a | 40.93 ± 0.35 a | 33.33 ± 1.39 a |
| **Total antioxidant capacity (mg CE/100g FW)** | **Control Open** | 10.19 ± 0.08 a | 9.60 ± 0.06 a | 8.38 ± 0.26 b | 7.41 ± 0.14 a | 6.30 ± 0.18 c | 4.86 ± 0.11 c |
| **Control Sealed** | 10.17 ± 0.08 a | 9.92 ± 0.12 a | 8.43 ± 0.12 b | 7.69 ± 0.17 a | 6.66 ± 0.12 bc | 5.51 ± 0.19 bc |
| **BAP(0.5 mM)** | 10.49 ± 0.11 a | 9.77 ± 0.11 a | 9.15 ± 0.09 a | 7.85 ± 0.17 a | 7.18 ± 0.09 ab | 5.72 ± 0.12 ab |
| **BAP(1.0 mM)** | 10.64 ± 0.06 a | 10.07 ± 0.13 a | 9.29 ± 0.08 a | 8.58 ± 0.13 a | 7.42 ± 0.04 a | 6.15 ± 0.13 ab |
| **BAP(1.5 mM)** | 10.49 ± 0.25 a | 10.07 ± 0.11 a | 9.35 ± 0.11 a | 8.88 ± 0.24 a | 7.70 ± 0.09 a | 6.34 ± 0.20 a |

**Table. 4. Effect of 6-benzylaminopurine treatments on total flavonoid content (mg RE/ g) and total antioxidant capacity (µmol TE/g FW) of *jamun* fruit during storage at low temperature. Vertical bars represent standard error of means (n= 3). Bars with different letters on each sampling day indicate significant difference (p ≤ 0.05) among treatments.**

**Conclusion**

This study investigated the effects of 6-Benzylaminopurine (BAP) on the postharvest quality and shelf life of Jamun (*Syzygiumcumini*Skeels) under controlled low-temperature storage conditions. After surface disinfection using 0.1% sodium hypochlorite, fruits were treated with BAP at concentrations of 0.5, 1.0, and 1.5 mM, while distilled water served as the control. Among the tested concentrations, 1.5 mM BAP proved to be the most effective, significantly improving key quality attributes such as weight retention, total soluble solids, and biochemical composition. Overall, BAP at 1.5 mM emerged as the optimal treatment for preserving postharvest quality and extending the shelf life of Jamun fruit during cold storage.



****

**Fig: 1 Appearance of 6-benzylaminopurine treated and untreated *jamun*fruit after 30 days of storage**

**References**

A.O.A.C. (2000). Official methods of analysis. 17th ed. Association of Official Analytical Chemists, Gaithersburg, MD.

An, J., Zhang, M., Lu, Q. and Zhang, Z. (2006). Effect of a prestorage treatment with 6-benzylaminopurine and modified atmosphere packaging storage on the respiration and quality of green asparagus spears. *Journal of Food Engineering,***77**(4), 951–957.

Apak, R., Guclu, K., Ozyurek, M. and Celik, S. E. (2008). Mechanism of antioxidant capacity assays and the CUPRAC (cupric ion reducing antioxidant capacity) assay. *Microchimica Acta*, **160**(4), 413-419.

Arya, S. S., Pegu, K., and Sadawarte, P. D. (2017). Bioactive Compounds and Health Benefits of Jamun (*Syzygiumcumini*). *Bioactive Molecules in Food*, **11**(02), 1-20.

Ayar, K. A., Sanjay Singh, S. S. and Chavda, J. C. (2011). Effect of post harvest treatments on quality of Jamun (*Syzygiumcuminii*Skeels) fruits during storage.

Ayyanar, M. and Subash-Babu, P. (2012). *Syzygiumcumini* (L.) Skeels: A review of its phytochemical constituents and traditional uses. *Asian Pacific Journal of Tropical Biomedicine*, **2**(3), 240-246.

Banerjee, A., Dasgupta, N. and De, B. (2005). In vitro study of antioxidant activity of *Syzygium  cumini* fruit. *Food Chemistry*, **90**(4), 727-733.

Baraiya, N. S., Rao, T. V. R. and Thakkar, V. R. (2015). Improvement of postharvest quality and storability of jamun fruit (*Syzygiumcumini* L. Var. Paras) by zein coating enriched with antioxidants. *Food and Bioprocess Technology*, **8**(11), 2225-2234

Bhargava, U. C., Westfall, B. A. and Siehr, D. J. (1968). Preliminary pharmacology of ellagic acid from *Juglans nigra* (black walnut). *Journal of Pharmaceutical Sciences*, **57**(10), 1728-1732.

Canli, F. A., Sahin, M., Temurtas, N. and Pektas, M. (2014). Improving Fruit Quality of Apricot by Means of Preharvest Benzyladenine and Benzyladenine Plus Gibberellin Applications. *Horttechnology*, **24**(4), 424-427.

Chen, B. and Yang, H. (2013). 6‐Benzylaminopurine alleviates chilling injury of postharvest cucumber fruit through modulating antioxidant system and energy status. *Journal of the  Science of Food and Agriculture*, **93**(8), 1915-1921.

Dagadkhair, A. C., Pakhare, K. N., Todmal, A. D. and Andhale, R. R. (2017). Jamun (*Syzygiumcumini*) Skeels: A traditional therapeutic tree and its processed food products. *International Journal of Pure and Applied,* **5**(5), 1202-1209.

de Carvalho, T. I. M., Nogueira, T. Y. K., Mauro, M. A., Gómez-Alonso, S., Gomes, E., Da- Silva, R. and Lago-Vanzela, E. S. (2017). Dehydration of jambolan (*Syzygiumcumini* (L.))  juice during foam mat drying: Quantitative and qualitative changes of the phenolic compounds. *Food Research International*, **102**, 32-42.

Efendi, D. and Hermawati, H. (2010). The use of bee wax, chitosan and BAP to prolong shelflife of Mangosteen (*Garcinia mangostana*L.) fruit. *JurnalHortikultura Indonesia*, **1**(1), 32-39.

Flores, F. B., Sánchez-Bel, P., Valdenegro, M., Romojaro, F., Martínez-Madrid, M. C. and Egea, M. I. (2008). Effects of a pretreatment with nitric oxide on peach (*Prunus persica* L.) storage at room temperature. *European Food Research and Technology*, **227**(6), 1599.

Franco-Mora, O., Morales-Pérez, A. A., Castañeda-Vildózola, Á., Morales-Rosales, E. J. and Sánchez-Pale, J. R. (2015). Sprays mixing resveratrol and benzylaminopurine previous harvest helps to preserve postharvest quality in cherimoya. *Journal of Agriculture and Life Science*, **2**(2), 16-24.

Jagetia, G. C. (2017). Phytochemical Composition and pleotropic pharmacological properties of jamun, *Syzygiumcumini*skeels. *Journal of Exploratory Research in Pharmacology*,**2**(2), 54-66.

Kachroo, A. and Robin, G. P. (2013). Systemic signaling during plant defense. *Current Opinion in Plant Biology*, **16**(4), 527-533.

Koley, T. K., Barman, K. and Asrey, R. (2011). Nutraceutical properties of Jamun *Syzygium  cumini* (L.) and its processed products. *Indian Food Industry*, **30**(3), 34-37.

Lees, D. H. and Francis, F. J. (1972). Standardization of pigment analyses in cranberries. *HortScience*, **7**, 83-84.

Maftoonazad, N., Ramaswamy, H. S. and Marcotte, M. (2008). Shelf‐life extension of peaches through sodium alginate and methyl cellulose edible coatings. *International Journal of Food Science and Ttechnology***, 43**(6), 951-957.

Massolo, J. F., Lemoine, M. L., Chaves, A. R., Concellón, A. and Vicente, A. R. (2014). Benzyl-aminopurine (BAP) treatments delay cell wall degradation and softening, improving quality maintenance of refrigerated summer squash. *Postharvest Biology and Technology*, **93**, 122-129.

Noomrio, M. H. and Dahot, M. U. (1996). Nutritive value of *Eugenia jambosa* fruit. *Journal of Islamic Academy of Sciences*, **9**(1), 9-12.

Palma, J. M., Freschi, L., Rodríguez-Ruiz, M., González-Gordo, S. and Corpas, F. J. (2019). Nitric oxide in the physiology and quality of fleshy fruits. *Journal of Experimental Botany*, **70**(17), 4405-4417.

Patil, S. S., Thorat, R. M. and Rajasekaran, P. (2012). Utilization of jamun fruit (*Syzygiumcumini*) for production of red wine. *Journal of Advanced Laboratory Research in Biology*, **3**(3), 200-203.

Rai, D. R., Chadha, S., Kaur, M. P., Jaiswal, P. and Patil, R. T. (2011). Biochemical, microbiological and physiological changes in Jamun (*Syzyiumcumini* L.) kept for long term storage under modified atmosphere packaging. *Journal of Food Science and Technology*, **48**(3), 357-365.

Raza, A., Saif-ul-Malook, N. S., Qasrani, S. A., Sharif, M. N., Akram, M. N.and Ali, M. U. (2015). Extraction of Bioactive Components from the Fruit and Seed of Jamun (*Syzygiumcumini*) Through Conventional Solvent Extraction Method. *American-Eurasian Journal of Agricultural and Environmental Sciences*, **15**(6), 991-996.

Saurabh, V., Barman, K. and Singh, A. K. (2019). Synergistic effect of salicylic acid and chitosan on postharvest life and quality attributes of jamun (SyzygiumcuminiSkeels) fruit. *Acta Physiologiae Plantarum*, **41**(6), 89.

Siddiqui, M. W., Singh, J. P., Nayyer, M. A., Barman, K., Ahmad, M. S. and Kumar, V. (2015). 6-Benzylaminopurine affects lipid peroxidation and membrane permeability and thereby preserves curd quality and antioxidants during storage of cauliflower. *Acta Physiologiae Plantarum*, **37**(5), 96.

Singleton, V. L., Orthofer, R. and Lamuela-Raventos, R. M. (1999). Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. *Methods in Enzymology*, **299**, 152-178.

Wingler, A., von Schaewen, A., Leegood, R. C., Lea, P. J. and Quick, W. P. (1998). Regulation of leaf senescence by cytokinin, sugars, and light: effects on NADH-dependent hydroxypyruvate reductase. *Plant Physiology*, **116**(1), 329-335.

Yahia, E. M., Contreras-Padilla, M. and Gonzalez-Aguilar, G. (2001). Ascorbic acid content in relation to ascorbic acid oxidase activity and polyamine content in tomato and bell pepper fruits during development, maturation and senescence. *Food Science and Technology*, **34**(7), 452-457.

Zhang, D., Xu, X., Zhang, Z., Jiang, G., Feng, L., Duan, X. and Jiang, Y. (2018). 6-Benzylaminopurine improves the quality of harvested litchi fruit. *Postharvest Biology and Technology*, **143**, 137-142.

Zhishen, J., Mengcheng, T. and Jianming, W. (1999). The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food Chemistry*, **64**(4), 555-559.