Review Article

Gut hormones in glucose homeostasis and current treatment approach in the control of T2DM - A succinct review

ABSTRACT

Gut-brain axis plays a key role in the regulation of energy homeostasis and glucose metabolism through various hormones. Gut hormones are peptides synthesized by specialized cells of enteroendocrine located in the epithelium of the stomach, small bowel and large bowel. Gut hormones activate neural circuits to signal peripheral organs for overall energy intake and assimilation. Incretins, Leptins, CCK, coordination of Oxyntomodulin, PYY and Gastrin are the major gut hormones involved in glucose metabolism. Group of gut peptides that are secreted after nutrient intake and stimulate insulin secretion together with hyperglycaemia are known as incretin hormones. Certain gut hormones like cholecystokinin (CCK) and gastrin are reported to activate pathways that promote islet neogenesis and improve glucose homeostasis in type 2 diabetes mellitus (T2DM). First-generation gut hormone treatments-GLP-1 analogues (incretin mimetics) are now available for the treatment of T2DM.Presently, hormonal synergy is of therapeutic interest for treatment of diabetes mellitus. Augmenting the biological activity of the "incretin" hormones to address many of the pathophysiological problems of diabetes is an effort in this direction. Gut hormones such as OXM, ghrelin and PYY play crucial role in the regulation of glucose. Pleirotropic actions of leptin reported to lower glucose is also, an area of investigation for hyperglycemia. Research has proved that these hormonal actions are a possible platform for therapeutic development in T2DM management.

Keywords: Gut hormones, glucosehomeostasis, T2DM, GLP-1 receptor agonists, DPP-4 inhibitors

1.INTRODUCTION

The diverse actions of gut peptides play an important role regulating the control of various physiological actions like satiety, gut motility, digestion and absorption of nutrient, disposal and energy storage. Gut hormones play role to initiate several physiological processes in multiple metabolically active tissues hence, attracted as therapeutic targets in the treatment of type 2 diabetes mellitus. In 1902 Bayliss and Starling described the first gastrointestinal (GI) hormone, secretin, establishing the role of the GI tract as an endocrine organ [1]. Considerable evidence is available on the important biological role of these endogenous hormones with direct bearing on glucose homeostasis.Incretins, Leptins, CCK, Oxyntomodulin, PYY and Gastrin are the gut hormones responsible for glucosehomeostasis.GIP and GLP-1 two major incretins along with gastrin, secretin, and

cholecystokinin play a key role in the pathophysiology of type 2 diabetes. Failure of pancreatic β-cell functioning cause insulin depletion as well as insulin resistance in organs is a pathophysiological disorder in Type 2 diabetes (T2DM). Impaired regulation of incretin hormones which reduce BG levels is another fundamental defect in the pathogenesis of Type 2 diabetes [2-4]. Leptin primarily produced in the adipose tissue although does not increase insulin levels, can potently increase insulin sensitivity [5-7] and participates in regulation of glucose absorption. CCK released from intra-islet neurons [8] along with GLP-1 (glucagon-like peptide-1) enhances insulin secretion. Oxyntomodulin (OXM) anther peptide secreted post-prandially is a dual agonist of the GLP-1 receptor and the glucagon receptor combining the effects of both hormones.PYY3-36 from PYY1-36 a satiety hormone processed by DPP-4 may also regulate glucose homeostasis by improving insulin sensitivity [9].Gastrin may contribute to incretin effect in combination with other hormones. Gastrin peptides are reported to stimulate insulin secretion independent of glucose [8,10]. These hormonal actions are now being viewed as possible platform for the apeutic development in T2DM management [11,12]. Incretin-based therapy has clearly emerged as one of the most sought out strategy in managing type 2 DM [13]. The paper is a concise review on gut hormones in glucose metabolism and the current therapeutic development to reduce hyperglycemic condition in T2DM subjects.

2. INCRETINS IN GLUCOSE METABOLISM

The term "incretin" was coined in 1932 to describe hormones that stimulated upper gut mucosa i.e., islet secretions of the pancreas[14,15]. Oral glucose elicits a higher insulin response than intravenous glucose at identical plasma glucose (PG) profiles (isoglycemia) is termed as incretin effect [16,17].Glucose-dependent insulinotropic polypeptide (GIP) and glucagon-like peptide-1(GLP-1) are the two major incretin hormones from the upper (GIP, 'K' cells) and lower (GLP-1, 'L' cells) gut [18]. The secretion of these incretins vary with individuals with same trend of secretion in both the hormones [19,20]. Thus, incretins are the gut hormones that potentiate insulin secretion after meal ingestion in a glucose-dependent manner. While, GIP and GLP-1 are major incretins, gastrin and cholecystokinin may also have minor roles to play in the pathophysiology of type 2 diabetes. GLP-1 and GIP enhance the effects of insulin, suppress glucagon release, and decrease hepatic gluconeogenesis to maintain BG levels in healthy subjects [21]. Reduced incretin effect is a consequence of the diabetic state and not a primary event in the development of type 2 diabetes [4,20,22]. Incretin effect is also, reported to be reduced in type 1 diabetes subjects and normal fasting glucose levels [2,23-25]. Glucagon producing α-cells, play a key role in glucose counter-regulation to avoid dangerous hypoglycemia. Glucagon-like peptide-1 (GLP-1) issecreted from the 'L'-cells located in the gut epithelium [26] with enteroendocrine cells distributed throughout the jejunum, ileum and colon [27].GLP1 circulates in two equipotent forms as GLP1₇₋₃₇ and GLP1₇₋₃₆ amide [28,29,30], but most circulating GLP1 in humans is GLP1₇₋₃₆ amide [31]. GLP-1enhances the differentiation of new *B*-cells from progenitor cells in the pancreatic duct epithelium [32] also, stimuling cell proliferation [33-35].GLP-1 is capable of inhibiting apoptosis of B-cells [36] to maintain a balance between apoptosis and proliferation [37].Glucose-dependent insulinotropic polypeptide (GIP) cells are found in the small intestinal mucosa [38] secreted from specific endocrine cells, known as 'K' cells in response to glucose, amino acids, and lipids [39,40]. GIP is a 42 amino acid peptide stimulating insulin together with GLP-1.Late phase of the insulinotropic response is particularly impaired in type 2 diabetes[22]. However, insulin resistance is independent of decreased GLP-1 [3,41]. GIP contains an alanine at position 2 and is a substrate for enzymatic inactivation by DPP4, an aminopeptidase. Insulinotropic actions of GIP are more prominent during hyperglycaemia [42].GIP effect was preserved in women who had a history of gestational diabetes and are therefore at high risk of developing type 2 diabetes[43]. The enteroglucagon peptides expressed by proglucagon gene, primarily in the 'L' cells of the distal intestine are glicentin and oxyntomodulin (OXM).

Table 1: Gut hormones in glucose homeostasis

Harmone	Site of secretion	Mode of action	Reference
GLP-1	Secreted from the L-cells located in the gut epithelium	Augmentation of insulin, inhibition of glucagon secretion, inhibits gastric emptying,food intake, and maximizing nutrient absorption	[144]
GIP	Secreted in intestinal mucosa from endocrine cells, called K cells	Mediates the postprandial potentiation of insulin secretion	[38,39]
Gherlin	Secreted primarily in the enteroendocrine cells as pro-hormone by P/D1 closed-type cells in gastric fundus.	Growth hormone secretagogue that stimulates pituitary release of growth hormone and stimulates hypothalamic centers to increase appetite Effects mediated through vagus nerve	[69]
Leptin	A peptide hormone containing 167 amino acids primarily produced in the adipose tissue	Regulates absorption of glucose	[94]
Gastrin	The main site of production gastrin in adults is the antroduodenal G-cells	Contributes to an incretin effect in combination with other hormones	[54,59]
PYY	Peptide YY is a short (36-amino acid) peptide released from cells in the ileum and colon in response to feeding.	Increases postprandial insulin and glucose response and also regulate glucose homeostasis through peripheral effects distinct from its interaction with islets	[107]
Cholecystokinin (CCK)	I-cells in duodenal mucosa, particularly with multiple molecular forms	Moderatespostprandial glycemia by slowing down gastric emptying. CCK along with incretin hormone GLP-1 enhances insulin secretion.	[142]
Oxyntomodulin	Oxyntomodulin (OXM) is a peptide hormone released from the gut in postprandial state.	Decreases food intake by suppressing appetite, enhances glucose stimulated insulin secretion and inhibits glucagon release, thereby reducing postprandial glucose levels	[143]

Glicentin is considered as only a discarded metabolite of proglucagon after the cleavage of GLP-1 and GLP-2 [44].OXM similar to GLP-1, a peptide of 37-amino acid secreted from the intestine following nutrient ingestion originating from the same proglucagon precursor [45]. OXM is a dual agonist of the GLP-1 receptor and the glucagon receptor combining the effects of both hormones. Like GLP-1,OXM decreases food intake by suppressing appetite,inhibits gastric emptying, enhances glucose-stimulated insulin secretion and inhibits

glucagon release, thereby reducing postprandial glucose levels by glucagon action thus a key peptide in therapeutic development [46-49]. Activated glucagon receptor (GCGR) can increase hepatic glucose production, but the overall metabolic effect of OXM is balanced toward improving glycemic control.

3. OTHER GUT HORMONES IN GLUCOSE METABOLISM

Cholecystokinin (CCK) peptides are released from intra-islet neurons [8] with two receptors, CCKAR (CCK1R) and CCKBR (CCK2R), CCKBR mediating the effects of CCK on the control of glucose homeostasis by the pancreas. CCK moderatespostprandial glycemia by slowing down gastric emptying which otherwise leads to poor glycemic control. CCK along with incretin hormone GLP-1 enhances insulin secretion.CCK has been shown to stimulate glucagon release from human islets in vitro. In vitro studies elucidate glucagon release by CCK from islets, and stimulation of insulin in a glucose-dependant manner in mice model. Infusion of CCK-8 increases plasma insulin concentration and reduces glucose excursion following meal ingestion in normal and T2DM subjects [50]. CCK has proliferative role on pancreatic β cells while CCK-8 can promote regeneration of β cells [51]. Short CCK peptides, CCK-8, CCK-5, and CCK-4 have been shown to release insulin in humans and in the isolated perfused porcine pancreas [8,52,53]. Gastrin as a humoral mediator of gastric acid secretion proposed in 1905 [54]. However, physiological proof of an acid-stimulating hormone from gastric antrum was presented in 1948 [55] and later isolation, structure and physiological functions were determined [56]. Gastrin may contribute an incretin effect in combination with other hormones as evident in mouse model under gastrin and GLP-1 dual agonist ZP3022 [57]. Gastrin when co-administrated with glucose more pronounced insulin release was evidenced supporting incretin effect [10]. The main site of production gastrin in adults is the antroduodenal G-cells targeting G-protein coupled receptors [58.59]. Human islet cells are well equipped with gastrin receptors [60,61].Gastrin is likely to induce β-cell proliferation, neogenesis and stimulate the secretion of insulin postprandially [62].Gastrin peptides are reported to stimulate insulin secretion independent of glucose [8,10]. Gastrin enhances islet mass from transdifferentiated exocrine pancreatic tissue [63]and induces the expression of glucagon genes in α-cells [64].Gastrin is expressed in fetal and neonatal pancreatic islets [65]

Gherlin is a 28-amino acid hormone is produced in the fasting state promoting hunger sensations [66-68].Gherlin is a endogenous ligand for the growth hormone secretagogue receptor (GHSR)1a, capable of stimulating growth hormone (GH) release from the anterior pituitary gland [69]. Secreted primarily in the enteroendocrine cells [69-71] as pro-hormone by P/D1 closed-type cells in gastric fundus. Gherlinto act on its own receptors. the growth hormone secretagogue receptor (GHSR 1a) must be cleaved and posttranscriptionally acylated by the enzyme ghrelin O acyltransferase (GOAT) a member of the membrane bound O acyltransferase (MBOAT) family [69,72-76].GHSR1a expressed by acells of the pancreatic islet are likely to contribute to the ability of GH to directly stimulate glucagon secretion [77]. Acylated bioactive ghrelin (AG) produced in ε cell of pancreatic islets [78],acts on β -cells of the *islets* promoting calcium release (Ca2+) as a messenger signal [79].Ghrelin inhibition of insulin secretion is reported in most animal studies [80,81].Blocking the function of endogenous ghrelin with GHSR1a showed low fasting glucose concentrations suggesting an inhibitory role for ghrelin in the control of insulin secretion [82].Ghrelin secretion reported to be on peak during fasting [83]. Plasma ghrelin and insulin levels are negatively correlated [84,85], with an inverse relationship between circulating ghrelin levels and insulin resistance [86]. While,AG is responsible for the decrease in insulin with a consequent rise in glucose levels [87] with UAG able to antagonize the effects of AG on insulin secretion [88,89]. Investigations reveled that ghrelin administration increased plasma levels of glucose and decreased plasma levels of insulin [90,91] with plasma concentration of glucose regulating ghrelin secretion from a-cells to stimulates insulin secretion [92].

In 1994, the human obese (OB) gene located on chromosome 7 and its product leptin were identified and characterized [93,94]. Leptin is a peptide hormone containing 167 amino acids primarily produced in the adipose tissue and in in small amounts in tissues of the stomach, mammary epithelium, placenta and heart [95]. Direct role of Leptin on glucose metabolism independent of body weight and food intake is demonstrated in leptin deficient mice [96]. Similarly, in vitro studies have shown mechanism and regulatory role of leptin in glucose absorption [97-99]. Although leptin does not increase insulin levels, it can potently increase insulin sensitivity as seen in animal models of T1DM [100-102]. The glucose lowering actions of leptin are largely facilitated through its role in many metabolic pathways due to its pleirotropic actions [103]. An increase in adipocyte leptin expression and circulating leptin is reported after overfeeding in healthy humans [104], with circulating leptin levels showing a diurnal pattern influenced by gender, age, exercise and glucose uptake [105]. Shanta and Gavin (2014) reviewed the potential role of peptide tyrosine tyrosine (PYY) in Glucose homeostasis [106]. Gut hormone Peptide YY (PYY) with 36 amino acids was first isolated from porcine intestine [107] and the biological activity is dependent on the presence of an amide group at the C-terminus. PYY a satiety hormone released from the enteroendocrine L cells. PYY increased postprandial insulin and glucose responses [108].PYY may also regulate glucose homeostasis through peripheral effects distinct from its interaction with islets [109]. In addition, PYY3-36 from PYY1-36 processed by DPP-4 may also regulate glucose homeostasis by improving insulin sensitivity [9].

4. GUT HORMONES AND T2DM CONTROL STRATEGIES

Synergistic effect of gut hormone combinations for glucose metabolism is seen as better alternative. Combination effects of GLP-1 and GIP with CCK and gastrin peptides are of clinical interest now for glucose metabolism [110-112]. GLP-1 due to rapid degradation by dipeptidyl peptidase-4 (DPP-4), which has very short half-life of >2 min in plasmais is a major limitation [113,114].DPP-4 resistant GLP-1 receptor agonists (incretin mimetics), and inhibitors of DPP-4 activity (incretin enhancers) are being successfully used clinically for treatment of T2diabetes mellitus currently. GLP-1 receptor agonists proved to be weightnegative anti-diabetes treatment option. Exenatide is a synthetic form of a natural peptide found in the saliva of Gila monster-Heloderma suspectum first analogue GLP-1 receptor agonists [115]. Liraglutide, Dulaglutide, and Semaglutide are other GLP-1 receptor agonists.DPP-4 inhibitors stimulate insulin secretion and inhibit glucagon secretion by elevating endogenous GLP-1 concentrations without an intrinsic hypoglycaemia risk. DPP-4 inhibitors raise only the proportion of active GLP-1 postprandial concentration [116], resulting in elevated plasma levels of GLP-1 does not produce GLP-1-related side effects [117,118]. DPP-IV inhibitor are small-molecules (gliptins) also demonstrated to be effective in antihyperglycemic state devoid of any major adverse events. Presently there are five DPP-4 inhibitors available vie., sitagliptin (2006), vildagliptin (2007), saxagliptin (2009), linagliptin (2011) and alogliptin (2013). Four more gliptins, namely teneligliptin, anagliptin, omarigliptin, and trelagliptin are approved in the Japanese and Korean markets. Generally, the DPP-4 inhibitors are eliminated primarily via the kidney[119-123], except linagliptin which is eliminated via the biliary pathway [124,125]. Inhibition of ghrelin can be a potential therapeutic target to regulate hyperglycemia opening a new avenue for type-2 diabetes subjects. The ghrelin receptor, growth hormone secretagogue receptor (GHSR1a) is expressed in a wide variety of tissues suggesting diverse biological activity. GHSR1a antagonism could be a promising therapy in the treatment of T2DM. Inhibition of posttranscriptional octanoylation by the enzyme ghrelin O acyltransferase (GOAT) can be a target to get improved glycemic control [126,127].LEAP2,Quinazolinone and Triazole are the presently known antagonists of GHSR1a.

Proton pump inhibitors (PPIs) are a group of medicines that decrease stomach acid production and can raise serum gastrin concentration significantly to affect glucose metabolism through promoting β-cell regeneration/expansion and also enhancing insulin

secretion [128].PPI lansoprazole increased serum gastrin which is associated with improved glycemia and increased pancreatic insulin content in rat models [129]. Gastrin with GLP-1 dual agonist showed incretin effect in animal models can be a area to be investigated [130,131]. The duodenal-jejunal bypass liner (DJBL; EndoBarrier; GI Dynamics, MA, USA) is a 60-cm-long impermeable sleeve-like device, suggests potential hormonal mechanisms for diabetes improvement needs further confirmations [132,133]. Although insulin therapy restores circulating leptin levels in type 1 diabetic patients [134], addition of leptin provides more glycemic control, with less-frequent insulin dosing. However, leptin and insulin cotherapy has a potential danger of hypoglycemia [134,135].PYY also, represents as a therapeutic tool after establishment of its role as anti-obesity and anti-diabetic effects. PYY is a key effector of the early recovery of impaired glucose-mediated insulin and glucagon secretion in bariatric surgery establishes principles in development of new non-surgical therapy for T2D correction [136].GLP-1, PYY, and oxyntomodulin combination therapy was reported to improved post-prandial glycemic control similar to that of RYGB patients[137]. However, use of PYY as a potential treatment needs further investigation (Batterhamet al., 2003). Infusion of oxyntomodulin a promising glucose-lowering and obese and therapy well-tolerated in human studies is also, reported but for its its short circulating half-life [138-141].

CONCLUSIONS

Gut-brain axis has a key role in the regulation of energy homeostasis and glucose metabolism. A better understanding of the gut-brain axis perhaps may the key for the development of successful therapies to manage diabetes and related metabolic disorders. Caution must be taken to avoid side effects when developing therapies, as gut hormones play role not only in glucose homeostasis but act on other physiological actions. Similarly, more attention is required towards comorbidity linked to diabetes. Bariatric surgery has given a new thinking on exploiting hormonal changes to target future medical therapies for Type 2 diabetes mellitus. It may be possible to reset metabolism and reverse diabetes taking the advantage of knowledge gained from bariatric surgery.

CONSENT (WHERE EVER APPLICABLE): Not applicable

ETHICAL APPROVAL (WHERE EVER APPLICABLE): Not applicable

REERENCES

- 1. Bayliss, W.M., & Starling, E.H. (1902). The mechanism of pancreatic secretion. J Physiol., 28:325–53.
- 2. Nauck, M., Stöckmann, F., Ebert R., et al. (1986). Reduced incretin effect in Type 2 (non-insulin-dependent)-diabetes. Diabetologia, 29,46–52. https://doi.org/10.1007/BF02427280
- 3. Muscelli, E., Mari, A., Casolaro, A., Camastra, S., Seghieri, G., Gastaldelli, A., et al. (2008). Separate impact of obesity and glucose tolerance on the incretin effect in normal subjects and type 2 diabetic patients. Diabetes, 57(5):1340-8.
- 4. Knop, F.K., Tina, Vilsboll., Patricia, V., Hojberg, Steen Larsen, Sten Madsbad Aage Volund, Holst, J.J., Thure, Krarup. (200).Reduced Incretin Effect in Type 2 Diabetes Cause or Consequence of the Diabetic State? Filip K. Diabetes, 56: 1951–1959.
- 5. Lin, C.Y., Higginbotham, D.A., Judd, R.L., White, B.D. (2002). Central leptin increases insulin sensitivity in streptozotocin-induced diabetic rats American Journal of Physiology-Endocrinology and Metabolism. 282:E1084-E1091.
- 6. German, A.J., Holden, S.L. Bissot, T., Morris, P.J., Biourge, V. (2010). A high protein high fibre diet improves weight loss in obese dogs. Vet J.,183(3):294-7. doi: 10.1016/j.tvjl.2008.12.004. Epub 2009 Jan 12. PMID: 19138868.

- 7. Denroche, H.C., Huynh, F.K., Kieffer, T.J. (2012). The role of leptin in glucose homeostasis. J Diabetes Investig., 3(2):115-29. doi: 10.1111/j.2040-1124.2012.00203.x. PMID: 24843554; PMCID: PMC4020728.
- 8. Rehfeld, J.F., Larsson, L.I., Goltermann, N.R., *et al.* (1980). Neural regulation of pancreatic hormone secretion by the C-terminal tetrapeptide of CCK. *Nature*,284:33–38.
- 9. van den Hoek, A.M., Heijboer, A.C., Corssmit, E.P., et al. (2004). PYY3-36 reinforces insulin action on glucose disposal in mice fed a high-fat diet. Diabetes.53:1949–1952.
- 10. Rehfeld, J.F., Stadil, F. (1973). The effect of gastrin on basal- and glucose stimulated insulin secretion in man. J Clin Invest.52:1415–1426.
- 11. Drucker, D.J. (2015).Deciphering metabolic messages from the gut drives therapeutic innovation: the 2014 Banting Lecture. *Diabetes*.64(2):317-326.
- Drucker, D.J. (2024). Prevention of cardiorenal complications in people with type 2 diabetes and obesity. Cell Metabolism. 36:338-353
- 13. Phung, O.J., Scholle, J.M., Talwar, M., Coleman, C.I. (2010). Effect of non insulin antidiabetic drugs added to metformin therapy on glycemic control, weight gain, and hypoglycemia in type 2 diabetes. J. Am. Med. Assoc.303 (14):1410–1418. doi:10.1001/jama.2010.405
- 14. La Barre, J. (1932). Sur les possibilités d'un traitement du diabète par l'incrétine. Bull Acad Royal Med Belg.12:620–634.
- 15. Rehfeld, J. (2018). The origin and understanding of the incretin concept. Front Endocrinol (Lausanne). 9:387
- 16. McIntyre, N., Holdsworth, C.D., Turner, D.S. (1964). New interpretation of oral glucose tolerance. Lancet.2:20-21.
- 17. Elrick, H., Stimmler, L., Hlad, C.J. Jr., Arai, Y. (1964). Plasma insulin response to oral and intravenous glucose administration. J. Clin. Endocrinol. Metab., 24:1076–1082.
- 18. Holst, J.J., & Gromada, J. (2004). Role of Incretin Hormones in the Regulation of Insulin Secretion in Diabetic and Non-diabetic Humans. American Journal of Physiology: Endocrinology and Metabolism.287.E199-E206. http://dx.doi.org/10.1152/ajpendo.00545.
- 19. Calanna, S., Christensen, M., Holst, J.J. et al. (2013). Secretion of glucose-dependent insulinotropic polypeptide in patients with type 2 diabetes: systematic review and meta-analysis of clinical studies. Diabetes Care.36:3346–3352. https://doi.org/10.2337/ dc13-0465
- 20. Nauck, M.A., El-Ouaghlidi, A., Gabrys, B., et al. (2004). Secretion of incretin hormones (GIP and GLP-1) and incretin efect after oral glucose in frst-degree relatives of patients with type 2 diabetes. Regul Pept.122:209–217. https://doi.org/10.1016/j.regpep.2004.06.020
- 21. Asmar, M., Simonsen, L., Madsbad, S., Stallknecht, B., Holst, J.J., Bulow, J. (2010). Glucose-dependent insulinotropic polypeptide may enhance fatty acid re-esterification in subcutaneous abdominal adipose tissue in lean humans. Diabetes. 2010;59(9):2160-3. doi: 10.2337/db10-0098. Epub 2010 Jun 14. PMID: 20547981; PMCID: PMC2927937.
- 22. Vilsboll, T., Krarup, T., Madsbad, S., Holst, J.J. (2002). Defective amplification of the late phase insulin response to glucose by GIP in obese Type II diabetic patients. Diabetologia.45(8):1111-9. doi: 10.1007/s00125-002-0878-6. Epub 2002 Jul 4. PMID: 12189441.
- 23. Greenbaum, C.J., Prigeon, R.L., D'Alessio, D.A. (2002).Impaired beta-cell function, incretin effect, and glucagon suppression in patients with type 1 diabetes who have normal fasting glucose. Diabetes.51(4):951-7. doi: 10.2337/diabetes.51.4.951. PMID: 11916912.
- 24. Bagger, J.I., Knop, F.K., Lund, A., Vestergaard, H., Holst, J.J., Vilsboll, T..(2011). Impaired regulation of the incretin effect in patients with type 2 diabetes. J Clin Endocrinol Metab.96(3):737-45. doi: 10.1210/jc.2010-2435. Epub 2011 Jan 20. PMID: 21252240.
- 25. Meier, J.J., Nauck, M.A. (2006).Incretins and the development of type 2 diabetes. Current Diabetes *Reports*,6(3): 194-201.
- 26. Solcia, E., Capella, C., Buffa, R., Fiocca, R., Frigerio, B., Usellini, L. (1980). Identification, ultrastructure and classification of gut endocrine cells and related growths. Investig. Cell Pathol.,3: 37–49.
- 27. Hansen, C.F., Vrang, N., Torp Sangild, P., Jelsing, J. (2013). Novel insight into the distribution of L-cells in the rat intestinal tract. Am. J. Transl. Res.5: 347–358.
- 28. Mojsov, .S., Kopczynski, M.G., Habener, J.F. (1990). Both amidated and nonamidated forms of glucagon-like peptide I are synthesized in the rat intestine and the pancreas. J. Biol. Chem., 265:8001–8008.
- 29. Orskov, C., Wettergren, A., Holst, J.J. (1993). Biological effects and metabolic rates of glucagonlike peptide-1 7-36 amide and glucagonlike peptide-1 7-37 in healthy subjects are indistinguishable. Diabetes.42:658–661. doi: 10.2337/diab.42.5.658.
- 30. Wettergren, A., Pridal, L., Wojdemann, M., Holst, J.J. (1998). Amidated and non-amidated glucagon-like peptide-1 (GLP-1): non-pancreatic effects (cephalic phase acid secretion) and stability in plasma in humans. Regul. Pept.,77:83–87. doi: 10.1016/s0167-0115(98)00044-5.
- 31. Orskov, C., Rabenhoj, L., Wettergren, A., Kofod, H., Holst, J.J. (1994). Tissue and plasma concentrations of amidated and glycine-extended glucagon-like peptide I in humans. Diabetes.43:535–539. doi: 10.2337/diab.43.4.535.

- 32. Zhou, J., Wang, X., Pineyro, M.A., Egan, J.M.(1990). Glucagon-like peptide 1 and exendin-4 convert pancrteatic AR42J cells into glucagon- and insulin producing cells. Diabetes.48:2358–2366.
- 33. Xu, G., Stofera, D.A., Habener, J.F., Bonner-weir, S. (1999). Exendin-4 stimulates both beta-cell replication and neogenesis, resulting in increased beta-cell mass and improved glucose tolerance in diabetic rats.. Diabetes.48 (12):2270–2276. https://doi.org/10.2337/diabetes.48.12.2270
- 34. Butler Alexandra, E., Martha Campbell-Thompson, Tatyana Gurlo, David W Dawson, Mark Atkinson, Peter C. Butler. (2013). Marked expansion of exocrine and endocrine pancreas with incretin therapy in humans with increased exocrine pancreas dysplasia and the potential for glucagon-producing neuroendocrine tumors. Diabetes. 2:62:2595-2604
- 35. Bai, L., Meredith, G., Tuch, B.E. (2005). Glucagon-like peptide-1 enhances production of insulin in insulin-producing cells derived from mouse embryonic stem cells. J Endocrinol.,186(2):343-52. doi: 10.1677/joe.1.06078. PMID: 16079260.
- 36. Farilla, L., Bulotta, A., Hirshberg, B., Li Calzi, S., Khoury, N., Noushmehr, H., Bertolotto, C., Di Mario, U., Harlan, D.M., Perfetti, R. (2003). Glucagon-like peptide 1 inhibits cell apoptosis and improves glucose responsiveness of freshly isolated human islets. Endocrinology,144(12):5149-58. doi: 10.1210/en.2003-0323. Epub 2003 Aug 28. PMID: 12960095.
- 37. Bonner-Weir. Susan. (2000). Life and Death of the Pancreatic β Cells.Trends in Endocrinology & Metabolism,11(9):375 378.
- 38. Mortensen, K., Christensen, L.L., Holst, J.J., Cathrine Orskov. (2003). GLP-1 and GIP are colocalized in a subset of endocrine cells in the small intestine. Regulatory Peptides, 14(2–3):189-196.
- 39. Falko James, M., Crockett Samuele, E., Samuel Cataland, et al. (1975). Gastric Inhibitory Polypeptide (GIP) Stimulated by Fat Ingestion in Man, *The Journal of Clinical Endocrinology & Metabolism.* 41(2):260–265,https://doi.org/10.1210/jcem-41-2-260
- 40. Buchan Alison, M.J., Polak Julia, M., Capella, C., Solcia, E., Pearse, A.G.E. (1978). Electronimmunocytochemical Evidence for the K Cell Localization of Gastric Inhibitory Polypeptide (GIP) in Man. Histochemistry, 56:37-44.
- 41. Rask Eva, Tommy Olsson, Stefan Söderberg, Owe Johnson, Jonathan Seckl, Jens Juul Holst, Bo Ahrén. (2001). Impaired Incretin Response After a Mixed Meal Is Associated With Insulin Resistance in Nondiabetic Men. *Diabetes Care*,24 (9):1640–1645. https://doi.org/10.2337/diacare.24.9.1640
- 42. Christensen, M., Vedtofte, L., Holst, J.J., Vilsboll, T., et al. (2011). Glucose-dependent insulinotropic polypeptide: a bifunctional glucosedependent regulator of glucagon and insulin secretion in humans. Diabetes,60:3103–3109. https://doi.org/10.2337/db11-0979
- 43. Meier, J.J., Gallwitz, B., Askenas, M., Vollmer, K., Deacon, C.F., Holst, J.J., et al. (2005). Secretion of incretin hormones and the insulinotropic effect of gastric inhibitory polypeptide in women with a history of gestational diabetes. *Diabetologia*,48:1872-1881.
- 44. Sinclair, E.M., Drucker, D.J., (2005).Proglucagon-derived peptides: mechanisms of action and therapeutic potential. Physiology (Bethesda),20:357–365.
- 45. Cohen, M.A., Ellis, S.M., Le Roux, C.W., *et al.* (2003).Oxyntomodulin suppresses appetite and reduces food intake in humans. J. Clin. Endocrinol. Metabol.,88(10):4696–4701.
- 46. Collie, N.L., Zhu, Z,,Jordan, S., Reeve, J.R. (1997). Oxyntomodulin stimulates intestinal glucose uptake in rats. Gastroenterology,112:1961–1970.

 doi: 10.1053/gast.1997.v112.pm9178688.
- 47. Jarrousse, C., Bataille, D., Jeanrenaud, B. (1984). A pure enteroglucagon, oxyntomodulin (glucagon 37), stimulates insulin release in perfused rat pancreas. Endocrinology, 115(1): 102-105.
- 48. Baggio, L.L., Huang, Q., Brown, T.J., Drucker, D.J. (2004). Oxyntomodulin and glucagon-like peptide-1differentially regulate murine food intake and energy expenditure. Gastroenterology, 127(2):546–558.
- 49. Pocai, A. (2012). Unraveling oxyntomodulin, GLP1's enigmatic brother. J. Endocrinol., 215(3):335–346.
- 50. Ahren, B., Holst, J.J. (2000). Efendic S. Anti-diabetogenic action of cholecystokinin-8 in type 2 diabetes. J. Clin. Endocrinol. Metab, .85:1043–1048.
- 51. Kuntz, E., Pinget, M., Damge, P. (2004). Cholecystokinin octapeptide: a potential growth factor for pancreatic beta cellsin diabetic rats. *Jop.***5**:464–475.
- 52. Ohgawara, H., Mizuno, Y., Tasaka, Y., Kosaka, K. (1969). Effect of the C-terminal tetrapeptide amide of gastrin on insulin secretion in man. J Clin Endocrinol Metab., 29:1261-1262.
- 53. Kaneto, A., Tasaka, Y., Kosaka, K., Nakao, K. (1969). Stimulation of insulin secretion by the C-terminal tetrapeptide amide of gastrin. Endocrinology,84: 1098-1106.
- 54. Edkins, J.S. (1905). On the chemical mechanism of gastric secretion. Proc Roy Soc Ser B.76:376–376.
- 55. Grossman, M.I., Robertson, C.R., Ivy, A.C.(1948). The proof of a hormonal mechanism for gastric secretion the humoral transmission of the distension stimulus. Am J Physiol., 153:1–9.
- 56. Gregory, R.A. (1974). The Bayliss-Starling lecture 1973. The gastrointestinal hormones: a review of recent advances. J Physiol.,241, 1–32.

- 57. Fosgerau, K., Jessen, L., Lind Tolborg, J., Osterlund, T., Schaeffer Larsen, K., Rolsted, K., Brorson, M., Jelsing, J., et al. (2013). The novel GLP-1-gastrin dual agonist, ZP3022, increases β-cell mass and prevents diabetes in db/db mice. Diabetes Obes Metab.,15(1):62-71. doi: 10.1111/j.1463-1326.2012.01676.x.Epub 2012 Sep 9. PMID: 22862961.
- 58. Kopin, A.S., Lee, Y.M., McBride, E.W., et al. (1992). Expression cloning and characterization of the canine parietal cell gastrin receptor. Proc Natl Acad Sci U S A, 89:3605-3609.
- 59. Wank, S.A., Harkins, R., Jensen, R.T., Shapira, H., de Weerth, A., Slattery, T.(1992). Purification, molecular cloning, and functional expression of the cholecystokinin receptor from rat pancreas. Proc Natl Acad Sci U S A, 89:3125-3129.
- 60. Dufresne, M., Seva, C., Fourmy, D. (2006). Cholecystokinin and gastrin receptors. Physiol. Rev., 86:805–847.
- 61. Reubi, J.C., Waser, B., Gugger, M., et al. (2003). Distribution of CCK1 and CCK2 receptors in normal and diseased human pancreatic tissue. Gastroenterology.,125:98–106.
- 62. Rehfeld, J.F..(1976). Disturbed islet-cell function related to endogenous gastrin release. Studies on insulin secretion and glucose tolerance in pernicious anemia. J Clin Invest., 58:41-49.
- 63. Rooman, I., Lardon, J., Bouwens, L. (2002). Gastrin stimulates beta-cell neogenesis and increases islet mass from transdifferentiated but not from normal exocrine pancreas tissue. Diabetes,51: 686–690.
- 64. Leung-Theung-Long, S., Roulet, E., Clerc, P., Escrieut, C., Marchal-Victorion, S., Ritz-Laser, B., et al. (2005). Essential interaction of Egr-1 at an islet-specific response element for basal and gastrin-dependent glucagon gene transactivation in pancreatic alpha-cells. J Biol Chem., 280:7976–7984.
- 65. Larsson, L.I., Rehfeld, J.F., Sundler, F., Håkanson, R. (1976). Pancreatic gastrin in foetal and neonatal rats. Nature, 262:609–610.
- 66. Wren AM, Seal LJ, Cohen MA *et al.* (2001). Ghrelin enhances appetite and increases food intake in humans. J. Clin. Endocrinol. Metabol.,86(12), 5992.
- 67. Tschop, M., Wawarta, R., Riepl, R.L., Friedrich, S., Bidlingmaier, M., Landgraf, R., et al. (2001). Post-prandial decrease of circulating human ghrelin levels. Journal of Endocrinological Investigation, 24:RC19e21.
- 68. Ibrahim Abdalla, M.M. (2015). Ghrelin Physiological Functions and Regulation. Eur Endocrinol.11(2):90-95. doi: 10.17925/EE.2015.11.02.90. Epub 2015 Aug 19. PMID: 29632576; PMCID: PMC5819073.
- 69. Kojima, M., Hosoda, H., Date, Y., Date, M., Nakazato, H., Matsuo Kangaa, K. (1999). Ghrelin is a growth-hormone-releasing acylated peptide from stomach, Nature, 402(6762);656–660.
- 70. Date, Y., Kojima, M., Hosoda, H., Sawaguchi, A., Mondal, M.S., Suganuma, T., Matsukura, S., Kangawa, K., Nakazato, M. (2000). Ghrelin, a novel growth hormone releasing acylated peptide, is synthesized in a distinct endocrine cell type in the gastrointestinal tracts of rats and humans. Endocrinology.141:4255–4261.
- 71. Dornonville de la Cour, C., Bjorkqvist, M., Sandvik, A.K., Bakke, I., Zhao, C-M., Chen, D., Håkanson, R. (2001). A-like cells in the rat stomach contain ghrelin and do not operate under gastrin control. Regul Pept.,99:141–150.
- 72. Gutierrez, J.A., Solenberg, P.J., Perkins, D.R., Willency, J.A., Knierman, M.D., Jin, Z., et al. (2008). Ghrelin octanoylation mediated by an orphan lipid transferase. Proceedings of the National Academy of Sciences of the United States of America. 105:6320e6325.
- 73. Jing Yang., Michael, S. (2008). Brown, Guosheng Liang, Nick V. Grishin, Joseph L. Goldstein. Identification of the Acyltransferase that Octanoylates Ghrelin, an Appetite-Stimulating Peptide Hormone. Cell.132 (3): 387 396
- 74. Barnett, B.P., Hwang, Y., Taylor, M.S., Kirchner, H., Pfluger, P.T., Bernard, V., et al. (2010). Glucose and weight control in mice with a designed ghrelin O-acyltransferase inhibitor. Science,330(6011):1689-1692.
- 75. Kirchner, H., Gutierrez, J.A., Solenberg, P.J., Pfluger, P.T., Czyzyk, T.A., Willency, J.A., et al. (2009).GOAT links dietary lipids with the endocrine control of energy balance. Nature medicine, 15(7), 741-745.
- 76. Zhao, T.J., Lian, G., Li, RL., Xie,X., Sleeman, M.W., Murphy, A.J., et al. (2010). Ghrelin O-acyltransferase (GOAT) is essential for growth hormone-mediated survival of calorie-restricted mice. Proceedings of the National Academy of Sciences, 107(16):7467-7472.
- 77. Chuang, JC., Sakata, I., Kohno, D., Perello, M., Osborne-Lawrence, S., Repa, J.J., et al. (2011). Ghrelin directly stimulates glucagon secretion from pancreatic alpha-cells. Molecular Endocrinology,25:1600e1611.
- 78. Prado, C.L., Pugh-Bernard, A.E., Elghazi, L., SosaPineda, B., Sussel, L. (2004). Ghrelin cells replace insulin-producing beta cells in two mouse models of pancreas development. Proc. Natl. Acad. Sci. U. S. A.101:2924–2929.
- 79. Date, Y., Nakazato, M., Hashiguchi, S., Dezaki, K., Mondal, M.S., Hosoda, H., et al. (2002). Ghrelin is present in pancreatic alpha-cells of humans and rats and stimulates insulin secretion. Diabetes.51:124e129.
- 80. Reimer, M.K., Pacini, G., Ahren, B.. (2003). Dose-dependent inhibition by ghrelin of insulin secretion in the mouse. Endocrinology.144:916e921.
- 81. Qader, S.S., Lundquist, I., Ekelund, M., Hakanson, R., Salehi, A. (2005). Ghrelin activates neuronal constitutive nitric oxide synthase in pancreatic islet cells while inhibiting insulin release and stimulating glucagon release. Regulatory Peptides, 128:51e56.
- 82. Dezaki, K., Sone, H., Koizumi, M., Nakata, M., Kakei, M., Nagai, H., et al. (2006). Blockade of pancreatic islet-derived ghrelin enhances insulin secretion to prevent high-fat diet-induced glucose intolerance. Diabetes, 55:3486e 3493.

- 83. Dezaki, K., et al. (2004). Endogenous ghrelin in pancreatic islets restricts insulin release by attenuating Ca2+ signaling in beta-cells: implication in the glycemic control in rodents. Diabetes,53:3142–3151.
- 84. Cummings, D.E., Purnell, J.Q., Frayo, R.S., Schmidova, K., Wisse, B.E., Weigle, D.S. (2001). A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. Diabetes, 50:1714e1719.
- 85. Flanagan, D.E., Evans, M.L., Monsod, T.P., Rife, F., Heptulla, R.A., Tamborlane, W.V., et al. (2003). The influence of insulin on circulating ghrelin. American Journal of Physiology Endocrinology and Metabolism, 284:E313e E316.
- 86. Tschop, M., Wawarta, R., Riepl, R.L., Friedrich, S., Bidlingmaier, M., Landgraf, R., et al. (2001).Post-prandial decrease of circulating human ghrelin levels. Journal of Endocrinological Investigation, 24:RC19e21.
- 87. Gauna, C., Delhanty, P.J.D., Hofland, L.J., et al. (2005). Ghrelin stimulates, whereas des-octanoyl ghrelin inhibits, glucose output by primary hepatocytes, *J*ournal of Clinical Endocrinology and Metabolism, 90 (2):1055–1060.
- 88. Broglio, F., Gottero, C., Prodam, F., et al. (2004). Non-acylated ghrelin counteracts the metabolic but not the neuroendocrine response to acylated ghrelin in humans, *J*ournal of Clinical Endocrinology and Metabolism,89 (6): 3062–3065.
- 89. Gauna, C., Delhanty, P.J., van Aken, M.O., et al.(2006). Unacylated ghrelin is active on the INS-1E rat insulinoma cell line independently of the growth hormone secretagogue receptor type 1a and the corticotropin releasing factor 2 receptor, Molecular and Cellular Endocrinology,251:103–111.
- 90. Guido, M., Romualdi, D., De Marinis, L., Porcelli, T., Giuliani, M., Costantini, B., et al. (2007). Administration of exogenous ghrelin in obese patients with polycystic ovary syndrome: effects on plasma levels of growth hormone, glucose, and insulin. Fertility and Sterility,88:125e130.
- 91. Tassone, F., Broglio, F., Destefanis, S., Rovere, S., Benso, A., Gottero, C., et al. (2003). Neuroendocrine and metabolic effects of acute ghrelin administration in human obesity. The Journal of Clinical Endocrinology and Metabolism,88:5478e5483.
- 92. Toshinai, K., Mondal, M.S., Nakazato, M., Date, Y., Murakami, N., Kojima, M., Kangawa, K., Matsukura, S.(2001). Upregulation of ghrelin expression in the stomach upon fasting, insulin-induced hypoglycemia, and leptin administration. BiochemBiophys Res Commun., 281:1220–1225.
- 93. Green, E.D., Maffei, M., Braden, V.V., Proenca, R., DeSilva, U., Zhang, Y., Chua, S.C Jr., Leibel, R.L., Weissenbach, J., Friedman, J.M. (1995). The human obese (OB) gene. RNA expression pattern and mapping on the physical, cytogenetic, and genetic maps of chromosome 7. Genome Res., 5: 5–12.
- 94. Zhang, Y., Proenca, R., Maffei, M., Barone, M., Leopold, L., Friedman. J.M.(1994). Positional cloning of the mouse obese gene and its human homologue. Nature, 372: 425–432.
- 95. Klok, M.D., Jakobsdottir, S., et al. (2007). The role of leptin and ghrelin in the regulation of food intake and body weight in humans: a review. Obesity reviews.8(1), 21-34.
- 96. Louis A Tartaglia. (1997). The Leptin Receptor. J Bilogical chemistry. 272 (10):6093-6096.
- 97. Gutierrez-Juarez, R., et al. (2004). Melanocortin-independent effects of leptin on hepatic glucose fluxes. J. Biol. Chem., 279(48): 49704-49715.
- 98. Balthasar, N., Coppari, R., Mc Minn, J., Liu, SM., Lee, CE., Tang, V., et al. (2004). Leptin receptor signaling in POMC neurons is required for normal body weight homeostasis. Neuron., 42(6):983-991.
- 99. Pereira, G.D.A.V., Morais, T.C., França, E.L., Daboin, B.E.G., Bezerra, I.M.P., Pessoa, R.S., et al. (2023). Leptin, adiponectin, and melatonin modulate colostrum lymphocytes in mothers with obesity. International Journal of Molecular Sciences., 24(3):2662.
- 100. Denroche, H.C., Levi, J., Wideman, R.D., *et al.* (2011). Leptin therapy reverses hyperglycemia in mice with streptozotocin-induced diabetes, independent of hepatic leptin signaling. Diabetes, 60: 1414–1423.
- 101. German, A.J., Holden, S.L., Bissot, T., Morris, P.J., Biourge, V., (2010). A high protein high fibre diet improves weight loss in obese dogs. Vet J.183(3):294-7. doi: 10.1016/j.tvjl.2008.12.004. Epub 2009 Jan 12. PMID: 19138868.
- 102. Lin, C.Y., Higginbotham, D.A., Judd, R.L., White, B.D. (2002). Central leptin increases insulin sensitivity in streptozotocin-induced diabetic rats American Journal of Physiology-Endocrinology and Metabolism, 282:E1084-E1091.
- 103. Anna, M., D'souza, Ursula, H., Neumann, Maria M. Glavas., Timothy J. Kieffer. (2017). The glucoregulatory actions of leptin. Molecular Metabolism,6 (9):1052-1065.
- 104. Kolaczynski, J.W., Ohannesian, J.P., Considine, R.V., Marco, C.C., Caro, J.F. (1996). Response of leptin to short-term and prolonged overfeeding in humans. *J Clin Endocrinol Metab.*,81:62–4165.
- 105. Ostlund, R.E. Jr., Yang, J.W., Klein, S., Gingerich, R. (1996). Relation between plasma leptin concentration and body fat, gender, diet, age, and metabolic covariates. J Clin Endocrinol Metab.,81: 3909–3913.
- 106. Shanta J. Persaud, Gavin A. Bewick. (2014). Peptide YY: more than just an appetite regulato. Diabetologia.,57:1762–1769.
- 107. Tatemoto, K, Mutt, V. (1980). Isolation of two novel candidate hormones using a chemical method for finding naturally occurring polypeptides. Nature.285:417–418.
- 108. Batterham, R.L., Bloom, S.R. (2003). The gut hormone peptide YY regulates appetite. Ann NY Acad Sci.,994:162–168.

- 109. Chandarana, K., Gelegen, C., Irvine, E.E., et al. (2013). Peripheral activation of the Y2-receptor promotes secretion of GLP-1 and improves glucose tolerance. Mol Metab., 2:142–152.
- 110. Pathak, V., Flatt, P.R., Irwin, N. (2018). Cholecystokinin (CCK) and related adjunct peptide therapies for the treatment of obesity and type 2 diabetes. Peptides.. 100:229-235.
- 111. Rehfeld, J.F.. (2011).Incretin physiology beyond glucagon-like peptide 1 and glucosedependent insulinotropic polypeptide: cholecystokinin and gastrin peptides. Acta Physiol (Oxf).,201:405-411.
- 112. Rehfeld. J.F. (2016). CCK, gastrin and diabetes mellitus. Biomark Med.,10: 1125-1127.
- 113. Drucker, D.J.. (2006). The biology of incretin hormones. Cell Metabol.,3(3):153-165.
- 114. Mentlein, R., Gallwitz, B., Schmidt, W.E. (Dipeptidyl-peptidase IV hydrolyses gastric inhibitory polypeptide, glucagon-like peptide-1(7–36)amide, peptide histidine methionine and is responsible for their degradation in human serum. Eur. J. Biochem...214; 3:829–835.
- 115. Kleinman, EngJ.W.A.,Singh,L., Singh, G., Raufman, J.P. (1992). Isolation and characterization of exendin-4, an exendin-3 analogue, from *Heloderma suspectum* venom: further evidence for an exendin receptor on dispersed acini from guinea pig pancreas. J Biol Chem.,1992;267:7402–5.
- 116. Herman, G.A., Bergman, A., Stevens, C., Kotey, P., Yi, B., Zhao, P., et al. (2006). Effect of single oral doses of sitagliptin, a dipeptidyl peptidase-4 inhibitor, on incretin and plasma glucose levels after an oral glucose tolerance test in patients with type 2 diabetes. J. Clin. Endocrinol. Metabolism,91(11):4612–4619. doi:10.1210/jc.2006-1009
- 117. Vilsboll, T., Krarup, T. (2001). Deacon C F, Madsbad S, Holst J J. Reduced postprandial concentrations of intact biologically active glucagon-like peptide 1 in type 2 diabetic patients. Diabetes., 50(3):609-613.
- 118. Sharma, S., Bhatia, V. (2021). Drug design of GLP-1 receptor agonists: Importance of in silico methods. Curr. Pharm. Des., 27:1015–1024. doi:10.2174/1381612826666201118094502
- 119. He, H., Tran, P., Yin, H., Smith, H., Batard, Y., Wang, L., Einolf, H., Gu, H., Mangold, J.B., Fischer, V., et al.(2009). Absorption, metabolism, and excretion of [14C]vildagliptin, a novel dipeptidyl peptidase 4 inhibitor, in humans. Drug Metab. Dispos. Biol. Fate Chem.,37:536–544. doi: 10.1124/dmd.108.023010.
- 120. Herman, G.A., Stevens, C., Van Dyck, K., Bergman, A., Yi, B., De Smet, M., Snyder, K., et al. (2005). Pharmacokinetics and pharmacodynamics of sitagliptin, an inhibitor of dipeptidyl peptidase IV, in healthy subjects: Results from two randomized, double-blind, placebo-controlled studies with single oral doses. Clin. Pharmacol. Ther.,78:675–688. doi: 10.1016/j.clpt.2005.09.002.
- 121. Covington, P., Christopher, R., Davenport, M., Fleck, P., Mekki, Q.A., Wann, E.R., Karim, A. (2008). Pharmacokinetic, pharmacodynamic, and tolerability profiles of the dipeptidyl peptidase-4 inhibitor alogliptin: A randomized, double-blind, placebo-controlled, multiple-dose study in adult patients with type 2 diabetes. Clin. Ther.30:499–512. doi: 10.1016/j.clinthera.2008.03.004.
- 122. Graefe-Mody, U,.Retlich, S., Friedrich, C. (2012). Clinical Pharmacokinetics and Pharmacodynamics of Linagliptin. Clin. Pharmacokinet.,51:411–427.
- 123. Makrilakis, K. (2019). The Role of DPP-4 Inhibitors in the Treatment Algorithm of Type 2 Diabetes Mellitus: When to Select, What to Expect. Int J Environ Res Public Health.16(15):2720. doi: 10.3390/ijerph16152720. PMID: 31366085.
- 124. Gallwitz, B.(2019). Clinical Use of DPP-4 Inhibitors. Front Endocrinol (Lausanne). 10:389. doi: 10.3389/fendo.2019.00389. PMID: 31275246; PMCID: PMC6593043.
- 125. Deacon, C.F (2019). Physiology and pharmacology of DPP-4 in glucose homeostasis and the treatment of type 2 diabetes. Front. Endocrinol. (Lausanne) 10, 80–14. 10.3389/fendo.2019.00080.
- 126. Esler, W.P., Rudolph, J., Claus, T.H., *et al.* (2007). Small-molecule ghrelin receptor antagonists improve glucose tolerance, suppress appetite, and promote weight loss. Endocrinology,148(11):5175–5185.
- 127. Yang, J., Zhao, T.J., Goldstein, J.L., Brown, M.S.(2008).Inhibition of ghrelin O-acyltransferase (GOAT) by octanoylated pentapeptides. Proc. Natl Acad. Sci. USA,105(31), 10750–10755.
- 128. Bodvarsdottir, T.B., Hove, K.D., Gotfredsen, C.F., et al. (2010). Treatment with a proton pump inhibitor improves glycaemic control in *Psammomysobesus*, a model of type 2 diabetes. Diabetologia,53:2220–2223. https://doi.org/10.1007/s00125-010-1825-6
- 129. Kirchner, H., Heppner, K.M., Tschop, M.H.. (2012). The role of ghrelin in the control of energy balance. Handbook Exp. Pharmacol.,209, 161–184.
- 130. Suarez-Pinzon, W.L., Power, R.F., Yan, Y., Wasserfall, C., Atkinson, M., Rabinovitch, A. (2008). Combination therapy with glucagon-like peptide-1 and gastrin restores normoglycemia in diabetic NOD mice. Diabetes., 2008;57:3281-3288.
- 131. Fosgerau, K., Jessen, L., Lind Tolborg, J., Osterlund, T., Schaeffer Larsen, K., Rolsted, K., Brorson, M., Jelsing, J., Skovlund Ryge Neerup, T. (2013). The novel GLP-1-gastrin dual agonist, ZP3022, increases β-cell mass and prevents diabetes in db/db mice. Diabetes Obes Metab.,15(1):62-71. doi: 10.1111/j.1463-1326.2012.01676.x.Epub 2012 Sep 9. PMID: 22862961.
- 132. Escalona, A., Pimentel, F., Sharp, A., *et al.* (2012). Weight loss and metabolic improvement in morbidly obese subjects implanted for 1 year with an endoscopic duodenal–jejunal bypass liner. Ann. Surg.,255(6):1080–1085.

- 133. De Moura, E.G., Martins, B.C., Lopes, G.S., *et al.* (2012).Metabolic improvements in obese Type 2 diabetes subjects implanted for 1 year with an endoscopically deployed duodenal-jejunal bypass liner. Diabetes Technol. Ther..2012; 14(2):183–189.
- 134. Soliman, A.T., Omar, M., Assem, H.M., et al.(2002). Serum leptin concentrations in children with type 1 diabetes mellitus: relationship to body mass index, insulin dose, and glycemic control. Metabolism, 51: 292–296.
- 135. Denroche, H.C., Levi, J., Wideman, R.D., *et al.* (2011). Leptin therapy reverses hyperglycemia in mice with streptozotocininduced diabetes, independent of hepatic leptin signaling. Diabetes,60: 1414–1423.
- 136. Claudia Guida, Sam D Stephen, Michael Watson, Niall Dempster, Pierre Larraufie, et al. (2019). PYY plays a key role in the resolution of diabetes following bariatric surgery in humans. EBioMedicine,40:67-76. doi: 10.1016/j.ebiom.2018.12.040. Epub 2019 Jan 11.
- 137. Behary, P., Tharakan, G., Alexiadou, K., Johnson, N., WewerAlbrechtsen N.J., Kenkre, J., et al. (2019). Combined GLP-1, oxyntomodulin, and peptide YY improves body weight and glycemia in obesity and Prediabetes/type 2 diabetes: a randomized single-blinded placebo controlled study. Diabetes Care, 8.
- 138. Batterham, R.L., Bloom, S.R. (2003). The gut hormone peptide YY regulates appetite. Ann NY Acad Sci.,994:162–168.
- 139. Cohen, M.A., Ellis, S.M., LeRoux C.W., Batterham, R.L., Park, A., Patterson, M. (2003). Oxyntomodulin suppresses appetite and reduces food intake in humans. Journal of Clinical Endocrinology and Metabolism,88:4696–4701. doi: 10.1210/jc.2003-030421.
- 140. BT, Baldissera FG., Mortensen, P.E., Holst, J., Christiansen, J. (1988). Oxyntomodulin. a potential hormone from the distal gut. Pharmacokinetics and effects on gastric acid and insulin secretion in man. European Journal of Clinical Investigation.18:499–503. doi: 10.1111/j.1365-2362.1988.tb01046.x.
- 141. Pocai, A. (2013). Action and therapeutic potential of oxyntomodulin. Mol Metab., 3(3):241-51. doi: 10.1016/j.molmet.2013.12.001. PMID: 24749050
- 142. lvy, A.C., Oldberg, E. (1928). A hormone mechanism for gallbladder contraction and evacuation. Am. J. Physiol., 86: 599-613.
- 143. Bataille, D., Gespach, C., Tatemoto, K., Marie, J.C., Coudray, A.M., et al. (1981). Bioactive enteroglucagon (oxyntomodulin): Present knowledge on its chemical structure and its biological activities. Peptides,2(2): 41-44.
- 144. Holst, J.J. (2019). The incretin system in healthy humans: The role of GIP and GLP-1. Metabolism.46-55. doi: 10.1016/j.metabol.2019.04.014.