

PHYTOCHEMICAL SCREENING AND ACUTETOXICITY STUDY OF AQUEOUS LEAF EXTRACT OF *Telfairia occidentalis* ON ALBINO RATS

ABSTRACT

Aim: This study investigated the phytochemical contents and toxicological effects of the aqueous leaf extract of *T. occidentalis* on albino rats.

Study Design: Toxicological study

Place and Duration of the study: Department of Biochemistry and Molecular Biology Federal University, Birnn Kebbi, Kebbi State Nigeia, Between July to October, 2024.

Methodology: Twenty (20) albino rats of both sexes were randomized into four (A, B, C and D) different groups of five (5) animals each. Group A was given distilled water, group B was administered with 200mg/kg body weight of aqueous leaf extract of *T. occidentalis*, Group C was administered 300mg/kg body weight of aqueous leaf extract of *T. occidentalis* and Group D was administered 400mg/kg body weight of aqueous leaf extract of *T. occidentalis* orally for 14 days. The animals were sacrificed and blood sample was collected for haematological and biochemical analyses. **Results:** Qualitative phytochemical screening of the aqueous leaf extract of *T. occidentalis* revealed the presence of Alkaloids, Flavonoids, Phenols, Saponins, Tannins, Terpenoids and Carbohydrate. A significant ($p < 0.05$) increase in the weight of the experimental rats compared to the control group was observed in all the groups treated with aqueous leaf extract of *T. occidentalis*. Oral administration of different doses of *T. occidentalis* to the experimental rats significantly ($p < 0.05$) increased the concentration of RBC, HCT, GRA, PLT, LYM, MCV, MCH, Hb and WBC in a dose dependent manner when compared with the control. Administration of different doses of *T. occidentalis* causes significant decrease ($P < 0.05$) in AST, ALT, ALP, TB and DB levels while significant increase ($p < 0.05$) of TP and albumin concentration were observed when compared with control. Similarly, administration of different doses of *T. occidentalis* significantly ($p < 0.05$) decreased the concentration of Urea, Creatinine, Na^+ and K^+ while the concentration of Cl^- was significantly ($p < 0.05$) increased in a dose dependent manner when compared with the control. **Conclusion:** In conclusion, the findings showed that the aqueous leaf extract of *Telfairia occidentalis* is relatively non-toxic at acute exposure doses exceeding 5000 mg/kg and exhibits potential anti-anemic effects.

Key words: Haematological, Kidney, Liver, *Telfairia occidentalis*, Toxicity

1.0 Introduction

Nigeria possesses a diverse array of genetic resources comprising cultivated, semi-wild, and wild species of crops utilized as traditional vegetables, with various ethnic groups consuming these for a multitude of reasons (Akinola *et al.*, 2020; Aro *&* Jayeola, 2024). Edible foliage from vegetable plants is incorporated into both supplementary food items and primary dishes (Akindele *et al.*, 2018). These leaves may exhibit aromatic, bitter, or neutral flavor profiles; however, they represent the most economical and readily available sources of proteins, vitamins, minerals, and essential amino acids. Furthermore, they contain certain hormonal precursors alongside energy (Olorunfemi *et al.*, 2014).

Leafy vegetables confer significant health benefits and serve a crucial role in disease prevention. They provide an invaluable source of dietary components that can be effectively harnessed to enhance and fortify the human body (Mutiu *&* Abidemi, 2013). These vegetables are characterized by their substantial carbohydrate, vitamin, and mineral concentrations.

The plant *Telfairia occidentalis* Hook. f. (Cucurbitaceae), commonly designated as “fluted gourd” or “fluted pumpkin,” is cultivated in West Africa primarily for its leaves and edible seeds (Mutiu *&* Abidemi, 2013). Within Nigeria, the leaves are consumed across various regions due to their nutritional and medicinal properties (Ojo *et al.*, 2012; Aro *&* Jayeola, 2024). It is recognized by distinct traditional names: “Ugu” among the Igbos, “Iroko” among the Yoruba, “Ubong” among the Efik, and “Umeke” among the Edo (Saaluet *al.*, 2010). In traditional medicine, the fresh leaves are employed in the treatment of conditions such as anemia, acute convulsions, and malaria (Mutiu *&* Abidemi, 2013). Prior research has documented the hypoglycemic, antinociceptive, and anti-inflammatory properties of extracts derived from *T. occidentalis* (Mutiu *&* Abidemi, 2013). It has anxiolytic and sedative properties (Mutiu *&* Abidemi, 2013), blood coagulation (Thomas *et al.*, 2013), immunomodulatory (Okokon *et al.*, 2012), testis-protective (Saaluet *al.*, 2010). Hence, this study aims to investigate the hematological effects and toxicological implications of the aqueous leaves extract of *T. occidentalis* on liver and kidney function indices.

2.0 Materials and Methods

2.1. Chemicals and Reagents

Chemicals and reagents of analytical grade were used for the experiment.

2.2. Collection of Plant Material and Aqueous Extraction

The fresh foliage of *Telfairia occidentalis* was procured from the vegetable section of the Birnin Kebbi Central Market, situated in Kebbi State. The leaves underwent a rigorous washing process and were subsequently chopped into diminutive fragments, followed by air-drying until a stable weight was achieved. The dry leaves were then subjected to milling into a fine powder utilizing an electric blender, resulting in 500 g of the pulverized substance. A quantity of one hundred grams (100 g) of the plant powder was subjected to maceration in 500ml of distilled water for duration of 72 hours. The resultant extract was then decanted and subjected to filtration twice

employing Whatman filter paper. The filtrate was concentrated to dryness utilizing a rotary evaporator set at 40°C.

2.3 Experimental Animals

Twenty (20) albino rats of both sexes weighing 100 – 200 g were used for the study. The rats were kept at the animals' house under normal environmental conditions and maintained with free access to pelletized growers' feed and access to water ad libitum. The albino rats were allowed to acclimatize for 14 days. All animal procedures were performed according to the Guide for the Care and Use of Laboratory Animals of the National Institute of Health and the guidelines of the Animal Welfare Act, 1999.

2.4 Experimental Design

Albino rats were used for this study. After acclimatization, all animals were housed in cages at regulated room temperature, allowed daily 12 hours light and dark cycle, feed with pelletized growers feed and allowed free access to water during the experiment.

The animals were grouped into four (4) groups of five (5) rats each.

Treatment details

Groupings	Treatments
Group A	Control (Administered distilled water and feeds only)
Group B	Administered aqueous extract of <i>Telfairia occidentalis</i> leaves extract at 200mg/kg body weight
Group C	Administered aqueous extract of <i>Telfairia occidentalis</i> leaves extract at 300mg/kg body weight
Group D	Administered aqueous extract of <i>Telfairia occidentalis</i> leaves extract at 400mg/kg body weight

2.5 Qualitative phytochemical screening

Five grams (5g) of crude extracts was dissolved in 40 ml of distilled water and thereafter subjected to phytochemical screening through established methodologies (Harborne, 1973; Trease & Evans, 1989; Sofowora, 1993).

2.6 Oral Acute Toxicity Study of *Telfairia occidentalis*

The median lethal dose (LD50) of the aqueous extract derived from the foliage of *Telfairia occidentalis* was assessed to determine appropriate safe dosages for the appraisal of the aqueous extract's effects. This assessment was executed utilizing the protocol delineated by Lorke (1983). During the preliminary phase, rats were categorized into three cohorts of three individuals each and were administered 10 mg, 100 mg, and 1000 mg of aqueous extract per kg of body weight via oral ingestion. They were monitored for 24 hours for manifestations of toxicity, encompassing alterations in behavior and mortality.

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In phase II, three rats were allocated into three separate groups, each consisting of one rat, and were treated with the aqueous extract in accordance with the findings obtained from the initial phase. In alignment with the survivorship observed in phase I, three rats were individually administered 1600, 2900, and 5000 mg/kg body weight of the aqueous extract fraction in the second phase, respectively, and the incidence of death within 24 hours were meticulously documented. The LD50 was derived from the outcomes of the final phase as the square root of the product of the minimal lethal dose and the maximal non-lethal dose, specifically, the geometric mean of the successive doses with recorded survival rates of 0% and 100% (Lorke, 1983).

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The mathematical expression of Geometric mean (LD₅₀) = $\sqrt{X * Y}$

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Where X and Y represents lowest lethal dose and the highest non-lethal dose respectively.

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2.7 Hepato and Renal toxicity Effect

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Twenty albino rats were divided into four groups of five rats each. The first group received distilled water (0.2ml) as control while group 1 to group 3 received 200mg/kgbw, 300mg/kg bw and 400mg/kgbw of the extract respectively. The animals were sacrificed two weeks after administration to assess the effects of aqueous leaves extract of *T. occidentalis* on biochemical parameters.

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2.8 Collection of Blood Sample

After duration of two weeks during which various concentrations of the extracts were administered, the albino rats underwent an overnight fasting period. The rats were subjected to anesthesia by being placed within a sealed container filled with cotton wool that had been saturated with diethyl ether in an inhalation apparatus. The albino rats were euthanized via decapitation, and subsequently, blood specimens were collected and subjected to centrifugation at 4000 ×g for duration of 10 minutes at a temperature of 4°C. The resulting supernatant was preserved at a temperature of 37°C for subsequent biochemical analyses.

2.9 Haematological Analysis

Blood was collected in an EDTA blood sample container. It was gently swirled to mix with the EDTA so as to prevent it from coagulating. The sample container was placed at the seeping inlet of the automated haematological analyser. ~~The automated haematological analyser (Medonic M32S Cell Counter, India) seeped in the blood sample and then analysed the~~ haematological parameters, like Packed Cell Volume (PCV), Haemoglobin concentration, Red Blood Cells count

(RBC), White Blood Cells count (WBC) (neutrophils, lymphocytes, eosinophils, monocytes, T. occedentalisphiles), Haematocrit (Hct), Platelets, Mean Cell Haemoglobin Concentration (MCHC) and Mean Cell Haemoglobin (MCH),~~was~~ ~~were~~ analysed using an automated haematological analyser (Medonic M32S Cell Counter, India).

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2.10 Determination of Liver Function Indices

Alkaline phosphatase (ALP) activity was evaluated by the Para-Nitrophenyl phosphate (PNPP) method (Kind and King, 1954), Alanine aminotransferase (ALT) and Aspartate aminotransferase (AST) according to the methods (Reitman and Frankel, 1957). Serum Total Protein (TP), Albumin Total and Direct bilirubin as described by (Tietz, 1976). Assay kits used were obtained from the Randox

2.11 Determination of Renal Function Indices

Urea and creatinine were estimated using the diagnostic kits supplied by Randox. In addition, blood electrolytes levels (sodium, potassium and chloride) were also measured using the Randox kit. The procedure described in each user's manual of the kit was followed while estimating the biomarker parameters.

2.12 Data Analysis

The results were presented as Mean and Standard error means (SEM), and n represents the number of animals used. The differences between means were carried out using one-way analyses of variance (ANOVA) using the statistical software SPSS version 20. The Duncan post Hoc comparison test was used to check differences between individual groups, and mean differences were considered significant when $P < 0.05$.

3.0 Results and Discussion

3.1 Extraction

A 40.6g of dark brownish solid extract with 20% yield was obtained after 72hours of extraction with aqueous. It was then ~~labeled~~ as crude extract.

3.2 Phytochemical composition of the crude extract

Table 1 showed the qualitative phytochemical screening of the aqueous extract of *T. occidentalis*. After the phytochemical screening, the presence of Alkaloids, Flavonoids, Phenols, Saponins, Tannins and Terpenoids and Carbohydrate were detected in the aqueous leaf extract. But, Anthraquinones and Anthracyanins were not detected in the sample

Table 1: Qualitative Phytochemical Content of Crude Aqueous Extract of *T. occidentalis*

Test	Crude Extract
Alkaloids	+
Anthraquinones	-
Flavonoids	+
Phenols	+
Saponins	+
Steroids	+
Tannins	+
Anthracyanins	-
Carbohydrate	+

Key: + = detected, - = not detected

3.3. Oral Acute Toxicity Studies

The Median Lethal Dose (LD50) of aqueous extract (leaves) on the tested *albino* rats was presented in Table 2. There was no mortality recorded at 10, 100, 1000, 1600, 2900 and 5000 mg/kg body weight respectively. The LD50 of aqueous extract(leaves) was taken to be >5000 mg/kg body weight.

Table 2: LD₅₀ of the aqueous extracts of *T. occidentalis*

	Dosage(mg/kg)bw.	Number of rats used	Mortality	% Lethality
Phase 1	10	3	0/3	0
	100	3	0/3	0
	1000	3	0/3	0
Phase 2	1600	1	0/1	0
	2900	1	0/1	0
	5000	1	0/1	0

The LD₅₀ of the extracts was ~~determined~~determined to be greater than 5000 mg/kg body weight

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3.4 Effect of *T. occidentalis* on Body Weight

Figures 1 and 2 showed the weight gained observed across the groups in phase I and II. There was an increased increase in body weight among the treated groups (B,C and D) when compared with the control group (A).

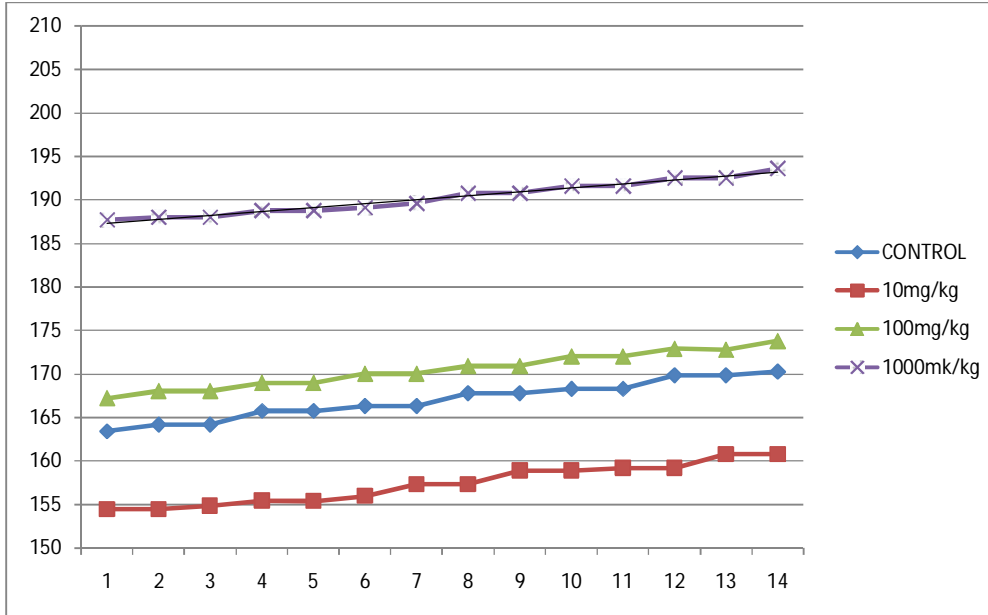


Figure 1: Change in body weight of the rats during acute toxicity study of *T. occidentalis* (Phase I).

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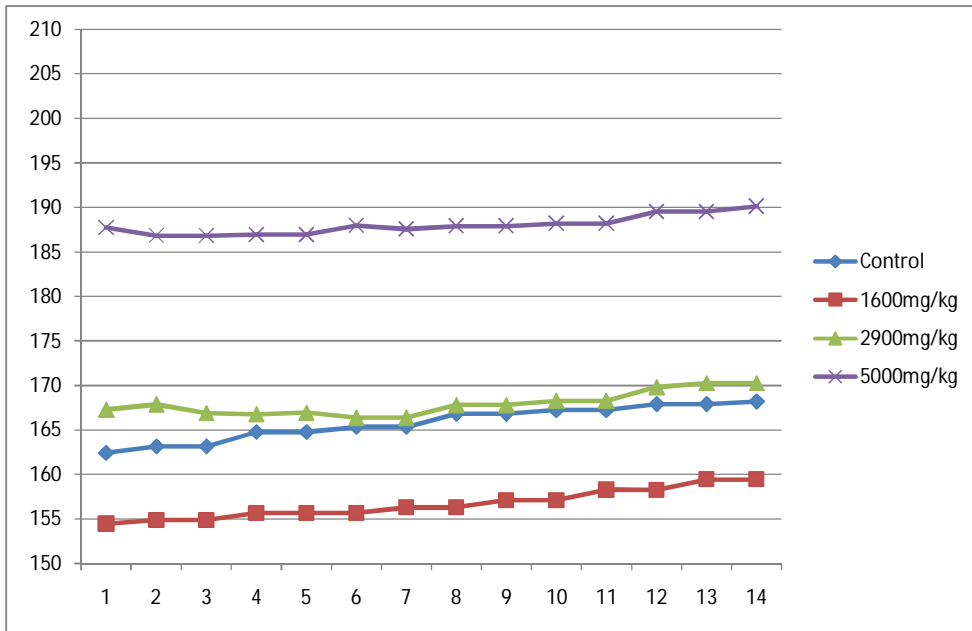


Figure 2: Change in body weight of rats during acute toxicity study of *T. occidentalis* (Phase II)

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3.5 Effect of *T. occidentalis* Haematological Indices

Table 3 showed the effect of oral administration of different doses of *T. occidentalis* haematological indices. *T. occidentalis* extract significantly ($p < 0.05$) increased the concentration of RBC, HCT, GRA, PLT, LYM, MCV, MCH, Hb and WBC in a dose dependent manner when compared with the control.

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Table 3: Effect of Administration of Different doses of *T. occidentalis* Haematological Indices

Doses (mg/kg)	RBC (10 ¹² /L)	HCT (g/dL)	GRA (pg)	PLT (fL)	MCHC (g/dL)	LYM (%)	MCV (x10 ⁹ /L)	MCH (x10 ⁹ /L)	Hb (g/dl)	WBC (x10 ⁹ /L)
Distilled Water (0.2ml)	5.9±0.3 ^a	33.4±0.5 ^a	0.3±0.2 ^a	67.4±0.5 ^a	41.3±0.1 ^a	75.4±0.5 ^a	53.6±0.6 ^a	15.2±0.3 ^a	10.6±0.1 ^a	8.3±0.3 ^a
200mg/kg	6.4±0.6 ^b	35.5±0.4 ^b	0.4±0.3 ^b	70.4±0.7 ^b	42.6±0.3 ^b	78.5±0.5 ^b	55.2±0.4 ^b	16.4±0.9 ^b	11.1±0.2 ^b	8.4±0.6 ^{ab}
300mg/kg	6.6±0.5 ^c	39.7±0.6 ^c	0.5±0.2 ^c	73.4±0.3 ^c	44.3±0.1 ^c	80.6±0.6 ^c	58.3±0.3 ^c	17.4±0.4 ^c	12.2±0.3 ^c	8.6±0.1 ^b
400mg/kg	7.6±0.3 ^d	43.4±0.3 ^d	0.7±0.1 ^d	76.3±0.2 ^d	45.8±0.2 ^d	83.3±0.4 ^d	60.2±0.3 ^d	19.1±0.1 ^d	13.4±0.2 ^d	9.2±0.2 ^c

Values were expressed as mean ± standard error of mean, n = 3. Mean values having common superscript letters in a column are not significantly different (P<.05) (one-way ANOVA followed by Duncan's multiple range test). RBC- Red Blood Count, MCHC-Mean Cell Haemoglobin Concentration, MCH- Mean Cell Haemoglobin, MCV- Mean Corpuscular Volume, HGB- Haemoglobin, PCV- Packed Cell Volume, PLT- Platelets, WBC- White Blood Count.

3.6 Effect of *T. occidentalis* on Liver Function Indices.

Table 4 shows the effect of administration of different doses of *T. occidentalis* on liver function indices. *T. occidentalis* causes a significant decrease ($P < 0.05$) in AST, ALT, ALP, TB and DB levels while a significant increase ($p < 0.05$) of ~~in~~TP and albumin concentrations ~~were~~ was observed when compared with control.

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Table 4: Effect of Oral administration of different doses of *T.occidentalis* on liver function indices

TREATMENT	AST (U/l)	ALT (U/l)	ALP (U/l)	TP (g/dl)	ALB (g/dl)	TB (mg/dl)	DB (mg/dl)
Distilled Water (2ml/kg)	13.50±0.11 ^c	38.67±1.76 ^b	70.67±0.88 ^c	7.29±0.12 ^a	5.50±0.01 ^a	1.05±0.08 ^c	0.32±0.03 ^b
200mg/kg	12.59±0.12 ^b	35.33±0.88 ^b	68.00±1.73 ^c	7.77±0.13 ^b	5.17±0.17 ^a	0.92±0.04 ^b	0.26±0.01 ^a
300mg/kg	12.35±0.04 ^b	30.33±0.88 ^a	60.33±2.03 ^b	8.00±0.06 ^b	6.53±0.20 ^b	0.86±0.01 ^{ab}	0.25±0.01 ^a
400mg/kg	11.170±0.23 ^a	26.67±0.88 ^a	46.67±1.45 ^a	7.93±0.09 ^b	7.33±0.09 ^c	0.74±0.01 ^a	0.22±0.01 ^a

Values were expressed as mean ± standard error of mean, n = 3. Mean values having common superscript letters in a column are not significantly different (P<.05) (one-way ANOVA followed by Duncan's multiple range test). ALP-Alkaline Phosphatase, AST-Aspartate Amino Transferase, ALT-Alanine Amino Transferase, ALB-Albumin, TP-Total Protein, DB-Direct Bilirubin and TB-Total Bilirubin,

3.7 Effect of *T. occidentalis* on Kidney Function Indices.

Table 5 shows the effect of oral administration of different doses of *T. occidentalis* on kidney indices. *T. occidentalis* significantly ($p < 0.05$) decreased the concentration of Urea, Creatinine, Na^+ and K^+ while the concentration of Cl^- was significantly ($p < 0.05$) increased in a dose dependent manner when compared with the control.

Table 5: Effect of Administration of Different Doses of *T. occidentalis* on Kidney Function Indices

Doses (mg/kg)	Urea (mg/dl)	Cr (mg/dl)	Na^+ (meq/l)	K^+ (meq/l)	Cl^- (meq/l)
Distilled Water (0.2ml)	27.35±0.04 ^d	6.51±0.04 ^c	147.28±0.03 ^d	4.63±0.03 ^d	104.32±0.01 ^a
200mg/kg	24.15±0.02 ^c	6.51±0.02 ^c	146.43±0.06 ^c	4.51±0.03 ^c	104.51±0.07 ^b
300mg/kg	23.20±0.03 ^b	6.31±0.03 ^b	145.57±0.05 ^b	4.30±0.02 ^b	104.54±0.01 ^b
400mg/kg	20.28±0.04 ^a	6.15±0.02 ^a	142.27±0.03 ^a	4.19±0.02 ^a	104.71±0.03 ^c

Values were expressed as mean ± standard error of mean (n = 5). Mean values having common superscript letters in a column are not significantly different ($P < 0.05$) (one-way ANOVA followed by Duncan's multiple range test). Cr-Creatinine, Na- Sodium, K- Potassium and Cl^- - Chloride

3.8 Discussion

Plants are widely recognized in the pharmaceutical industry for their phytochemical content and diverse pharmacological activities, which can be utilized in the treatment of both chronic and infectious diseases (Junejoet *et al.*, 2021). The preliminary phytochemical screening of *Telfairia occidentalis* revealed the presence of tannins, flavonoids, glycosides, phenols, and terpenoids. These findings are consistent with previous studies conducted by Ojo *et al.* (2012), Ogbonnaya *et al.* (2010), Mutiu & Abidemi (2013). These phytochemicals are known to have various therapeutic applications (Shivashankar *et al.*, 2019; Ukwuani-Kwaja *et al.*, 2021; Ugwah, 2023). For example, tannins are diverse organic compounds with astringent properties that promote wound healing and soothe inflamed mucous membranes (Orlowskiet *et al.*, 2018). Additionally, tannins exhibit antibiotic properties (Motaharesadatet *et al.*, 2025). Alkaloids, another class of phytochemicals, are reported to possess analgesic, anti-inflammatory, and adaptogenic activities, which help alleviate pain, enhance disease resistance, and improve stress endurance (Mukta *et al.*, 201).

In the present study, administering an aqueous leaf extract of *T. occidentalis* significantly enhanced the percentage weight gain in treated rats compared to the normal control group. This effect is likely attributed to the nutrient-rich composition of *T. occidentalis* leaves. These results align with the findings of Akinola *et al.* (2020). Furthermore, Olorunfemi *et al.* (2014) reported that *T. occidentalis* exhibits inhibitory effects on intestinal microbes such as *E. coli* and *S. typhi*, which could enhance gastrointestinal tract efficiency by improving the absorption of digested nutrients.

The acute toxicity of the aqueous leaf extract of *T. occidentalis* was evaluated through the calculation of its median lethal dose (LD₅₀), defined as the dose required to kill 50% of a test population. The LD₅₀ was found to be greater than 5000 mg/kg, indicating a wide safety margin in acute toxicity tests conducted on albino rats (Lorke, 1983). This finding aligns with the study by (Adisa *et al.*, 2024) but contradicts the results of (Mutiu and Abidemi, 2013). The observed discrepancies may be attributed to geographical and environmental factors, as well as variations in plant age and harvesting time. Hayes (Hayes, 1989) reported that no dose-related toxicity should be considered above 5000 mg/kg body weight while the Hodge and Sterner Scale of toxicity classes categorized products with LD50 value > 5000 mg/kg as practically non-toxic (Dawoud *et al.*, 2015)

Results of studies of haematological properties of *T. occidentalis* leave showed a significant of haematological parameters i.e., RBC, Hb concentration, MCV, Mean Corpuscular Haemoglobin MCHC and PLT. This finding agreed with the report of (Udosen and Osu, 2022; Suleiman *et al.*, 2022; Alada, 2000; Ochokwuet *al.*, 2021). The increases in the haematological parameters could be due to the chemical composition of the leaves of *T. occidentalis*. According to Ochokwuet *al.* (2021) and Okonwuet *al.* (2018) the nutrient composition of the *T. occidentalis* includes protein, fat, carbohydrate, calcium, iron, vitamin A, vitamin E, vitamin K, vitamin C, thiamine and riboflavin. Most of these constituents are well-known haematological factors that have direct influence on the production of blood from the bone marrow (Udosen and Osu, 2022). For instance, iron is a well-established haemopoietic factor and deficiency of it produces anaemia. The haematological activity of *T. occidentalis* lead extracts is thought to be enhanced by these compounds. Many bioactive components of *T. occidentalis* such as vitamin C and E, phenolic and flavonoids have been reported to have natural antioxidant potential. According to the Okonwuet *al.* (2018), vitamin C is a major antioxidant in the human body and possesses the capacity to participate in enzymatic and hydroxylation reactions, participates as well as in the oxidation-reduction processes. It also promotes the absorption of microelements such as iron and copper, is involved in trace element metabolism and protects red blood cells from damage caused by free radicals and environmental pollution.

ALT, AST and ALP activities are commonly measured to monitor potential of plant in drug induced hepatic injury in both pre-clinical studies and human patients and thus, they serve as biomarkers of liver toxicity (Meunier and Larrey, 2019). Non-significant increase of serum ALT, AST and ALP activities in albino rats administered with *T. occidentalis* is an indication that *T. occidentalis* has no significant effect on cellular integrity of the liver. A dose dependent decrease in ALT, AST and ALP activities following administration with crude extract of *T. occidentalis* suggests that *T. occidentalis* might possess hepatoprotective potentials. This finding was in line with report of Osonuga (Osonuga *et al.*, 2020). The outcomes of this study also agree with the reports of Oboh, Ekpenyong and Agada (Oboh, 2005; Ekpenyong *et al.*, 2021; Agada *et al.*, 2024) but contradict the reports of Ogunmoyole and Eze (Ogunmoyole *et al.*, 2019; Eze, *et al.*, 2020).

Albumin and total proteins are globular proteins synthesized by the liver and found in the serum. A decrease in their levels often indicates a reduced synthetic capacity of the liver or impaired hepatocellular function (Yakubu *et al.*, 2003). In the present study, the absence of alterations in serum total protein levels suggests that liver physiology remained unaffected. Bilirubin, a

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byproduct of hemoglobin breakdown, is processed and excreted by the liver. Elevated bilirubin levels can indicate liver dysfunction or an increased rate of red blood cell (RBC) destruction. This study observed no significant differences in serum total and direct bilirubin concentrations between the control group and the groups treated with *Telfairia occidentalis*. These findings indicate that *T. occidentalis* did not interfere with bilirubin metabolism in the liver.

The kidneys play a vital role in the elimination of metabolic waste products and toxic substances. When exposed to toxic substances, kidney function can be compromised, leading to the leakage of biochemical substances into the bloodstream (Frąket *et al.*, 2024). Urea and uric acid, the primary nitrogenous end-products of protein catabolism, along with creatinine, a byproduct of muscle energy metabolism, are transported to the kidneys for excretion. Healthy kidneys effectively remove these compounds from the bloodstream, ensuring their elimination in the urine. In this study, the serum levels of urea and creatinine were unaffected across all groups treated with *T. occidentalis*. This indicates that *T. occidentalis* did not cause any adverse effects on kidney function, suggesting its potential safety for consumption. The outcome of this study agreed with finding of [Akinola](#) (Akinola *et al.*, 2020).

3.9 Conclusion

The findings indicate that the aqueous leave extract of *Telfairia occidentalis* is relatively non-toxic at acute exposure doses exceeding 5000 mg/kg and exhibits potential anti-anemic effects.

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COMPETING INTERESTS

Authors have declared that no competing interests exist

Disclaimer (Artificial intelligence)

Option 1:

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

References

1. Adisa, W., N. A., & Osim, E. (2024). Effect of *Telfairia occidentalis* on Markers of Oxidative Stress in Indomethacin Induced Gastric Ulcer. *African Journal of Biomedical Research*, 17, 137- 142.
2. Agada, S., Odama, R., & Kenchukwu, C. (2024). Antioxidant and hepatoprotective effects of methanolic seed extract of *Telfairia occidentalis* on carbon tetrachloride induced hepatic damage in wistar rats. *Discovery of Medicine*, 35-42.
3. Akindele, A., Oladimeji-Salami, J., Oyetola, R., & Osiagwu, D. (2018). Sub-Chronic Toxicity of the Hydroethanolic Leaf Extract of *Telfairia occidentalis* Hook. f. (Cucurbitaceae) in Male Rats. *Medicines*, 5(4), 1-22.
4. Akinola, B., Olawuy, T., & Ogunmokuwa, A. (2020). The protective effects of *Telfairia occidentalis* on potassium bromate induced hepatotoxicity in adult Wistar rats. *African Journal of Biological Sciences*, 2(3), 51-61.
5. Alada, A. (2000). The Haematological Effect of *Telferia Occidentalis* Diet Preparation. *African Journal of Biomedical Research*, 3, 185-186.
6. Aro, e., & Jayeola, d. (2024). Effects of aqueous leaf extract of *Telfairia occidentalis* on acyclovir induced renal. *Journal of Applied Science Environment Management*, 28 (6), 1679-1684.
7. Dawoud, a., Shayoub, m., & Shayoub, s. (2015). Acute Toxicity Studies of Ethanolic Extract of *Eucalyptus Camaldulensis* Dehnh Leaves. *Journal of Network Communications and Emerging Technologies (JNCET)*, 1-5.
8. Ekpenyong, C., Akpan, E., & Udoh, N. (2012). Phytochemistry and Toxicity Studies of *Telfairia occidentalis* Aqueous Leaves tract on Liver Biochemical Indices in Wistar Rats. *American Journal of Medical Science*, 5(2), 103-110.
9. Eze, B., Ogbodo, E., Ezejindu, D., Ezeugwunne, I., Analike, R., Onuora, I., *et al.* (2020). Hepatoprotective potential of the aqueous leaf extract of *Telfairia occidentalis* on the Liver function parameters in Adult Wistar Rats. *Annals of Geriatric Education and Medical Sciences*, 7(1), 39–42.
10. Frąk, W., Dąbek, B., Balcerzyk-Lis, M., Motor, J., Radzioch, E., Młynarska, E., *et al.* (2024). Role of Uremic Toxins, Oxidative Stress, and Renal Fibrosis in Chronic Kidney Disease. *Antioxidants*, 13(687).
11. Harborne, J. (1973). *Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis*. (1st Edition ed.). London.: Chapman and Hall, UK.

12. Hayes, A. (1989). Guidelines for acute oral toxicity testings. In *Principles and Methods of Toxicity, 2nd ed* (p. 105). New York, USA: Raven Press Ltd.
13. Junejo, J., Zaman, K., Rudrapal, M., Celik, S., & Attah, E. (2021). Antidiabetic bioactive compounds from *Tetrastigma angustifolia* (Roxb.) Deb and *Oxalis debilis* Kunth.: Validation of ethnomedicinal claim by in vitro and in silico studies. *South African Journal of Botany, 143*, 164-175.
14. Kind, R., & King, E. (1954). Determination of alkaline phosphatase activity by colorimetric method. *Journal Clinical Pathology*, 1-10.
15. Lorke, D. (1983). A new approach to practical acute toxicity testing. *Archives of Toxicology, 54*, 272-289.
16. Lorke, D. (1983). A new approach to practical acute toxicity testing. *Archives of Toxicology, 275-287*.
17. Meunier, L., & Larrey, D. (2019). Drug-Induced Liver Injury: Biomarkers, Requirements, Candidates, and Validation. *Frontiers Pharmacology, 10*(1482), 25-38.
18. Motaharesadat, H., Lalehvasht, M., Leonie, B., Silvia, C., Dietmar, W. H., & S., F. .. (2025). The multifaceted role of tannic acid: From its extraction and structure to antibacterial properties and applications. *Progress in Polymer Science, 160*, 1-33.
19. Mukta, G., Naresh, S., Monica, G., Reena, G., Kalvatala, S., & Bhupinder, K. (2021). Herbal bioactives in treatment of inflammation: An overview. *South African Journal of Botany, 205-225*.
20. Mutiu, Y. A., & Abidemi, J. A. (2013). Anxiolytic and sedative properties of hydroethanolic extract of *Telfairia occidentalis* leaves in mice. *Brazilian Journal of Pharmacognosy, 23*(2), 301-309.
21. Oboh, G. (2005). Hepatoprotective property of ethanolic and aqueous extracts of *Telfairia occidentalis* (Fluted pumpkin) leaves against gastric – induced oxidative stress. *Journal of Medicinal Food, 560-563*.
22. Ochokwu, I. J., Taiwo, M., & Bashir, S. (2021). Haematological Indices and Carcass Composition of African Catfish *Clarias gariepinus* (Burchell, 1822) Fingerlings Fed with Fluted Pumpkin Leaf (*Telfairia Occidentalis*) as Feed Additives. *Nigerian Journal of Biotechnology, 33*(1), 83-90.
23. Ogbonnaya, E., Anthony, A., Monago, C., Comfort, B., & Chuka, D. (2010). Phytochemical Screening and Acute- and Organ- Toxicity Evaluation of *Telfairia occidentalis* Root Aqueous Extract on Normal Wistar Rats. *Research Journal of Pharmacognosy and Phytochemistry, 2*(5), 417-420.
24. Ogunmoyole, T., Oladele, F., Aderibigbe, A., & Johnson, O. (2019). Hepatotoxicity of *Telfaria occidentalis* root extracts on wistar albino rat. *Heliyon, 1-6*.

25. Ojo, N. A., Adawaren, E., Tijjani, M. B., Chiroma, M., Simon, J., Afisu, B., *et al.* (2012). Acute Toxicity and Effect of Ethanolic Extract of *Telfairia occidentalis* Leaves on Blood Glucose Level in Normal Rats. *Vom Journal of Veterinary Science* , 25-31.
26. Okokon, E., Dar Farooq, A., Choudhary, M. I., & Antia, B. (2012). Immunomodulatory, Anticancer and Anti-inflammatory Activities of *Telfairia occidentalis* Seed Extract and Fractions. *International Journal of Food Nutrition and Safety*, 2(2), 72-85.
27. Okonwu, K., Akonye, L., & Mensah, S. (2018). Nutritional Composition of *Telfairia occidentalis* Leaf Grown in Hydroponic and Geoponic Media. *J. Appl. Sci. Environ. Manage.*, 22(2), 259 – 265.
28. Olorunfemi, A. E., Munavvar, A. S., & Hassaan, A. R. (2014). A Review of the Pharmacological and Biological Activities of the Aerial Parts of *Telfairia occidentalis* Hook.f. (Cucurbitaceae). *Tropical Journal of Pharmaceutical Research* , 13(10), 1761-1769.
29. Orłowski, P., Zmigrodzka, M., Tomaszewska, E., Ranożek-Soliwoda, K., Czupryn, M., Antos-Bielska, M., *et al.* (2018). Tannic acid-modified silver nanoparticles for wound healing: the importance of size. *Int J Nanomedicine.*, 16(13), 991-1007.
30. Osonuga, O., Faponle, A., Ezima, E., Adenowo, T., & Adelegan, A. (2020). Effects of aqueous leaf extract of *Telfairia occidentalis* on haematological parameters and liver enzymes in male Wistar rats. *Annals of Health Research*, 44-50.
31. Reitman, S., & Frankel, S. A. (1957). colorimetric method for the determination of serum glutamic oxaloacetic and glutamic pyruvic transaminase. 25(59).
32. Saalu, L., Kpela, T., Benebo, A., Oyewopo, A., Anifowope, E., & Oguntola, J. (2010). The Dose-Dependent Testiculoprotective and Testiculotoxic Potentials of *Telfairia occidentalis* Hook f. Leaves Extract in Rat. *International Journal of Applied Research in Natural Product*, 3(3), 7-38.
33. Shivashankar, S., Murali, A., & Sangeetha, M. (2019). Molecular interaction of phytochemicals with snake venom: Phytochemicals of *Andrographis paniculata* inhibits phospholipase A2 of Russell's viper (*Daboia russelli*). *Biocatalysis and agricultural biotechnology*, 1-10.
34. Sofowora, A. (1993). *Medicinal Plants and Traditional Medicine in Africa*. (2nd Edition ed.). Ibadan: Spectrum Books, Ibadan, Nigeria.
35. Suleiman, Z., Salahuddeen, Y., & Sabiu, U. (2022). Effect Of Methanolic Leaves Extract of *Telfairia occidentalis* on 2,4-Dinitrophenylhydrazine Induced Anaemic Rats. *UMYU Journal of Microbiology Research*, 7(1), 66-69.
36. Thomas, N., Ernest, U. O., Nkoyo, N. I., Elvis, S., Chukwubuzor, O. U., Ayodele, U. C., *et al.* (2013). Effects of methanolic seed extract of *Telfairia occidentalis* on blood coagulation in Albino rats. *Nigerian Journal of Experimental and Clinical Biosciences*, 1((1&2)), 10-13.

37. Tietz, N. W. (1976). *Fundamental of Clinical Chemistry*. W. B. Saunders Company Philadelphia.
38. Trease, G., & Evans, W. (1989). *Trease and Evans Pharmacognosy*. (13th Edition ed.). London: Bailliere Tindall.
39. Udosen, I., & Osu, S. (2022). Phytochemistry, and Effects of *Telfairia occidentalis* Leaf Extracts on the Growth and. *Journal of Applied Science and Environmental Management*, 26(2), 317-322.
40. Ugwah, E. (2023). Phytochemical analysis of six anti-venom medicinal plants. *Journal of Medicinal Plants Studies* 2023, 11(3), 71-79.
41. Ukwuani-Kwaja, A., Sani, I., & Kindzeka, L. (2021). Acute and Subchronic Toxicity Studies of Methanol Leaves Extract of *Ficus platyphylla* on Albino Rats. *Drug Discovery*, 15(35), 115-121.
42. Yakubu, M., Bilbis, L., Lawal, M., & Akanji, M. (2003). Evaluation of selected parameters of rat liver and kidney function following repeated administration of yohimbine. *Biokemistri*, 15(2), 50-56.