THE PREVALENCE OF COCCIDIOSIS IN POULTRY FARMS IN UYO METROPOLIS, AKWA IBOM STATE, NIGERIA.

Abstract

Despite the presence of large number of chickens in Nigeria, contribution to national economy or benefit from this sector of activity is very limited due to diseases and management, out of which poultry coccidiosis is a leading problem. This study was conducted to investigate the prevalence and incidence of coccidiosis in poultry farms in Uyo metropolis, Akwa Ibom State, Nigeria. Selected numbers of poultry farms within Uyo metropolis were used for this study. A total of 100 faecal samples from live birds comprising 62 young and 38 adult birds were randomly collected as bird's fresh droppings from both young and adult sections of the farm. Samples were analyzed using standard parasitological methods and techniques. An overall prevalence rate of 46%% was obtained. Higher prevalence rates were recorded in young bird as 48.38% while adult had 42.10%. Then five species of Eimeria that were identified includes E. tennela, E. maxima, E. necatrix, E. alervuhinaand E. brunette at 28%, 25%, 15.6%, 18.8% and 12.5% respectively. E. tennelarecorded highest prevalence followed by E. maxima, while E. brunette was the least. Coccidiosis is a major problem in the farm with inadequate hygienic measures and factors such as age, sex and biosecurity which are the most common factors that contribute for the occurrence of coccidiosis. Therefore, appropriatecontrol strategies should be designed considering important risk factors and focus should be given to biosecurity practices in he prevention and control of coccidiosis, and in addition, further studies are needed to be conducted to identify the prevalentEimeria species for strategic control.

Keywords: Incidence, Coccidiosis, Poultry Farms, Uyo, Akwa Ibom State, Nigeria.

INTRODUCTION

1.1 Introduction

In Agriculture, poultry has a wide distribution in and out of Africa. Among members of the poultry family, fowl or chicken is the most common with approximately 98% of the total poultry population in Africa (UNDESA, 2017). Over the decades, the world has been experiencing a continuous growth in human population. According to UN DESA (2017), the current human population of 7.6 billion has been projected to a population of 9.8 and 11.2 billion respectively by 2050 and 2100. To meet this ever-growing population, food, shelter and clothing become imperative. Food production carries a larger percentage of these basic human needs as its availability determines the existence of human population (Erb *et al.*, 2012). In view of this, demand has been on the agricultural sector of each nation to increase food production that is safe for human consumption that will meet the ever-growing population.

Poultry industry is one of the fastest growing sub-sectors of agriculture that contributes to global nutrition (Mottet and Tempio 2017) and thus a major driving force of the economy. Chicken, a major poultry bird contributes greatly to agricultural production through the supply of meat and eggs (Hald, 2010). However, chickens are also host to many deadly diseases which hamper on productivity and compromise welfare resulting in high mortality in

Comment [KO1]: Italize

Comment [KO2]: A repeat, can be deduced from the preceding sentence

some cases. Among many diseases that affect chickens globally, coccidiosis is a house-hold name associated with high level of mortality in poultry industry (Blake and Tomley 2014).

Poultry play a vital role through their contribution to the socio-economic and cultural lives of small-holder farmers (Nyoni and Masika 2012). Village chickens can be used as tokens of appreciation for services rendered and are often given to visitors as gifts (Kusina and Kusina 1999). Their role in national economies is through improved nutritional status and income of many small-holder farmers as well as landless and marginalized communities (Tarwireyi and Fanadzo 2013).

*Gallus gallus*Linnaeus (1758) commonly called domestic fowl or chicken is reared by urban and rural farmers who use their eggs and meat as a source of animal protein and farm manure (Frantovo 2000). The greatest impediment to poultry production in Nigeria is diseases (Lawal *et al.*, 2001). Parasites can be a significant factor limiting the productivity of chickens by affecting the growth rate of chickens and causes organ damage and eventually death (Negbenebor and Ali, 2018). The *Gallus domesticus* can easily be infested with several types of bacterial, viral, fungal and parasitic pathogen (Tarwireyi and Fanadzo, 2013). Parasites are among the infectious agents that cause an alarming problem to the industry, posing adverse economic effects. Gastrointestinal parasitism leads to significant economic losses in poultry (Aklilu, 2007).

The parasite colonizes and infect the intestinal tract of different animals and birds (Haug *et al.*, 2008) and the infection is been established through ingestion of feed or water contaminated with sporulated oocycts (Allen and Fetterer, 2002). Infection with this parasite is characterized by dysentery, enteritis, diarrhea, which may bloody with certain *Eimeria species*, lower feed intake, interference with normal digestion and nutrient absorption, emaciation, dropping wings, and delayed sexual maturation (Rehman *et al.*, 2010; Awais *et al.*, 2012).

Poultry coccidiosis, caused by the protozoan parasite of the genus Eimeria, remains one of the most important parasitic diseases in poultry industry worldwide (Bera *et al.*, 2011; Jadhav *et al.*, 2011). Several studies established the prevalence and economic importance of coccidiosis as a major parasitic disease in both local and exotic breeds of poultry worldwide (Dakpogan and Salifou, 2013) The prevalence of coccidiosis was reported in many countries such as Iran (Nematollahi *et al.*, 2009), Egypt (Ahmed *et al.*, 2012), Ethiopia (Oljira*et al.*, 2012), India (Sharma *et al.*, 2015); South Africa (Mwale and Masika, 2011) and Nigeria (Musa *et al.*, 2010; Jatau *et al.*, 2012).

These pathogenicity of these species in poultry have caused huge damage and great economic loss to the agricultural industry due to mismanagement (Jones and Prescott,2020), ratio, weight loss, decreased feed conversion, low egg production and death in young birds as a result of damage intestine (Balqis, 2013). Primarily, farmers have been forced to either keep the business or hold or completely out of business as a result of high mortality. Poultry mismanagement is one of the major causes for the prevalence of parasitic infections. This resulted to 64% infection rate of 300 chickens in Uyo metropolis (Usip*et al.*, 2017). Therefore, it is important to understand the current rate of prevalence of coccidiosis, thereby developing new or improving already existing methods of prevention, control and treatments in poultry farms in Uyo. This study will provide resourceful information in regards the coccidia species and site of infestation in the local fowl. Additionally, it is pertinent for prompt disease diagnosis for further actions. The main objective of this study is to investigate

Comment [KO3]: Already covered by dysentery

Comment [KO4]: Italize all scientific names
Comment [KO5]:

the prevalence and incidence of coccidiosis in poultry farms in Uyo metropolis, Akwa Ibom State, Nigeria.

2.0 MATERIALS AND METHODS

2.1. Study Area

The study was conducted in Uyo, Akwa Ibom State Uyo metropolis. Uyo is the capital of Akwa Ibom State, in the South South region of Nigeria which stands at a total area of about 140sq miles. It lies approximately between latitude 5° 2' 51 North and Longitude 8° 2' 41 East. The climate condition of the study area is characterized by both rainy and dry seasons, which ranges between March to October and November to February respectively. The annual rainfall is about 2500 m with a mean annual temperature of 32 °C and a relative humidity of about 75%.

2.2. Sample collection

The poultry farms were visited for specimen collection. Fresh faecal droppings and carcasses were collected in well labelled sterile universal bottles and polythene leather bags respectively and were transported to the Department of Animal and Environmental Biology, Faculty of Biological Sciences, University of Uyo Laboratory for further processes.

2.3. Data collection

Information on temperature and humidity were taken from record kept at the farms. Data on the period of the year when the outbreak was noticed, age of bird affected, clinical signs observed, mortality rates, therapeutic and preventive measures taken were obtained through a well-structured questionnaire.

2.4. Laboratory analysis of samples

This was done by wet mount smears of the faecal droppings as described by Fleck and Moody (1993). Faecal samples were examined for the p. sence of Eimeria oocyst using the floatation techniques. Positive samples were further subjected to the Mcmaster counting technique so as to determine the intensity of Eimeria infection. Post mortem examination was carried out after opening of the carcasses, and the intestine was removed aseptically to notice the presence of lesions and intestine was scraped and observed under the microscope using X10 objective and X40 objective, the method as described by Jordan (1999). Sporulation was performed in wet chamber at 24-26 °C in a 2.5% aqueous solution of potassium dichromate (K₂Cr₂O₇)

2.5. Data Analysis

Data obtained from the prevalence study were analysed using descriptive statistics and the results summarized as percentages. Students' T test was used to analyse the difference between the prevalence of parasitic species. Variation in the prevalence of gastrointestinal parasite in relation to parasite species, sex and age group were analysed by using Chi- square statistics. Probabilities (P) was significant at $p \le 0.05$.

3.0. RESULTS

3.1. Prevalence of Coccidiosis infection from poultry farms in Uyo L.G.A.

Formatted: Highlight

Comment [KO6]: What informed selection of the study area?

Comment [KO7]: Address the following:

1. How many poultry farms were sampled? 2. How were the farms selected to ensure they are representative of study area? 3. What informed the sample size, provide a formula used for sample size calculation 4. How many samples were taken per farm? 5. How were the chickens housed in each farm, single-caged or multiple/group housing? 6. If group housing, did you pool the faecal samples or how did you match the samples with the chickens, and how did you know which sample came from which age group? What measures were taken during sample collection to avoid cross-contamination? 8. How were the samples kept and preserved during transportation to the laboratory? 9. How long were they stored before analysis at the laboratory?

Comment [KO8]: Author(s) has not addressed the following:

- the following: 1. What was the need for the temperature and humidity data? 2. The term 'bird' is confusing. use 'chicken'
 - uniformly throughout the text

Comment [KO9]: Author(s) has not addressed: 1. Why was direct faecal smear (wet mount0 method used despite its known low sensitivity in the initial screening?

 Specify which floatation techniques were used and how they were done? Magnification used, salts used and their concentration as well as preparation methods

3. McMaster technique should have been used to screen all the samples as it is relatively more sensitive than the other floatation methods,

explain why this was not done

4. What do you mean by intestines being removed aseptically in a dead chicken?5.Describe how the intestines were scraped and

examined? Why didn't you use impression smears? 6. How long did the sporulation take? Why was it

important that you do it, how did you use this information to identify the Eimeria species

Comment [KO10]: Prevalence is a categorical data (proportion), Chi-square is more suited for this comparison

Out of the one hundred and twenty specimens collected, that it one hundred faecal droppings and twenty carcasses, 45 faecal droppings were positive for oocysts of coccidiosis and 6 carcasses were positive of different species of Eimeria. Infection rate was higher among the younger birds (Table 1). Number of faecal samples examined in Farm A, B, C, D, E, F, G and J were 11, 16, 10, 12, 11, 9, 6, 8, 10 and 7 respectively. In Farm A-J, the numbers of young sample examined were more than the adult except Farm G that recorded same (3 each). For Farm A, B, C, D, E, F, G and J, the prevalence of infection in young and adult were 50.0 % and 33.3%, 50.0% and 50.0%, 57.1% and 67.0%, 38.0% and 25.0%, 50.0% and 40.0%, 80.0% and 25.0%, 33.3% and 33.3%, 40.0 and 67.0%, 50.0% and 25.0%, and 25.0% and 67.0% respectively.

Table 1: Showing the prevalence of coccidiosis infection from poultry farms in Uyo L.G.A.

Farms	No. of faecal droppings examined	6 /	Age	Prevalence of infection according to Age		
		Young	Adult	Young (%)	Adult (%)	
Α	11	8	3	4 (6.45)	1 (2.63)	
В	16	10	6	5 (8.10)	3 (7.89)	
С	10	7	3	4 (6.45)	2 (5.26)	
D	12	8	4	3 (4.83)	1 (2.63)	
Ε	11	6	5	3 (4.83)	2 (5.26)	
F	9	5	4	4 (6.45)	1 (2.63)	
G	6	3	3	1 (1.61)	1 (2.63)	
Η	8	5	3	2 (3.22)	2 (5.26)	
Ι	10	6	4	3 (4.83)	1 (2.63)	
J	7	4	3	1 (1.61)	2 (5.26)	
Total	100	62	38	30 (48.38)	16 (42.10)	
Total Prevalence				46 (46%)		

confusing as details that are supposed to be under materials and methods are mixed with results. P-values and X^2 are given below the tables but we don't know what they are comparing. Need a thorough reorganization

Comment [KO11]: Section is poorly written and

Significant at $p \le 0.05$; $X^2 = 14.54$

Table 2: Showing the different Eimeria species identified in this study

Farms	No. of	E. tennela	E. maxima	E. necatrix	E. alervuhina	E. brunette	Total
	Carcasses	(%)	(%)	(%)	(%)	(%)	
Α	4	2	1	0	2	1	6
В	2	1	1	2	0	0	4
С	2	1	0	0	1	1	3
D	1	0	1	0	1	0	2
Ε	0	0	0	0	0	0	0
F	3	2	1	1	0	1	5
G	2	1	1	1	1	0	4
Η	2	1	1	1	0	0	3
Ι	1	0	0	0	0	1	1
J	3	1	2	0	1	0	4
Total	20	9 (28)	8 (25)	5 (15.6)	6 (18.8)	4 (12.5)	32

Significant at $p \le 0.05$; $X^2 = 10.72$

3.2. Different Eimeria species identified and their percentage intensity

Table 2 reveals 5 species of Eimeria to be prevalent in the 10 farms under study. They include *E. tennela, E. maxima, E. necatrix, E. alervuhina* and *E. brunette*. For the carcasses used for the analysis, farm E recorded 0, thus no presence of parasite. Farm A recorded the highest number of carcasses (4) with all species present except *E. necatrix*. Farm F and J recorded 3, with all species present except *E. alervuhina* and *E. necatrix* and *E. brunette* respectively. Farm B, C, G and H recorded 2, with all species present except *E. alervuhina* and *E. brunette* respectively. The highest and lowest prevalence of coccidiosis were *E. tenella* and *E. brunette*.

3.0. DISCUSSION, CONCLUSION AND RECOMMENDATIONS

3.1. Discussion

Epidemiological studies on the prevalence and intensity of Eimeria species are useful tools for the prevention and control of coccidiosis and specifically in Nigeria (Ola-Fadunsin and Ademola 2013). It has been reported from consistent literatures that coccidiosis is the most common parasitic disease of poultry and also a major constraint to the success of poultry farming in Nigeria (Lawal *et al.*, 2016).

The prevalence of coccidiosis in this study revealed *E. tennela* (28%), *E. maxima* (25%), *E. necatrix*(15.6%), *E. alervuhina* (18.8%) and *E. brunette* (12.5%) in both young and adult poultrysampled in 10 different farms in Uyo, Akwa Ibom State, Nigeria and were significantly different at p<0.05.

A total of 100 droppings of poultry were observed. The prevalence of coccidiosis in this research was 46% which corroborates with previous reports of Lucas and Zainab (2016) in Gombe that reported 42.7%, Agishiet al., (2016) that reported 40.1% at Slaughter Houses in Makurdi, Benue State, Usman et al., (2022) in Bauchi that reported 43.3%. Generally, the variation in the prevalence in coccidiosis in poultry is likely attributed to a range of factors which includes sampling periods (rainy or dry), sample size (number of concerned samples, geographical area (tropical or temperate) and climatic conditions observed in different study areas. It should be borne in mind however, that incidence of coccidiosis is high in highly humid geographical areas like (Akwa Ibom State, Nigeria) explaining the higher prevalence reported in these parts of Nigeria. The relatively high prevalence reported in this work could also be due to poor management practices in the farms. One of these practices as observed by the researcher's during sampling is the perpetual spillage of water on the litters from the poultry drinkers, which encourages Eimeria build-up and subsequent sporulation and infection. This is consistent with the reports by Taylor et al. (2007) and Omer et al., (2011). It is reported that most livestock farmers are not expert and do not seek professional advices and consultation (Okon et al., 2020; Silas et al., 2024).

However, the prevalence of this result was higher than the prevalence of coccidiosis in poultry farm in Maiduguri (Jallailudeen*et al.*, 2016), Eke *et al.*, (2016), Lawal *et al.*, (2016) and Dechas*et al.*, (2023) who reported low (28%, 37.1% and 27.1 respectively) prevalence.

The prevalence of the disease in farms in Uyo metropolis might be attributed to poor sanitation practices and unhygienic environmental conditions which encourages and pampers the survival of oocyts. Five species of *Eimeria* were identified in the present study, *Eimeria tenella9* (28%),*E. maxima* 8 (25%), *Eimeria necatrix*5 (15.6%), *Eimeria acervulina*6 (18.8%) and *E. brunette* 4 (12.5%).

Comment [KO12]: Provide distinguishing features used to identify the different Eimeria species

All these species have been identified in literatures of Haug *et al.*, (2008); Amer *et al.*, (2010); Hadipour *et al.*, (2011); Jatau *et al.*, (2012); Hadipour *et al.*, (2013); Meskerem *et al.*, (2013); Olanrewaju and Agbor (2014); Kaboudi*et al.*, (2016); Agishi*et al.*, (2016) and Dechas*et al.*, (2023). These reports have recorded at least 3 of the Eimeria species which have been identified and recognized poultry in this study. The coccidian parasite, *Eimeria* has special preferences to the chicken's intestinal tract with predilection in different anatomical sites of the tract, thus it prevalence in poultry. It is associated with bloody diarrhea, low productivity due to low feed conversion rates (production losses), reduced welfare of birds and increased mortality resulting from the extensive damage of the gastrointestinal tracts.

The young samples reported higher prevalence than the adult at 48.38% and 42.10% respectively. This study was similar to report recorded by Hembram *et al.*, (2015); Sheikh *et al.* (2016); Lawal *et al.*, 2016); Bandi *et al.*, (2020); Zalizar*et al.*, (2021); Oriaku*et al.*, (2024), who reported the prevalence of coccidiosis to be higher in young than adult poultry. Oriaku*et al.*, (2024) added that high rate of infection in young birds may be due to decreased immunity as well as continuous exposure to infections from the contaminated litter.

Amongst all species of prevalence, *E. tennela* and *E. maxima* had the highest at 28% and 25% respectively. This was in conformity with a study in Bauchi that reported most prevalent of coccidiosis in poultry to be *E. tennela*at 29.1% (Auwal *et al.*, 2022), but recorded *E. maxima* as the least with 5.8% (Auwal *et al.*, 2022; Abdullahi *et al.*, 2023). This result does not agree with Abdullahi *et al.*, (2023) who reported 7.84% and 6.86 for *E. tennela* and *E. maxima* respectively. Lawal *et al.* (2016) in Maiduguri also reported four species of *Eimeria* except for *E. brunetti*. Jatau *et al.* (2012) in his work carried out in Zaria, Nigeria reported about seven species. Muhammad *et al.* (2011) in Iran and Bachaya*et al.* (2015) in Pakistan all reported four Eimerian species except *E. brunetti*which suggests that those species of *Eimeria* are widely distributed in many countries.

The Eimerian species identified in this study are considered to be among the most economical important parasitic species of poultry. Jatau *et al.* (2012) reported that in Nigeria, coccidiosis is caused by *E. tenella, E. necatrix, E. acervulina, E. mitis, E. brunneti,* and *E. praecox.* The identification of *E. tenella and E. necatrix* this study agrees with Abdullahi *et al.*, (2023) who reported that they are the two most pathogenic species of *Eimeria.* Although in this study, *E. brunette* had the lowest prevalence of 12.5%.

5.3. Conclusion

Coccidiosis is an important enteric parasitic disease of poultry associated with significant economic losses to poultry farmers worldwide. It has been indicated that there are several species of *Eimeria* affect chickens with varying pathogenicity. The occurrence of coccidiosis dependent on agent, host and management as well as environment associated risk factors. This study has shown that Eimeria infection is endemic among poultry in Uyo, Akwa Ibom State, Nigeria, as 46% of the sampled farms were infected. The presence of lesions and part of intestine affected in combination with histopathology could help in better diagnosis of coccidiosis. Anticcocidials and good management are important for control and prevention of coccidiosis in domestic chickens.

5.4. Recommendations

Based on above findings and conclusion, farmers are advised to carryout good biosecurity measures such aproper stocking density, keeping litter dry by repairing all leaking watering

trough, appropriate use of anticoccidial drugs prescribed by registered veterinarians, daily cleaning of litters to avoid accumulation of feaces which help in oocysts development, proper prophylactics treatment and management to decrease the impact of coccidiosis on the economy of developing country. These measures when practiced will be effective in reducing the losses induced by the disease. In addition, to control this economically important parasitic disease of poultry, further studies need to be undertaken to come up with sustainable and cost-effective prevention and control methods.

REFERENCES

- Abdullahi, N., Abdulrahman, I. S., Abdulazeez, S. O., Ayuba, S. O., Odey, A. S., Adejoh, V. A. and Pam, A. V. (2023). Prevalence of Avian Coccidiosis and Identification of Eimeria spp in Local Broilers and Chickens in Lafia Modern Market, Nassarawa State, Nigeria. *EAS Journal* of Parasitology and Infectious Diseases. 5 (3):23-35.
- Agishi, G., Luga, I. I. and Rabo, J. S. (2016). Prevalence of Coccidiosis and Eimeria species in Layers and Broilers at Slaughter Houses in Makurdi, Benue State. *The International Journal* of Engineering and Science (IJES), 5(2):08-11
- Ahmed, A. A., Olfat, A. M., Aida, A. N. E. and Mohamed, S. A. (2012). Studies on coccidia of Egyptian Balady breed chickens. *Life Sci. J.*, 9(3): 568-576.
- Aklilu, H. M. (2007). Village Poultry in Ethiopia: Socio-Technical Analysis with Farmers, [Ph.D. thesis], Wageninsen University, Wageningen, Netherlands.
- Allen, P. C. and Fetterer, R. H. (2002). Recent advances in biology and immunology of Eimeria species and in diagnosis and control of infection with coccidian parasites of poultry. *Clin. Microbiol. Rev.*, 15:58-65.
- Amer, M. M., Awaad, M. H. H., El-Khateeb, R. M., Abu-Elezz, N. M. T., Sherein-Said, A., Ghetas, M.M. and Kutkat, M.A. (2010). Isolation and Identification of *Eimeria* from Field Coccidiosis in Chickens. *Journal Am. Science.*, 6(10):1107-1114. -170.
- Auwal, M. M., Nasiru, A. Y., Danyaya, A. I., Abubakar, A. B. and Muhammad, A. A. (2022). Prevalence of Avian Coccidiosis among Exotic Breed of Chickens within Bauchi Metropolis *Iconic Research and Engineering Journals (IRE)*, 6 (6):232-238.
- Awais, M. M., Akhtar, M., Iqbal, Z., Muhammad, F. and Anwar, M. I. (2012). Seasonal prevalence of coccidiosis in industrial broiler chickens in Faisalabad, Punjab, Pakistan. *Trop. Anim. Health Prod.*, 44:323-328.
- Bachaya, H. A., Abbas, R. Z., Raza, M. A., Iqbal, Z., Rehman, T. U., Baber, W. and Hussein, R. (2015). Existence of Coccidiosis and Associated Risk Factors in Broiler Chickens in Southern Punjab, Pakistan. *Pakistan Veterinary Journal*, 35(1), 597-600.
- Balqis, U., Hambal, M., Darmami, S. and Utami, C. (2013). Histological changes in intgestine of chicken (*Gallus gallus*) infected naturally by *Ascaridiagalli*. International Conference on Multidiscipline Research (ICMR).
- Bandi, A., Pattipati, M., Chennuru, S., Pentela, R. and Kokila, S. (2020). A cross-sectional study on gastrointestinal parasites in backyard poultry in Krishna district, Andhra Pradesh, India. *International Journal of Livestock Research* 10(2): 46–60.
- Bera, A. K., Bhattacharya, D., Pan, D., Dhara, A., Kumar, S. and Das, S. K. (2010) Evaluation of economic losses due to coccidiosis in poultry industry in India. *Agric. Econ. Res. Rev.*, 23: 91-96.
- Blake, D. P. and Tomley, F. M. (2014) Securing poultry production from the ever-present Eimeria challenge. *Trends Parasitol* 30:12–19.
- Dakpogan, H. B. and Salifou, S. (2013) Coccidiosis prevalence and intensity in litter based high stocking density layer rearing system of Benin. J. Anim. Plant Sci., 17(2):2522-2526.

- Dechas, M., Adem, D. V. M and Mohammedkemal, M. A.(2023). Factors in and around Haramaya District, Ethiopia. *Vet Med Open J.* 2023; 8(1): 9-17. doi: 10.17140/VMOJ-8-172
- Eke, S. S., Ibeh, E. O., Omalu, I. C. J., Otuu, C. A., Hassan, S. C. and Ubanwa, E. D. (2016). Prevalence of coccidiosis in chickens in three poultry farms in Minna, Niger State. *Nigerian Journal of Parasitology. Volume* 37(2):154-157.
- Erb, K. H., Mayer, A., Kastner, T., Sallet, K. E. and Haberl, H. (2012). The impact of industrial grain fed livestock production on food security: an extended literature review. Commissioned by compassion in world farming. The Tubney Charitable Trust and World Society for the Protection of Animals, London, UK
- Fleck, S. L. and Moody, A. H. (1993). Diagnostics Tecniques in Meidcal Parasitology, 11th ed. Cambridge, Uttar Peadesh, India , p. 10-14
- Frantovo, D. (2000). Some parasitic nematodes (Nematoda) of birds (Aves) in the Czech Republic. Acta SocietatisZoologicaeBohemicae. 66(1):13-28
- Hadipour, M. M., Olyaie, A., Naderi, M., Azad, F. and Nekouie, O. (2011). Prevalence of Eimeria species in scavenging native chickens of Shiraz, Iran. *Afr J Microbiol Res* 5: 3296-3299.
- Hald, T. (2010) Analysis of the baseline survey on the prevalence of Campylobacter in broiler batches and of Campylobacter and Salmonella on broiler carcasses in the EU, 2008, Part A: Campylobacter and Salmonella prevalence estimates. European Food Safety Authority 2010.
- Haug, A., Gjevre, A. G., Thebo, P., Mattsson, J. G. and Kaldhusdal, M. (2008) Coccidial infections in commercial broilers: epidemiological aspects and comparison of Eimeria species identification by morphometric and polymerase chain reaction techniques. *Avian pathol* 37: 161-70.
- Hembram, A., Panda, M. R., Mohanty, B. N., Pradhan, C. R., Dehuri, M., Sahu, A. and Behera, M. (2015). Prevalence of gastrointestinal helminths in Banaraja fowls reared in semi-intensive system of management in Mayurbhanj district of Odisha. *Veterinary World* 8(6): 723–726.
- Jadhav, B. N., Nikam, S. V., Bhamre, S. N. and Jaid, E. L. (2011). Study of *Eimeria necatrix*in broiler chicken from Aurangabad District of Maharashtra State India. *Int.Multidiscip. Res. J.*, 1(11): 11-12.
- Jallailudeen, R. L., Saleh, M. J., Umar I.I., Yaqub, A.G., Isa A.G., Gambo M. and Benjamin U. I. (2016). Prevalence of Coccidiosis among Village and Exotic Breed of Chickens in Maiduguri, Nigeria. Veterinary World, 9(6), 653-659.
- Jatau, I. D., Sulaiman, N. H. and Musa, I. W (2012). Prevalence of coccidian infection and preponderance Eimeria species in free range indigenous and intensively managed exotic chickens during hot-wet season. Asian Journal of Poultry Science; 6(3): 79-88. doi:10.3923/ ajpsaj.2012.79.88
- Jones, E. K. M. and Prescott, N. B. (2020). "Visual cues used in the choice of mate by fowl and their potential importance for the breeder industry". World's Poultry Science Journal. 56 (2): 127– 138.
- Jordan, F., Pattison, M., Alexander, D. and Faragher, T (2002). Parasitic diseases. In: Poultry Disease (5th Edn) Hong Kong, WB Saunders, 405-420.
- Kusina, J. F. and Kusina. N. T. (1999). Feasibility study of agricultural and household activities as they relate to livestock production in Guruve District of Mashonaland central Province with emphasis on village chicken production, Report prepared for Household Agricultural Support Programme, Harare, Zimbabwe.
- Lawal, A. J., Igbozuike, O. O., Okubanjo, O. O. and Natala, A. J. (2001). A comparative study of parasitism in the free range, deep litter and battery cage chickens in Zaria. *Journal of Tropical Biosciences*. 1(1): 89-92
- Lawal, J. R., Jajere, S. M., Ibrahim, U. I., Geidam, Y. A., Gulani, I. A., Musa, G. and Ibekwe, B. U. (2016) Prevalence of coccidiosis among village and exotic breed of chickens in Maiduguri, Nigeria, *Veterinary World*, 9(6), 653-659.

- Lawal, J. R., Jajere, S. M., Ibrahim., U. I. (2016). Prevalence of Coccidiosis Among Village and Exotic Breed of Chickens in Maiduguri, Nigeria *Veterinary World*, 9(6): 653–659,
- Mottet, A. and Tempio, G. (2017) Global poultry production: current state and future outlook and challenges. *World's Poult Sci J.* 73(2):245–256.
- Muhammad, M. H., Ahad, O., Mohammed, N., Fariborz, A. and Omid, N. (2011). Prevalence of *Eimeria* Species in Scavenging Native Chickens of Shiraz, Iran. *African Journal of Microbiology Research*, 5(20), 3296-3299.
- Mwale, M. and Masika, P. (2011) Point prevalence study of gastro-intestinal parasites in village chickens of Centane district, South Africa. *Afr. J. Agric. Res.*, 6: 2033-2038.
- Negbenebor, H. E. and Ali, M. (2018). Prevalence of gastro-intestinal parasites of local chickens (*Gallus gallus domestica*) in Kano, Nigeria. *Ann. Microb. Infec. Dis.* 1(4): 45-49
- Nematollahi, A., Gholamali, M. and Reze, F. P. (2009). Prevalence of *Eimeria* species among broiler chicks in Tabriz, Iran. *Munis. Entomol. Zool.*, 4: 53-58.
- Nyoni, N. M. B. and Masika, P. J. (2012). Village chicken production practices in the Amatola Basin of the Eastern Cape Province, South Africa. *Journal of Agricultural Research* 7, 2647–2652
- Okon, A. O., Udoinyang, E. P. and Essien, E. A. (2020). Growth performance of the African Catfish, *Clarias gariepinus*fingerlings fed four commercial feeds. *Journal of Wetlands and Waste Management*, 4 (1):51-55.
- Ola-Fadunsin, S. D. and Ademola, I. O. (2013)."Direct Effects of MoringaoleiferaLam (Moringaceae) Acetone Leaf Extract on Broiler Chickens Naturally Infected with Eimeria Species," Tropical Animal Health and Production. 45(6) 1423–1428.
- Oljira, D., Melaku, A. and Bogale, B. (2012). Prevalence and risk factors of coccidiosis in poultry farms in and around Ambo Town, Western Ethiopia. *Am. Euras. J. Sci.Res.*, 7(4): 146-149.
- Omer, S. A., Apio, A., Wronski, T. and Mohammad, O. B. (2011) A new coccidian parasite (*Eimeria farasanii*n. sp.) indicates parasite-host specificity in endemic Farasan gazelle. *Int. J. Zool. Res.*, 7: 85-92.
- Oriaku, L. O., Silas, I. I., Nnanna-Chigozie, E. P., Inyang, S. X., Gbadebo, A. O., and Jesse, S. B. (2024). "Prevalence of Gastrointestinal Parasites of Poultry Chicken Slaughtered and Sold in Uyo Metropolis, Akwa Ibom State, Nigeria". *International Journal of Pathogen Research* 13 (5):23-31. <u>https://doi.org/10.9734/ijpr/2024/v13i5306</u>.
- Rehman, T. U., Khan, M. N., Sajid, M. S., Abbas, R. Z., Arshad, M., Iqbal, Z. and Iqbal, A. (2010). Epidemiology of Eimeria and associated risk factors in cattle of district Toba Tek Singh, Pakistan. *Parasitol. Res.*, 108:1171-1177.
- Sharma, S., Iqbal, A., Azmi, S., Mushtag, I., Wani, A. Z. and Ahmad, S. (2015) Prevalence of poultry coccidiosis in Jammu region of Jammu and Kashmir State. J. Parasit.Dis., 39(1): 85-89.
- Sheikh, B. A., Ahmad, F. and Sofi, T. A. (2016). Morphology and prevalence of some helminth parasites in *Gallus domesticus* from Gurez valley of Jammu and Kashmir, India. *Journal of Fisheries and Livestock Production* 4:151-159.
- Silas, I. I., Essien, E. A., Anietiem H. N., Inyang, S. X. and Akpan, A. E. (2024). Comparative study on the prevalence of gastrointestinal parasite between wild and pond raised African Sharptooth Catfish *Clarias gariepinus*in Akwa Ibom State. *South Asian Journalof Parasitology* (SAJP), 7(2):133-140.
- Tarwireyi, L. and Fanadzo, M. (2013) Production of indigenous chickens for household food security in rural KwaZulu-Natal, South Africa: A situation analysis. *African Journal of Agricultural Research*. 8:5832–5840.
- Taylor, M. A., Coop, R. L. and Wall, R. L. (2007) Parasites of poultry and game birds. In: Anderson, J.M. and Macfadyen, A., editors. Veterinary Parasitology. Blackwell Publishing, Iowa State, USA. p459-557.

- UN DESA (United Nations Department of Economic and Social Affairs (2017) World Population Prospects: the 2017 Revision. Retrieved via https://www.un.org/development/desa/en/news/population/worldpopulation-
- Usip, L. P. E., Okoro, M. F. and Udo, K. S. (2017). Prevalence of Intestinal Parasite of Poultry Chicken in Uyo Urban, Akwa Ibom State, Nigeria. World Journal of Applied Science and Technology.9(2)165 -172.
- Usman A. M., Malann Y.D. and Babeker E. A. (2022). Prevalence of Coccidiosis among Local and Exotic Breeds of Reared Chickens in Azare Metropolis, Bauchi State Nigeria. *Dutse Journal of Pure and Applied Sciences (DUJOPAS)*, 8 (3b): 109-114.
- Zalizar, L., Winaya, A., Malik, A., Widodo, W., and Anggraini, A. D. (2021). Species identification and prevalence of gastrointestinal helminths in Indonesian native chickens, and its impact on egg production. *Biodiversitas*. 22(10):4363-4369.