**Isolation and Control of Post-Harvest Apple Rot Pathogens Using Ginger (*Zingiber officinale* Roscoe.) Extracts in Mubi, Nigeria**

**ABSTRACT**

A survey for the rot of apple fruits sold in two markets of Mubi North Local Government, Adamawa State (Mubi Main Markets and Mubi New Markets) was conducted between June and September 2023. Also, the *in vitro* effect of ginger (*Zingiber officinale*) aqueous and ethanolic root extracts on rot control was carried out. Five fungal pathogens viz; *Aspergillus niger*, *Ulocladium botrytis*, *Rhizopus stolonifer*, *Rhizopus microsporus*, and *Mucor hiemalis* were identified and proven through pathogenicity test to be pathogenic. Out of the total sample size of 32 fruits (16 per market), *R.stolonifer* had the highest percentage incidence in both markets with 37.50 % in the main market and 31.25 % in the new market while the least was *M. hiemalis* (6.25 %) in the main market and U. botrytis (6.25 %) Newmarket. Virulence of pathogens showed that *R. stolonifer* had very high, *A. niger* and *R. microsporus* had high while *U. botrytis* and *M. hiemalis* had moderate. Control trails on the pathogenic organisms (invitro) show that both extracts were able to inhibit the growth of pathogen with the ethanolic extract having high inhibition diameter of 0.70 cm in *A. niger*, 0.93 cm in *U. botrytis* and 0.49 cm in *R. stolonifer*. In comparison, aqueous had a higher mean zone of inhibition in *M. hiemalis* (0.63 cm) and *R. microsporus* (0.33 cm). There was a statistically significant difference between all the extract concentrations and the control at P≤ 0.005 It was recommended that the active components of both extracts be identified and isolated so that they can be incorporated to form fungicides that can be used commercially to control pre- and post-harvest fruit rot.

**Keywords**: Pathogens, Pathogenicity, *Zingiber officinale,* Apple, and root extracts

**Introduction**

“The apple tree (*Malus domestica* Borkh.) is a deciduous tree in the Rose family best known for its sweet, Pomaceous fruit, the apple. It is cultivated worldwide as a fruit tree and is the most widely grown species in the genus Malus. The tree originated in central Asia where its wild ancestor, *Malus sieversii* is still found today. It is an important part of the human diet” (Khanizadehet al*.,* 2008). “The main apple cultivars can be roughly divided into red varieties such as ‘Red Delicious’, ‘Fuji’, and ‘Royal Gala’ and non-colored varieties such as ‘Golden Delicious’, ‘Granny Smith’, and ‘Orin’” (Chen et al. 2021). “Research suggests that apples may reduce the risk of colon cancer, prostate cancer, and lung cancer” (NRCR, 2008). “They may also help with heart disease, weight loss, and controlling cholesterol, as they have fiber, which reduces cholesterol by preventing re-absorption and are bulky for their caloric content like most fruits and vegetables” (Sharma, 2005; AKYFH, (2008).

Approximately 20-25% of the harvested fruits are deteriorated by pathogens during postharvest handling even in advanced countries (Droby, 2006). “Postharvest losses are often harsher in developing countries due to a lack of storage and transportation facilities. Fruit infections by fungi may appear during the growth period, harvesting, handling, transportation, post-harvest stockpile, and marketing conditions, or after procuring by the consumer. Fruits incorporate high levels of nutrients, elements, and sugars and their low pH values make them exceptionally desirable to fungal decay” (Sharma et al., 2024). “Based on the cultivar, season, and production area, fungi are considered an essential post-harvest loss agent of different fruits” (Ewekeye et al., 2016). “Infections caused during post-harvest conditions lower the shelf life and adversely affect the market value of fruits” (Tripathi et al., 2008). “Moreover, mycotoxins the secondary metabolites produced by moulds have adverse effects on humans and animals” (Zain, 2011). “The contamination of fruits with mycotoxins has not only caused health hazards but also resulted in economic losses, especially for exporting countries. Two mechanisms of infection can be determined, primary and secondary infection. The primary infection usually happens in the field (orchards) or during sorting before storage after harvesting. The secondary infection happens when fungal mycelium and spores spread from infected apples to uninfected apples” (Dutot et al., 2013).

“Apple rot is an economically significant disease on apples (*Malus* *domestica* Borkh) and can be caused by several filamentous fungi with a worldwide distribution like *Penicillium expansum* Link. and *Botrytis cinerea* Pers. which are common and causing significanteconomic losses in the USA and Europe” (Sutton et al., 2014). “Apple rot incidence may vary depending on the cultivar” (Sever et al., 2012; Weber, 2011) and harvest time (Borve et al., 2013). Sutton et al. (2014) reported that *Alternaria alternata* causes core rot (Niem *et al.*, 2007) and rot around injuries or at calyx. “Grey mould caused by *Botrytis cinerea*, *Fusarium* rot caused by several *Fusarium* species, and brown rot caused by *Monilinia* *fructigena* (Aderh and Ruhland) Honey and blue mould decay caused by *P. expansum* were found more rarely” (Borve et al., 2013). “Among the different fungal pathogens, *Alternaria alternata, Botrytis cinerea, Glomerella cingulata, Monilinia fructigena* and *Penicillium expansum* are the dominant ones causing post-harvest losses in apple” (Kumari et al., 2019).

Ginger (*Zingiber officinale*) belongs *to Zingiberaceae* family (Sharma, 2010). Ginger is a perennial, herbaceous, monocotyledonous plant, cultivated in tropical and sub-tropical areas worldwide. Rhizome of ginger has a pungent taste and smell, it is mostly used as a food additive (Jangam and Thorat, 2010). According to Godowoli and Lawan (2024), ginger has direct anti-microbial activity and thus can be used in the treatment of bacterial infections. The chemical constituents of ginger extract and essential oil are the poly-phenolic ketones called gingerols or Oleoresin (Park et al., 2008). The antifungal effect of ginger is due to gingerone, dihydrogingerone, and dehydroshogaol (Ficker et al., 2003). Some other alkaloids, terpenes, and terpenoid derivatives have antimicrobial activities on different bacterial pathogens (Huang et al., 2022). “Ginger has been used as a spice and medicine for over 200 years in Traditional Chinese Medicine” (Shahrajabian et al., 2019). “The obtained findings suggest the potential of ginger extract as an additive in the food and pharmaceutical industries” (Shahrajabian et al.,2019; Peng et al., 2022). “Results indicate that ginger contains monoterpenoids, sesquiterpenoids, phenolic compounds, and its derivatives, aldehydes, ketones, alcohols, and esters, which provide a broad antimicrobial spectrum against different microorganisms and make it an interesting alternative to synthetic antimicrobials” (Lim and Wong, 2018; Gao et al., 2022). “For example, 6-gingerol inhibits the proliferation of human cervical cancer cells and induces their apoptosis” (Moorkoth et al., 2021). “Recent studies have shown that both ginger extract and ginger essential oil have antifungal activities against plant pathogens, such as *Fusarium oxysporum* and *Colletotrichum falcatum”* (Abdullahi et al., 2020; Bordoh et al., 2020). Noshirvani et al*.* (2017) reported that “ginger essential oils could be used to plasticize chitosan-carboxymethyl cellulose films while improving moisture permeability and maintaining antifungal activity. These findings indicate that ginger extract might be useful as an alternative to fungicides and bactericides” (Noshirvani et al., 2017). Yusuf et al*.* (2019) reported “ginger contains phytochemicals which include flavonoids, saponins, steroids, glycosides, resins, and tannins to be the major phytochemicals present in the plant. Ginger is also a rich source of mineral elements and vitamins which include zinc, iron, phosphorus, calcium, magnesium, potassium, vitamin E, and Vitamin C” (Yusuf *et al.*, 2019).

**MATERIALS AND METHODS**

**Study Area**

The study was conducted in Mubi town which comprised both Mubi-North and Mubi-South Local Government Areas (L.G. As) of Adamawa State, Nigeria. The town is located in the North Eastern region of Nigeria between latitudes 100141N and 100181N of the equator and longitudes 130141E and 130191E (Tizhe et al. 2019). It occupies a land of about 725.85 Km2. The area has a tropical climate with an average temperature of 320C and lies within the Sudan savannah vegetation zone of Nigeria. The area has an average relative humidity ranging from 28 % - 45 % and an annual rainfall of about 1056 mm (Adebayo, 2004).

# **3.2 Sources of Samples and Sample Size**

A total of 16 samples were collected from the two different Markets at four different locations using a stratified sampling technique (8 samples from each market) in a sterile polythene bag. The Markets were; Mubi main market (M MM) and Mubi new market (MNM).

# **3.3 Medium of Isolation**

The medium used for isolation was Potato Dextrose Agar (PDA), which was prepared according to standard microbiological procedures (Saira et al. 2022). The prepared media was autoclaved for 15 minutes, 10Ib pressure and allowed to cool. The Petri dishes used for solid media were sterilized in an oven at 160oC for 6 hours using a sterilization can. The needle and cork borer used for inoculation were sterilized by flaming and then cooled by dipping into a methylated spirit. The inoculation of the organisms was conducted in an inoculating chamber. The bench top was sterilized with 95% ethanol and then UV light was used to sterilize the inoculating chamber for 30 minutes.

**Incidence and Virulence of Apple Fruit Rot Pathogens in the Markets**

Apple samples were collected at random from the two markets selected, from different traders at different locations in the market. The incidence of apple fruit rot in the Market was determined by counting the infected apples from the samples collected from each market.

Virulence of fruit rot was assessed and scored according to the visual scale of 1-5 in which;

1 1- 20 % of apple fruit infected,

1. 21-40 % of apple fruit infected,
2. 41-60 % of apple fruit infected,

4 61-80 % of apple fruit infected,

5 More than 80 % of apple fruit infected.

The reactions of the isolates based on the 1-5 visual scale were grouped into the following categories based on the modified Ratanacherdchail et al. (2010) pathogenic potential of rating isolates

1-20 % - Low Virulent group

21-40 % - Moderately Virulent group

41-60 % - High Virulent group

61-80 % - Very High Virulent group

Above 80 % - Totally Virulent group

**Isolation and Identification of Pathogens**

About 5 mm square pieces of apple fruit showing rot lesions were cut with a sterilized blade and sterilized in 0.1 % mercuric chloride for 30 seconds. They were then rinsed in three changes of sterile distilled water and blotted dry between sterile Whatman No. 1 filter papers and were plated out on PDA. The plates were incubated at 25 0 C for 24 hrs and observed for any growth. The resulting spore colonies were transferred to fresh Potato Dextrose Agar (PDA) plates and in a refrigerator for further studies. The slides of the organism isolated were prepared and stained with lactophenol cotton blue observed under the microscope and subsequently identified by comparing the morphological characteristics of the organisms under the microscope with already identified structures (Dabban et al. 2024).

**Pathogenicity Test**

The healthy semi-ripe apple fruits (uniform in size) were selected and washed, surface sterilized in 0.5 % Sodium hypochlorite solution (bleach) for 3 minutes and rinsed in 3 changes of sterile distilled water and then air dried according to the method of Zakawa et al. (2019). The surface sterilized apple fruits were wounded with cork borer of 5mm diameter and bored tissue was removed (Ekwekeye et al., 2023). The wounded apple fruits were inoculated with a 4mm disc of the inocula and sealed with a sterile vesper prepared from Vaseline. The control experiment was set up in the same way except that sterile distilled water was used instead of inocula. All the wounded apple fruits were wrapped in black polythene bags (Manici and Cerato, 1994). Inoculated fruits were incubated in desiccators that had been sterilized. Regular observations were made for isolates and comparison with original isolates.

**Preparation of Plant Root Extracts of** ***Zingiber officinale***

Stem of *Zingiber officinale* was collected from Mubi main market, Mubi North Local Government of Adamawa State. The fresh stem was collected, and washed with tap water and with sterile distilled water, this were dried under the shade and were pulverized with mortar and pestle into a powdery form. Seventy grams (70 g) of the root powder was soaked in 1000 ml of cool distilled water and ethanol for 24 hours. The suspensions were then filtered using sterile cheese cloths. The extracts were then poured into a clean sterile conical flask plugged with cotton wool and heated between 30oC to 60oC in a water bath. This was then allowed to cool, wrapped in aluminium foil, and kept until when required.

**Determination of in vitro Efficacy of Root Extracts of Ginger on the Pathogens**

***In vitro test***

The aqueous and ethanolic stem extract used for the antifungal activity test were prepared into three (3) different concentrations ranging from 500 mg/ml to 1500 mg/ml (i.e. 500, 1000, and 1500 mg/ml). The extract concentration was prepared by weighing 1.5 g of the extract into 1 ml of sterile distilled water (1500 mg/ml). Serial dilution of the extract was carried out into two (2) different labeled bottles to obtain concentrations of 1000 mg/ml and 500 mg/ml respectively (Zakawa et al., 2021). The control contains sterile distilled water in place of the root extracts, three (3) replicates were used for each pathogen with the extract concentration and control. The disc was used to cut from 7-day-old culture of fungi and was incubated at room temperature of 250C for 7 days in which zones of inhibition were measured starting from two days after inoculation.

**Qualitative Phytochemical Analysis**

The extracts were analyzed for alkaloids, flavonoids, tannins, saponins, phenols, and glycosides. The method described by Silva et al. (2017), Gul et al. (2017), Kumar et al. (2018). And Uma et al. (2018) were adopted.

**Experimental Design and Data Analysis**

The experimental layout was a Completely Randomised Design (CRD) of the two extracts, each extract at the same concentration (i.e. 500 mg/ml, 1000 mg/ml, and 1500 mg/ml) and the experiment will be replicated three (3) times. All data were analyzed using Analysis of Variance (ANOVA) to test for significance and means were separated using the Ducan Multiple Range Test (DMRT). The statistical package used was the SPSS computer software program.

# **RESULTS**

# **Incidence and Virulence of Apple Fruit Rot Pathogens in the Markets**

The incidence of each fungal pathogen from the two markets visited was recorded with Mubi main markethaving *Rhizopus stolonifer* with the highest incidence at 37.50% followed by *Aspergillu niger* at 25%, *Rhizopus microsporus* at 18.75%, *Ulocladium botrytis* at 12.50% while *Mucor hiemalis* had the lowest incidence at 6.25% respectively. Mubi new main had *R. stolonifer* with the highest incidence (31.25%), *R. microsporus* (25%), *M. hiemalis,* and *A. niger* (18.75) while *U. botrytis* had the lowest incidence with 6.25% as shown in Figure 1.

The virulence of pathogens showed that all pathogens were virulent at different degrees, *R. stolonifer* virulence was scored as very high as fruit rot covered up to 70% of the fruit within a few days, *A. niger* and *R. microsporus* had a high virulence while *M. hiemalis* and *U. botrytis* were scored as moderate virulence (Table 1).

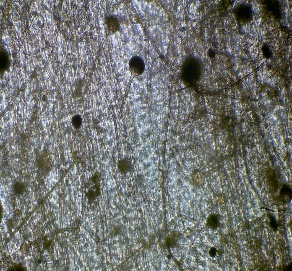
Figure 1: Percentage Incidence of Rot Caused by Pathogens in the Markets

# **Table 1: Virulence of Apple Fruit Rot Pathogens**

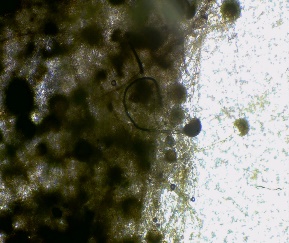
|  |  |  |
| --- | --- | --- |
| Organisms | Virulence |  |
| *A. niger* | High |  |
| *U. botrytis* | Moderate |  |
| *R. stolonifer* | Very High |  |
| *M. hiemalis* | Moderate |  |
| *R. microspores* | High |  |
|  |  |  |

# **Isolation and Identification of Pathogens**

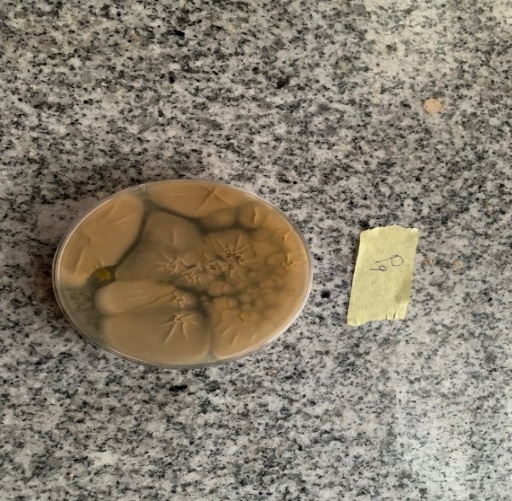
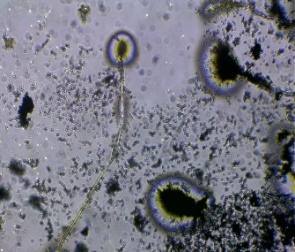
Five fungal pathogens (*Apergillus niger*, *Ulocladium botrytis*, *Rhizopus stolonifera*, *Mucor hiemalis,* and *Rhizopus microsporus*) were successfully isolated from the infected apple fruits. They were confirmed through a pathogenicity test to be responsible for apple fruit rot disease in the study area (Figure 2 (a-k)).



a b c d



e f g h

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i j k

Figure 2: (a) Micrograph of *A.niger* X 40 (b) Seven-Day Old Pure Culture of *A. niger,* (c) Micrograph of *M. hiemalis* X 40 (d) Seven Day Old Pure Culture of *M. hiemalis* (e) Micrograph of *R. microspores* X 40 (f) Seven Day Old Pure Culture of *R. microspores* (g) Micrograph of *R. stolonifer* X 40 (h) Seven-Day Old Pure Culture of *R. stolonifer* (i) Micrograph of *U. botrytis* X 40 (j/k) Seven Day Old Pure Culture of *U. botrytis*

**Pathogenicity Test**

The pathogenicity test showed that all five fungal pathogens were pathogenic to apple fruit at different levels (Figure 3). Rot was not observed on the first day of incubation in all organisms until the second day. There was a statistically significant difference between the control and all fungal pathogens starting from day two. *R. stolonifer* had the highest rot diameter, followed by *M. hiemalis* while *A. niger* had the lowest rot diameter respectively (Table 2)

# **Table 2: Pathogenicity Test of the Isolated Apple Fruit Rot Pathogens**

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| Organisms | Days/Inhibition Zones (cm) | | | | |  |
| 2 | 3 | 4 | 5 | 6 | 7 |
| Control | 0.00a | 0.00a | 0.00a | 0.00a | 0.00a | 0.00a |
| *Aspergillus niger* | 0.17a | 0.74b | 1.10b | 3.90b | 4.83b | 6.70b |
| *Ulocladium botrytis* | 0.25ab | 0.73b | 2.40c | 4.65c | 5.85c | 7.93c |
| *Rhizopus microsporus* | 0.27ab | 0.80b | 2.38c | 4.68c | 6.18c | 8.20c |
| *Mucor hiemalis* | 0.32ab | 1.40c | 4.28e | 8.25e | 9.48d | 10.37d |
| *Rhizopus stolonifer* | 0.63b | 2.50d | 3.87d | 7.72d | 10.32e | 10.50d |

Means in the same column followed by the same superscript are not significantly different at P ≤ 0.05 D.M.R. T



Figure 3: (a) Selected Samples of Apple Fruits from different Markets in Mubi North (b) Sample of semi-rapped Apple fruits for Pathogenicity test day one (c) Sample of Apple fruits after seven days during pathogenicity test showing rot.

# **Determination of *in vitro* Efficacy of Root Extracts of *Zingiber officinale* on the Pathogens**

Results from the *in vitro* efficacy determination of both aqueous and ethanolic extracts of *Z. officinale* on the apple fungal pathogens are presented in Table 3. There was a statistically significant difference between all extract concentrations and the non-treated control at p≤0.05 for all five fungal pathogens. The interaction between the two extracts of solvent used also showed a statistically significant difference at p≤0.05 for only three fungal pathogens (*A. niger, R. stolonifer,* and *R. microsporus*) (Table 3)

# **Table 3: Effect of Extract Concentrations on Zones of Inhibition (cm) of Apple Fruit Rot Pathogens**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  |  |  | Organisms |  |  |
| Treatment | *A. niger* | *U. botrytis* | *R. stolonifer* | *M.hiemalis* | *R. microsporus* |
| **Conc. (mg/ml) – (C)** |  |  |  |  |  |
| 0 | 0.00a | 0.00a | 0.00a | 0.00a | 0.00a |
| 500 | 0.42b | 0.80b | 0.35b | 0.50b | 0.17b |
| 1000 | 1.03c | 1.07b | 0.32b | 0.70bc | 0.29c |
| 1500 | 0.95c | 1.17b | 0.32b | 1.07c | 0.67d |
| SE ± | 0.09 | 0.15 | 0.08 | 0.15 | 0.04 |
| **Extract – (E)** |  |  |  |  |  |
| Aqueous | 0.50a | 0.59a | 0.00a | 0.63a | 0.33b |
| Ethanolic | 0.70b | 0.93a | 0.49b | 0.50a | 0.23a |
| SE± | 0.06 | 0.10 | 0.06 | 0.11 | 0.03 |
| **Interaction**  C x E | \* | NS | \* | NS | \* |

Means in the same column followed by the same superscript are not significantly different at p ≤ 0.05 D.M.R.T

Keys:

* = Significant at p ≤ 0.05

NS = Not significant at p ≤ 0.05

**Effect of aqueous extracts of *Zingiber officinale* on the fungal rot pathogens of apple**

The result for the effect of aqueous extracts concentrations of *Zingiber officinale* on the growth of the fungal pathogens are presented in Table 4 There was a statistically significant difference between the non-treated control and the other treatments at p≤0.05. For *A. niger*, treatment with 1000 mg/ml had the highest zone of inhibition with 1.20 cm while 500 mg/ml had the lowest with 0.10 cm. For *U. botrytis*, treatment with 1500 mg/ml had the highest zone of inhibition with 1.03 cm while 500 mg/ml had the lowest with 0.47 cm. *M. hiemalis* had treatment with 1500 mg/ml with the highest zone of inhibition at 1.13 cm although there was no statistically significant difference when compared with 1000 mg/ml while 500 mg/ml had the lowest zone of inhibition at 0.40 cm. For *R. microspores*, treatment with 1500 mg/ml had the highest zone of inhibition with 0.70 cm while treatment with 500 mg/ml had the lowest with 0.23 cm. There was, however, no statistically significant difference between extract concentrations for *R. stolonifera* at p≤0.05.

**Table 4: Effect of Aqueous Extract Concentrations of *Zingiber officinale* on Zones of Inhibition (cm) of Apple Rot Pathogens**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  |  |  | Organisms |  |  |
| Treatment | *A. niger* | *U. botrytis* | *R. stolonifer* | *M.hiemalis* | *R. microsporus* |
| Conc. (mg/ml) |  |  |  |  |  |
| 0 | 0.00a | 0.00a | 0.00a | 0.00a | 0.00a |
| 500 | 0.10b | 0.47b | 0.00a | 0.40b | 0.23b |
| 1000 | 1.20d | 0.87c | 0.00a | 1.00c | 0.38b |
| 1500 | 0.70c | 1.03d | 0.00a | 1.13c | 0.70c |
| SE ± | 0.13 | 0.21 | 0.12 | 0.21 | 0.05 |

Means in the same column followed by the same superscript are not significantly different at p ≤ 0.05

**Effect of ethanolic extracts of Zingiber officinale on the fungal rot pathogens of apple in vitro**

The results for the effect of ethanolic extract concentrations of *Zingiber officinale* on the growth of the fungal pathogens are presented in Table 5. There was a statistically significant difference between the non-treated control and the other treatments at p≤0.05. For *A. niger*, treatment with 1500 mg/ml had the highest zone of inhibition with 1.20 cm while 500 mg/ml had the lowest with 0.73 cm. For *U. botrytis*, treatment with 1500 mg/ml had the highest zone of inhibition with 1.30 cm while 500 mg/ml had the lowest with 1.13 cm. *M. hiemalis* had treatment with 1500 mg/ml with the highest zone of inhibition at 1.00 cm while 1000 mg/ml had the lowest zone of inhibition at 0.40 cm. For *R. microspores*, treatment with 1500 mg/ml had the highest zone of inhibition with 0.63 cm while treatment with 500 mg/ml had the lowest with 0.10 cm. There was, however, no statistically significant difference between extract concentrations for *R. stolonifer* at p≤0.05

# **Table 5: Effect of Ethanolic Extract Concentrations of *Zingiber officinale* on Zones of Inhibition (cm) of Apple Rot Pathogens**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  |  |  | Organisms |  |  |
| Treatment | *A. niger* | *U. botrytis* | *R. solonifer* | *M.hiemalis* | *R. microsporus* |
| Conc. (mg/ml) |  |  |  |  |  |
| 0 | 0.00a | 0.00a | 0.00a | 0.00a | 0.00a |
| 500 | 0.73b | 1.13b | 0.70b | 0.60b | 0.10b |
| 1000 | 0.87b | 1.27bc | 0.63b | 0.40b | 0.20b |
| 1500 | 1.20c | 1.30c | 0.63b | 1.00c | 0.63c |
| SE ± | 0.13 | 0.21 | 0.12 | 0.21 | 0.05 |

Means in the same column followed by the same superscript are not significantly different at p ≤ 0.05

**Qualitative Phytochemical Analysis**

The result for the phytochemical analysis of aqueous and ethanolic extracts of *Zingiber officinale* is presented in Table 6 Ethanolic extract of *Z. officinale* had the excess presence of alkaloid, flavonoid, and phenols while they are moderately present in the aqueous extract. Saponins however occurred in excess for the aqueous extract compared to ethanolic extract which was moderate, tannin occurred in trace amounts in both extracts while glycoside did not occur in both extracts.

# **Table 6: Qualitative Phytochemical Composition of *Zingiber officinale* Extracts**

|  |  |  |
| --- | --- | --- |
| Phytochemicals | Ethanolic Extracts | Aqueous Extracts |
| Alkaloid | +++ | ++ |
| Flavonoid | +++ | ++ |
| Tannins | + | + |
| Saponins | + | ++ |
| Glycosides | - | - |
| Phenols | ++ | + |

Keys:

-absent

+ trace

++ moderate

+++ excess

**Discussion**

**The study showed that various fungi are associated with post-harvest rot diseases of Apple in the study area (Mubi North Local Government, Adamawa State). This survey from Mubi in 2023 shows that Apple fruit rot occurred in all locations (Markets). However, *R. stolonifer* had the highest percentage incidence in the Main Market with 37.50 %, *A. niger* at 25 %, *R. microsporus* at 18.75 %, *U. botrytis* at 12.50 %, and *M. hiemalis* at 6.25 %. In the New Markets, *R. stolonifer* had 31.25%, *R. microsporus* had 25.00 %, *A. niger* and *M. hiemalis* had 18.75 %, while *U. botrytis* had 6.25 %. The incidence of these pathogenic organisms in the study area could be partly because of our handling during transportation as well as storage by the sellers in the market. This confirms the report of** Anand and Barau (2022) that harvesting, sorting, grading, inadequate storage, handling, and transportation are the major operational causes of post-harvest loss and waste of fruits and vegetables in the agri-fresh produce supply chains**. The organisms' Virulence indicates that *R. stolonifer* had very high virulence, *A. niger,* and *R. microsporus* had high virulence, and *U. botrytis* and *M. hiemalis* had moderate virulence. This agrees with Zakawa et al. (2018) who reported that *Aspergillus* spp (*A. niger* and *A. flavus*) have a high virulence in mango fruit with rot covering 41-60 % of the fruits.**

**The fungal pathogen isolated and confirmed through the pathogenicity test to cause Apple fruit rot disease in Mubi include *A. niger, U. botrytis, R. stolonifer, M. hiemalis,* and *R. microspores.* This finding agrees with that of Muqeet et al. (2020) who reported *A. niger* as one of the major fungal pathogens causing rot in three cultivars of apples (Royal Gala, Golden Delicious, and Kulur) in India. Kumari et al. (2019) reported *Botrytis cinerea* and *A. niger* to be among the most dominant fungal pathogens causing rot in Apple fruits. Abdullah et al. (2016) reported *M. hiemalis* and *A. niger* among the fungal pathogens responsible for Apple fruit rot in Yemen. Magga and Zakawa (2018) reported *Mucor* spp, *R. stolonifer,* and *A. niger* as fungal pathogens responsible for fruit rot of papaya in Mubi.**

**From the results obtained from the anti-fungal study, it has been shown that the inhibition of mycelial growth of the isolates was higher with the ethanoic extracts of ginger compared to Aqueous extracts. This was in agreement with the works of Shahida et al. (2022) who** also observed that the ethanolic extracts of ginger showed higher antibacterial and antifungal activity than the aqueous extract. The ethanol extracts were considered a powerful inhibitor compared to the extracts of methanol and chloroform (Sharaf and Al-Zaidi, 2021)**.** These results were similar to a previous study conducted by Ahmad et al. (2024) showing the active antimicrobial potential of ethanolic extracts of ginger, as compared to the aqueous extracts on fungal pathogens. Results obtained from the *in vitro* antifungal activities of this research showed that ginger ethanolic extracts were more effective in the inhibition of *U*. *botrytis* followed by *A. niger, M. hiemalis, R. microspores* and were lowest on *R*. *stolonifer* while the aqueous extract was more effective on *A. niger*, followed by *M. hiemalis*, *U. botrytis*, *R. microsporus* and wasn’t effective against *R*. *stolonifer*. This is in agreement with Amienyo and Ataga (2007) who reported the antifungal activities of the ethanolic extracts of *Z. officinale* against the mycelial elongation of *Rhizopus stolonifer*, *Botrydiplodia theobromae, A. niger*, *F. solani*, and *F. oxysporum*. However, it is not in agreement with Ishwa et al. (2024) who reported that aqueous ginger extract was able to inhibit the mycelial growth of *R. stolonifera*. This difference may be due to the concentration of extract used, their mode of extraction, or partly because the Z. officinale plant was grown in different ecological regions.

Numerous studies by Giriraju and Yunus (2013); Bordoh et al. (2020) and Makhuvele et al. (2020) have shown that “ginger extract has a wide range of antimicrobial activities and acts as a botanical fungicide by inhibiting spore germination and growth of plant pathogens”. Bordoh et al. (2020) reported that “ginger crude extract at 10.0 mg/mL showed an effective antifungal effect against *Colletotrichum gloeosporioides* in vitro; it suppressed conidial germination and mycelial growth by 88.48% and 87.50%, respectively” (Bordoh et al., 2020). Essential oils obtained from hydro-distillation of ginger or aqueous extracts from ginger seed had previously shown antimicrobial activity against a wide range of spoilage pathogens, including *A. niger*, *Aspergillus flavus* Link, *F. oxysporum*, *Fusarium roseum* Link:Fr. and *R. stolonifer* (Tripathi et al., 2008).

Phytochemical analysis of the two extracts showed the presence of some active components which include; *flavonoids, phenols, tannins, saponins,* and *alkaloids* while glycosides were absent in both aqueous and ethanolic extracts. The botanical bio-pesticides represent an alternative for control with low environmental impact and high food safety. The same results were reported by Kela et al. (2023) in Gombe State, Nigeria. Antimicrobial activity could be attributed to the presence of *gingerol* and shogaol (phenolic compounds) which are active ingredients in ginger (Poplawska et al., 2024). The antimicrobial activity of ginger is reported to depend on the chemical composition, extraction solvent, and method (Park et al., 2008; Beristain-Bauza et al., 2019). Plants generally contain chemical compounds (such as saponins, tannins, oxalates, phytates, trypsin inhibitors, flavonoids, and cyanogenic glycosides) known as secondary metabolites, which are biologically active (Azu and Onyeagba, 2007). Secondary metabolites may be applied in nutrition and as pharmacologically active agents. Flavonoids (quercetin) have inhibitory activity against disease-causing organisms in animals. In vitro studies show that flavonoids also have anti-allergic, anti-inflammatory, antimicrobial, anti-cancer, and anti-diarrheal activities (Kela et al., 2023).

**Conclusion**

It may be concluded from this study that *Aspergillus niger*, *Rhizopus stolonifer,* *Rhizopus microspores*, *Mucor hiemalis,* and *U. botrytis* are common pathogenic fungi that cause apple fruit rot in the study area (Mubi North). The result from the pathogenicity test indicated that all the isolated fungi are pathogenic and attributed to the cause of apple rot in Mubi North. The inhibitory effect of the plant extracts against fungal isolates could be due to the presence of antifungal substances in the extracts. Higher inhibition of fungal growth observed at higher concentrations of both aqueous and ethanolic extracts was recorded. The result also indicates that ethanolic extracts of ginger have more inhibitory compounds than the aqueous extracts. This shows a clear indication of the potential of plant extracts to control fungal pathogens. It is also clear from the result that both the test ginger extracts significantly reduce the radial growth of isolated fungi. It seems that the antifungal effects are a result of many compounds acting synergistically. This can be formulated and successfully used to produce fungicides with local technology, which can be applied in both pre-and post-harvest fruit management.

**Disclaimer (Artificial intelligence)**

Authors hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

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