

Original Research Article

Nutritional Potential of Watermelon (*Citrullus lanatus* (Thunb.) Matsum. & Nakai) Genotypes with Different Flesh Colours

ABSTRACT

Watermelon is a popular and widely consumed fruit that is thirst-quenching and nutritious, containing abundant health-promoting phytonutrients, minerals, and antioxidants. The present study assessed the nutritional potential of twenty watermelon genotypes with different flesh colours, such as red, pink, orange, yellow, and white. This study was conducted in the experimental field of the Department of Vegetable Science at Kerala Agricultural University, Thrissur during two growing seasons (December-March) of 2021-2022 and 2022-2023. A significant variation ($p=0.05$) was seen in nutritional quality characteristics such as lycopene, beta-carotene, citrulline, and ascorbic acid levels across various watermelon genotypes with different flesh colours. Lycopene and beta-carotene content varied from 0.037 to 66.963 $\mu\text{g/g}$ and 0.037 to 7.423 $\mu\text{g/g}$, respectively, based on fresh weight. Citrulline content ranged from 843.810 ppm to 2589.750 ppm. Ascorbic acid content ranged from 21.410 to 49.827 mg/kg on a fresh weight basis. The findings indicated that watermelon genotypes with red flesh are good source of lycopene and ascorbic acid. While the highest beta-carotene content was found in orange flesh genotypes. Both orange and yellow flesh watermelon genotypes exhibited supremacy of citrulline content compared to those with red and pink flesh. White-fleshed watermelon genotypes exhibited the lowest levels of lycopene and beta-carotene content.

Keywords: watermelon; fruit quality; lycopene; beta-carotene; citrulline; ascorbic acid.

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1. INTRODUCTION

Watermelon (*Citrullus lanatus*), a major cucurbitaceous vegetable crop with a chromosome number of $2n=2X=22$, is primarily cultivated for its fresh and nutritious fruit. It is largely consumed as a refreshing summer fruit, much appreciated because of its refreshing capability, attractive colour, delicate taste, and high water content to quench the summer thirst (Asfaw, 2022). The fruit is abundant in lycopene and possesses a total antioxidant capacity comparable to that of tomatoes (Perkins-Veazie *et al.*, 2001). Additionally, the fruits serve as an excellent source of beta-carotene, vitamins (B, C, and E), minerals (K, Mg, Ca, and Fe), amino acid (citrulline) and phenolic compounds.

Flesh colour is a key quality trait that determines attractiveness and signifies the health-promoting benefits of watermelon (Bang *et al.*, 2010). A variety of flesh colours are available in watermelon including white, yellow (pale, canary, salmon), orange and red (King *et al.*, 2009). These diverse flesh colours not only add visual appeal but also have nutritional significance, as they are based on the carotenoid composition and content (Song *et al.*, 2023).

Among carotenoids, beta-carotene is an important dietary source of vitamin A, which is essential for good eyesight. Generally, higher beta-carotene content would enhance the nutritive value of the fruits (Venkatesan *et al.*, 2016). Lycopene imparts red colour in watermelon and has received much scientific attention in recent years due to its strong antioxidant properties (Edwards *et al.*, 2003). Research indicates that lycopene is beneficial in human diet for the prevention of cardiovascular diseases as well as certain types of cancer and it may protect the skin from ultraviolet light damage. Many of these advantageous effects have been linked to the presence of phytochemicals with antioxidant effects (Perera and Yen, 2007).

Citrulline is the most abundant amino acid found in ripe watermelon (Joshi *et al.*, 2019; Zhong *et al.*, 2019). This non-essential amino acid is generated as an intermediate in the urea cycle (Bahri *et al.*, 2013). Citrulline acts as a precursor to arginine, another important amino acid present in watermelon (Joshi *et al.*, 2019; Song *et al.*, 2020). This vital amino acid has a substantial role in the immune, gastrointestinal, respiratory, pulmonary, renal, and hepatic systems, and it also supports wound healing (Wu *et al.*, 2000; Flynn *et al.*, 2002; Collins *et al.*, 2007). Additionally, citrulline may contribute to vasodilation and cardiovascular functions, as arginine is a conditionally essential amino acid linked to these processes (Levine *et al.*, 2012; Hong *et al.*, 2015). Studies suggest that the profiles of citrulline and arginine are crucial in fighting cancer (Lam *et al.*, 2009; Di *et al.*, 2022), heart disease (Tang *et al.* 2009; Hong *et al.*, 2015), acute hydrocephalus (Perez-Neri *et al.*, 2007), minor intestinal disorders, blood poisoning, trauma, and pulmonary hypertension (Beyer *et al.*, 2008; Santarpia *et al.*, 2008), indicating their utility in healing various ailments.

Ascorbic acid is the active form of vitamin C that can impart a sour taste and its concentration differs across various species of fruits and vegetables (Manchali *et al.*, 2021; Soumya and Rao, 2014). From a nutritional perspective, it is important due to its antioxidant properties (Dhillon *et al.*, 2019). Humans and other primates can no longer synthesize vitamin C, making it necessary to obtain it through diet. The suggested daily consumption (SDC) of ascorbic acid for non-smoking adults is established at 75 mg/day for women and 90 mg/day for men (Monsen, 1996). Hence, the current study was undertaken to assess the nutritional composition levels of lycopene, beta-carotene, citrulline, and ascorbic acid in watermelon fruits with varying flesh colours.

2. MATERIALS AND METHODS

The current research titled "Nutritional potential of watermelon (*Citrullus lanatus* (thunb.) Matsum. & nakai) genotypes with different flesh colours" was conducted in the experimental field of the Department of Vegetable Science at Kerala Agricultural University, Thrissur, situated at 10.5° North latitude and 76.2 ° East longitude, with an elevation of 11.13 m above mean sea level. This region experiences a warm and humid tropical climate. The results presented here represent the mean of two growing seasons (December – March) of 2021-2022 and 2022-2023. A total of twenty watermelon genotypes exhibiting various flesh colours (Fig. 1), such as red (Arka Manik, Arka Muthu, Asahi Yamato, CL-4, Sugar baby, and AHW/BR-43), pink (AHW/BR-5, AHW/BR-6, AHW/BR-7, AHW-19, and AHW-65), orange (CL-17, CL-18, CL-19 and CL-14), yellow (CL-5, CL-7, and CL-10), and white (AHW/BR-9 and AHW/BR-13), were included in this investigation (table 1).



Fig. 1. Watermelon genotypes with various flesh colours

Table 1. Source of watermelon genotypes with different flesh colours

Sl. No.	Watermelon genotype	Flesh colour	Source
1	Arka Manik	Red	IIHR
2	Arka Muthu	Red	IIHR
3	Asahi Yamato	Red	IIHR
4	CL-4	Red	KAU
5	Sugar Baby	Red	KAU
6	AHW/BR-43	Red	CIAH
7	AHW/BR-5	Pink	CIAH
8	AHW/BR-6	Pink	CIAH
9	AHW/BR-7	Pink	CIAH
10	AHW-19	Pink	IIHR
11	AHW-65	Pink	IIHR
12	CL-17	Orange	KAU
13	CL-18	Orange	KAU
14	CL-19	Orange	KAU
15	CL-14	Orange	KAU
16	CL-5	Yellow	KAU
17	CL-7	Yellow	KAU
18	CL-10	Yellow	KAU
19	AHW/BR-9	White	CIAH
20	AHW/BR-13	White	CIAH

IIHR- Indian Institute of Horticultural Sciences, Bengaluru, KAU – Kerala Agricultural University, Thrissur, CIAH – Central Institute of Arid Horticulture, Bikaner.

The seeds of selected watermelon genotypes underwent a water bath treatment for 3 hours at 40°C and were soaked overnight in 1% KNO₃ before sowing in portrays filled with cocopeat, vermiculite and perlite in a 3:1:1 ratio. Two weeks old seedlings were transplanted into the main field at a spacing of 2m x 1m. The experiment was laid out in a randomized block design with three replications, containing six plants in each replication. Three fruits from each treatment replication were randomly harvested 30-35 days after pollination and the following biochemical analyses were performed.

The lycopene and beta-carotene content in individual fruits were assessed using a spectrophotometer set at wavelengths of 452 nm and 503 nm, respectively (Ranganna, 1986). The citrulline concentration in the dried juice powder of the watermelon samples were measured using gradient HPLC as described by Ridwan *et al.* (2018). Initially, the fruit juice was lyophilized and freeze-dried to obtain the dried form. A sample weighing 14–26 mg of the freeze-dried substance was used, with 10 ml of deionized filtered water to dissolve it. Meanwhile, a 1 mg/ml stock solution for the citrulline standard was prepared by dissolving it in deionized filtered water. The analysis was conducted using an Agilent Technologies 1200 infinity Series HPLC system. Ascorbic acid content in fresh watermelon samples were determined by volumetric method (Sadasivam and Manickam, 1992).

The data obtained from the various nutritional quality parameters were analyzed using analysis of variance (ANOVA) and statistical evaluation was conducted through Least Square Design (LSD) with KAU GRAPES software version 1.1.1 at a 5% significance level.

3. RESULTS AND DISCUSSION

The findings regarding different nutritional quality parameters of watermelon genotypes with varying flesh colours are shown in Table 2. A significant variation ($P=0.05$) was found among the watermelon genotypes concerning their lycopene, beta-carotene, citrulline, and ascorbic acid levels. The findings revealed that the lycopene content in watermelon flesh exhibited a considerable variation among the tested genotypes, ranging from 0.037 to 66.963 µg/g. The red-fleshed watermelon varieties showed the highest lycopene content, followed by pink-fleshed types, whereas white-fleshed genotypes had the lowest levels, trailed by yellow-fleshed types. Several studies have recorded similar results regarding lycopene content in watermelon flesh. Lycopene is the main carotenoid present in watermelons with red and pink flesh (Tadmor *et al.*, 2005; Kang *et al.*, 2010). Naz *et al.* (2014) reported that watermelon is a potential source of lycopene and red-fleshed watermelons have a lycopene concentration that is over 40 per cent greater than that of tomatoes. Choo and Sin (2012) evaluated the lycopene levels in red and yellow-fleshed seeded watermelons and noted that red-fleshed varieties contained the highest lycopene (9.5 mg/kg) compared to yellow-fleshed ones (0.04 mg/kg). Perkins-Veazie *et al.* (2006) also found that lycopene content varied significantly among cultivars; in red-fleshed varieties, it ranged from 33.9 to 75.7 µg g⁻¹ (mean value 50 µg g⁻¹), while in the two yellow-fleshed varieties, it was below 5 µg g⁻¹. According to Tadmor *et al.* (2005), red-fleshed watermelons are rich in lycopene and have very little beta-carotene (less than 5 percent). Commercial red-fleshed watermelons have been reported to contain lycopene levels of 45.1-53.2 µg g⁻¹ fresh weight (FW), with a mean value of 48.2 µg g⁻¹ (Zhao *et al.*, 2013). Perkins-Veazie *et al.*, (2007) reported that only red-fleshed watermelons contained significant amounts of lycopene, while yellow and orange-fleshed types contained less than 6 mg/kg. Shonima, a red-fleshed seedless watermelon released from Kerala Agricultural University exhibited highest lycopene content which was on par with red fleshed seeded Durgapur Lal, whereas the lowest (0.03 mg/100g) was found in the yellow-fleshed seedless watermelon Swarna (Kaladhar *et al.*, 2024).

Beta-carotene serves as a crucial dietary source of vitamin A (Haskell, 2012; Tang, 2012). Higher beta- carotene content would enhance the nutritional value of the fruits (Venkatesan *et al.*, 2016). Significant differences in beta-carotene among the different watermelon genotypes were recorded, with levels ranging from 0.037 to 7.423 µg g⁻¹. The highest beta-carotene content was recorded in orange-fleshed watermelon genotypes, while the lowest levels were recorded in white-fleshed genotypes. Distinct carotenoid patterns were observed in red-

fleshed and yellow-fleshed watermelons. The red-fleshed watermelon varieties contain high lycopene and varying amounts of beta-carotene (Tadmor *et al.*, 2005). The flesh of immature white watermelons contains only minimal quantities of carotenoids as a result of the differentiation of chromoplasts from non-photosynthetic plastids, which probably develop directly from undifferentiated proplastids (Wang *et al.*, 2019). The accumulation of carotenoids is linked to the development of flesh colour (Tadmor *et al.*, 2005; Bang *et al.*, 2007, 2010). Yellow-fleshed watermelon primarily contains violaxanthin and neoxanthin, while orange-fleshed varieties are mainly composed of beta-carotene (Tadmor *et al.*, 2005; Lewinsohn *et al.*, 2006; Bang *et al.*, 2007, 2010; Liu *et al.*, 2012). White-fleshed watermelons accumulate colourless phytofluene and light-yellow ζ -carotene (Kang *et al.*, 2010; Grassi *et al.*, 2013).

The findings indicated that there were statistically significant differences in citrulline content among watermelon genotypes, ranging from 843.810 ppm to 2589.750 ppm. The highest citrulline content was noted in orange-fleshed genotypes followed by yellow-fleshed types, while the lowest content was observed in pink and red-fleshed genotypes. These findings are in line with Rimando and Perkins-Veazie (2005), who stated that citrulline, a non-essential amino acid, is found abundantly in watermelon, particularly in yellow and orange varieties, which have higher concentrations compared to red ones. According to Kaladhar *et al.* (2024), Swarna, a yellow-fleshed seedless watermelon hybrid released from Kerala Agricultural University, exhibited higher citrulline content compared to the red-fleshed seedless hybrid shonima. Therefore, the current study indicates the supremacy of citrulline content in orange and yellow-fleshed watermelon genotypes compared to red and other fleshed genotypes.

A significant variation in ascorbic acid content among different watermelon genotypes was recorded, with values ranging from 21.410 to 49.827 mg/kg of fresh watermelon. The highest ascorbic acid levels were noted in red-fleshed genotypes, while the lowest were found in white-fleshed genotypes. Comparable results regarding ascorbic acid levels in watermelon genotypes with varying flesh colours were reported by Sultana *et al.* (2023) and Choo and Sin (2012), who found that red-fleshed watermelons had a higher ascorbic acid content than yellow-fleshed types.

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Table 2. Nutritional quality parameters of watermelon genotypes with different flesh colours

Sl. No.	Watermelon genotypes	Lycopene (µg/g)	Beta-carotene (µg/g)	Citrulline (ppm)	Ascorbic acid (mg/kg)
1	Arka Manik	60.517	6.030	2014.430	49.827
2	Arka Muthu	66.963	6.397	1791.677	49.010
3	Asahi Yamato	49.167	6.153	1968.610	43.717
4	CL-4	47.200	2.630	1531.617	38.580
5	Sugar Baby	45.880	4.110	1403.710	37.670
6	AHW/BR-43	58.297	4.677	2248.297	42.527
7	AHW/BR-5	43.273	2.837	843.810	33.903
8	AHW/BR-6	32.707	1.173	1607.337	32.510
9	AHW/BR-7	31.723	1.373	1490.747	30.237
10	AHW-19	39.697	2.210	1472.273	34.603

11	AHW-65	36.233	2.230	1463.320	32.160
12	22K YF 200	17.350	7.390	2348.930	43.680
13	22K YF 201	15.033	7.163	2240.023	40.603
14	22K YF 202	19.443	7.423	2589.750	44.467
15	CL-14	16.237	6.910	2310.337	38.553
16	CL-5	3.447	4.290	2313.883	33.727
17	CL-7	4.213	4.653	2290.803	33.863
18	CL-10	4.553	4.583	2121.063	36.870
19	AHW/BR-9	0.086	0.173	1357.057	26.640
20	AHW/BR-13	0.037	0.037	1750.617	21.410
	SE(m)	0.681	0.055	27.908	0.952
	CD (0.05)	1.951	0.157	79.898	2.725
	CV (%)	3.987	2.297	2.602	4.429

Pearson's correlation matrix revealed the relationships among nutritional quality attributes in watermelon fruits (Table 3). In this analysis, the lycopene content in watermelon exhibited a significant positive correlation ($p=0.01$) with the ascorbic acid content. At the same time, it showed a weak positive correlation with the beta-carotene content and a slight negative correlation with the citrulline content. Furthermore, the beta-carotene content showed a strong positive correlation with both citrulline and ascorbic acid contents.

Table 3. Pearson's correlation coefficients between nutritional quality attributes in watermelon

Nutritional quality attributes	Ascorbic acid	Citrulline	beta- carotene
Citrulline	0.449		
beta- carotene	0.845**	0.714**	
Lycopene	0.596**	-0.302	0.162

** Correlation is significant at 0.01 level (two tailed)

4. CONCLUSION

Nutritional quality parameters such as lycopene, beta- carotene, citrulline and ascorbic acid content of twenty watermelon genotypes with five different flesh colours (red, pink, orange, yellow and white) were assessed, leading to the following conclusions drawn from the findings. The red flesh of cultivated watermelons is due to lycopene accumulation, while orange flesh results from beta-carotene accumulation. Watermelons with orange and yellow flesh exhibit higher levels of citrulline content. In contrast, red-fleshed genotypes exhibit higher levels of ascorbic acid. Thus, watermelon genotypes with diverse flesh colours could be utilized for developing new hybrids with enhanced nutritional qualities.

REFERENCES

1. Asfaw, M. D. (2022). Review on watermelon production and nutritional value in Ethiopia. *Journal of Nutrition Science Research*, 7(4), 173.
2. Perkins-Veazie, P., Collins, J. K., Pair, S. D. & Roberts, W. (2001). Lycopene content differs among red-fleshed watermelon cultivars. *Journal of the Science of Food and Agriculture*, 81, 983-987. doi:10.1002/jsfa.880
3. Bang, H., Davis, A., Kim, S., Leskovar, D. and King, S. (2010). Flesh color inheritance and gene interactions among canary yellow, pale yellow, and red watermelon. *Journal of the American Society for Horticultural Science*, 135, 362-368.
4. King, S. R., Davis, A. R. & Bang, H. (2009). New flesh colors in watermelon?. *HortScience*, 44, 576.
5. Song, H., Lee, K., Subburaj, S., McGregor, C., & Lee, G. (2023). CIPSY1 gene-based SNP markers identified from whole-genome resequencing for the determination of orange flesh color and carotenoid content in watermelon. *Scientia Horticulturae*, 318, 112120.
6. Venkatesan, K. B., Reddy, B. M., & Senthil, N. (2016). Evaluation of Muskmelon (*Cucumis melo* L.) genotypes for growth, yield and quality traits. *Electronic Journal of Plant Breeding*, 7(2), 443-447.
7. Edwards, A. J., Vinyard, B. T., Wiley, E. R., Brown, E. D., Collins, J. K., Perkins-Veazie, P., et al. (2003). Consumption of watermelon juice increases plasma concentrations of lycopene and beta-carotene in humans. *Journal of Nutrition*, 133(1), 043-50.
8. Perera, C. O. & Yen, G. M. (2007). Functional properties of carotenoids in human health. *International Journal of Food Properties*, 10, 201-30.
9. Joshi, V., Joshi, M., Silwal, D., Noonan, K., Rodriguez, S., & Penalos, A. (2019). Systematised biosynthesis and catabolism regulate citrulline accumulation in watermelon. *Phytochemistry*, 162, 129-140. doi: 10.1016/j.phytochem.2019.03.003
10. Zhong, Y., Shi, J., Zheng, Z., Nawaz, M. A., Chen, C., Cheng, F., et al. (2019). NMR-based fruit metabolomic analysis of watermelon grafted onto different rootstocks under two potassium levels. *Scientia Horticulturae*, 258, 108793. doi: 10.1016/j.scienta.2019.108793
11. Bahri, S., Zerrouk, N., Aussel, C., Moinard, C., Crenn, P., Curis, E., et al. (2013). Citrulline: from metabolism to therapeutic use. *Nutrition Journal*, 29, 479-484. doi: 10.1016/j.nut.2012.07.002
12. Song, Q., Joshi, M., Dipiazza, J., & Joshi, V. (2020). Functional relevance of citrulline in the vegetative tissues of watermelon during abiotic stresses. *Frontiers in Plant Science*, 11, 512. doi: 10.3389/fpls.2020.00512
13. Wu, G., Meininger, C. J., Knabe, D. A., Bazer, F. W., & Rhoads, J. M. (2000). Arginine nutrition in development, health and disease. *Current Opinion in Clinical Nutrition & Metabolic Care*, 3(1), 59-66. <https://doi.org/10.1097/00075197-200001000-00010>
14. Flynn, N., Meininger, C., Haynes, T., & Wu, G. (2002). The Metabolic Basis of Arginine Nutrition and Pharmacotherapy. *Biomedicine & Pharmacotherapy*, 56(9), 427-438. [https://doi.org/10.1016/s0753-3322\(02\)00273-1](https://doi.org/10.1016/s0753-3322(02)00273-1)
15. Collins, J. K., Wu, G., Perkins-Veazie, P., Spears, K., Claypool, P. L., Baker, R. A., et al. (2007). Watermelon consumption increases plasma arginine concentrations in adults. *Nutrition*, 23(3), 261-266. <https://doi.org/10.1016/j.nut.2007.01.005>
16. Levine, A. B., Punihale, D., & Levine, T. B. (2012). Characterization of the Role of Nitric Oxide and Its Clinical Applications. *Cardiology*, 122(1), 55-68.

doi:10.1159/000338150

17. Hong, M. Y., Hartig, N., Kaufman, K., Hooshmand, S., Figueroa, A., & Kern, M. (2015). Watermelon consumption improves inflammation and antioxidant capacity in rats fed an atherogenic diet. *Nutrition Research*, 35(3), 251–258. <https://doi.org/10.1016/j.nutres.2014.12.005>
18. Lam, T. L., Wong, G. K., Chong, H. C., Cheng, P. N. M., Choi, S. C., Chow, T. L., *et al.* (2009). Recombinant human arginase inhibits proliferation of human hepatocellular carcinoma by inducing cell cycle arrest. *Cancer Letters*, 277, 91–100.
19. Di Sano, C., Lazzara, V., Durante, M., D'Anna, C., Bonura, A., Dino, P., *et al.* (2022). The Protective Anticancer Effect of Natural Lycopene Supercritical CO₂ Watermelon Extracts in Adenocarcinoma Lung Cancer Cells. *Antioxidants*, 11, 1150. doi.org/10.3390/antiox11061150
20. Tang, W. H., Wang, Z., Cho, L., Brennan, D. M., & Hazen, S. L. (2009). Diminished global arginine bioavailability and increased arginine catabolism as metabolic profile of increased cardiovascular risk. *Journal of the American College of Cardiology*, 53, 2061–7.
21. Perez-Neri, I., Castro, E., Montes, S., Boll, M. C., Coll, J. B., Soto-Hernández, J. L., *et al.* (2007). Arginine, citrulline and nitrate concentrations in the cerebrospinal fluid from patients with acute hydrocephalus. *Journal of Chromatography B*, 851, 250–256.
22. Beyer, J., Kolditz, M., Ewert, R., Rubens, C., Opitz, C., Schellong, S., *et al.* (2008). L-arginine plasma levels and severity of idiopathic pulmonary arterial hypertension. *Vasa*, 37, 61–67.
23. Santarpia, L., Catanzano, F., Ruoppolo, M., Alfonsi, L., Vitale, D. F., Pecce, R., Pasanisi, F., *et al.* (2008). Citrulline blood levels as indicators of residual intestinal absorption in patients with short bowel syndrome. *Annals of Nutrition and Metabolism*, 53, 137–142.
24. Manchali, S., Murthy, K. N. C., Vishnuvardana, & Patil, B. S. (2021). Nutritional composition and health benefits of various botanical types of melon (*Cucumis melo* L.). *Plants*, 10, 1755.
25. Soumya, V., & Rao, T. V. R. (2014). Nutritional quality evaluation of four icebox cultivars of watermelon fruit during their development and ripening. *International Food Research Journal*, 21(2), 631–639.
26. Dhillon, N. S., Sharma, P., Kumar, P., & Sharma, V. (2019). Comparative performance of tomato genotypes for yield and quality characters under protected environment. *International Journal of Chemical Studies*, 7(3), 1678–1680.
27. Monsen, E. R. (1996). New dietary reference intakes proposed to replace the recommended dietary allowances. *Journal of American Dietetic Association*, 96, 754–755.
28. Ranganna, S. (1986). *Manual for Analysis of Fruit and Vegetable Products*, pp 83–104. Tata McGraw Hill, New Delhi.
29. Ridwan, R., Abdul Razak, H. R., Adenan, M. I., & Md Saad, W. M. 2018. Development of isocratic RP-HPLC method for separation and quantification of L-citrulline and L-arginine in watermelons. *International Journal of Analytical Chemistry*, 1–9.
30. Sadasivam, S., & Manickam, A. (1992). *Biochemical Methods*. (3rd Ed.). New Age International (P) Ltd., Publishers, New Delhi, 270p.
31. Tadmor, Y., King, S., Levi, A., Davis, A., Meir, A., Wasserman, B., *et al.* (2005). Comparative fruit colouration in watermelon and tomato. *Food Research International* 38, 837–41.
32. Kang, B., Zhao, W., Hou, Y., & Tian, P. (2010). Expression of carotenogenic genes during development and ripening of watermelon fruit. *Scientia Horticulturae*, 124, 368–

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33. Naz, A., Butt, M. S., Sultan, M. T., Qayyum, M. M. N., & Niaz, R. S. 2014. Watermelon lycopene and allied health claims. EXCLI Journal, 13, 650.
34. Choo, W. S., & Sin, W. Y. (2012). Ascorbic acid, lycopene and antioxidant activity of red-fleshed and yellow-fleshed watermelons. Advances in Applied Science Research, 3(5), 2 779–84.
35. Perkins-Veazie, P., Collins, J. K., Davis, A. R. and Roberts, W. (2006). Carotenoid content of 50 watermelon cultivars. Journal of Agricultural and Food Chemistry, 54, 2593- 2597. doi:10.1021/jf052066p
36. Zhao, W., Lv, P., & Gu, H. (2013). Studies on carotenoids in watermelon flesh. Agricultural Science, 4(7A), 13–20.
37. Perkins-Veazie, P., Collins, J. K., Clevidence, B. & Wu, G. (2007). Watermelons and health. Acta Horticulturae, 731,121.
38. Kaladhar, A., Kutty, M. S., Pradeepkumar, T., Johnson, J. M. & Anoop, E. V. (2024). Root colonization by Piriformospora indica promotes growth, induces earliness in flowering, enhances yield and citrulline content in watermelon (*Citrullus lanatus* (Thunb.) Matsum. & Nakai). Journal of Horticultural Sciences ,19(2)
39. Haskell, J. M. (2012). The challenge to reach nutritional adequacy for vitamin A: beta-Carotene bioavailability and conversion-evidence in humans. The American Journal of Clinical Nutrition, 96(5), 1193S-203S.
40. Tang, G. (2012). Techniques for measuring vitamin A activity from beta-carotene. The American Journal of Clinical Nutrition, 96(5), 1185S-1188S.
41. Wang, C., Qiao, A., Fang, X., Sun, L., Gao, P., Davis, A. R., et al. (2019). Fine mapping of lycopene content and flesh color related gene and development of molecular marker–assisted selection for flesh color in watermelon (*Citrullus lanatus*). Frontiers in Plant Science,10. doi: 10.3389/fpls.2019.01240
42. Bang, H., Kim, S., Leskova, D. & King, S. (2007). Development of a codominant CAPS marker for allelic selection between canary yellow and red watermelon based on SNP in lycopene β -cyclase (LCYB) gene. Molecular Breeding, 20, 63-72. doi:10.1007/s11032-006-9076-4
43. Lewinsohn, E., Sitrit, Y., Bar, E., Azulay, Y., Meir, A., Zamir, D., et al. (2006). Carotenoid pigmentation affects the volatile composition of tomato and watermelon fruits, as revealed by comparative genetic analyses. Journal of Agricultural and Food Chemistry, 53, 3142–3148. doi: 10.1021/jf047927t
44. Liu, C., Zhang, H., Dai, Z., Liu, X., Liu, Y., et al. (2012). Volatile chemical and carotenoid profiles in watermelons [*Citrullus vulgaris* (Thunb.) Schrad (Cucurbitaceae)] with different flesh colors. Food Science and Biotechnology, 21, 531-541. doi:10.1007/s10068-012-0068-3
45. Grassi, S., Piro, G., Lee, J. M., Zheng, Y., Fei, Z. J., Dalessandro, G., et al. (2013). Comparative genomics reveals candidate carotenoid pathway regulators of ripening watermelon fruit. BMC Genomics, 14 (1), 781. doi: 10.1186/1471-2164-14-781
46. Rimando, A. M. & Perkins-Veazie, P. M. (2005). Determination of citrulline in watermelon rind. Journal of Chromatography A, 1078(1-2), 196-200.
47. Sultana, H., Mallick, S. R., Hassan, J., Gomasta, J., Kabir, M. H., Sakib, M. S. A., et al. (2023). Nutritional composition and bioactive compounds of mini watermelon genotypes in Bangladesh. Agricultural and Food Sciences, P.22 {https://api.semanticscholar.org/CorpusID:262465705}

Commented [AJ5]: correct references