

EFFECTS OF CURCUMIN ON COGNITIVE IMPAIRMENT IN AGED FEMALE WISTAR RATS SUBJECTED TO INTESTINAL ISCHEMIA-REPERFUSION

ABSTRACT

Cognitive deficit, more common with age, it affects memory and cognitive functions. Intestinal ischemia-reperfusion injury (IIRI) causes systemic inflammation that can damage the brain, particularly the hippocampus. Curcumin, an active ingredient found in turmeric have neuroprotective properties. The aim of this study is to investigate the effect of curcumin on intestinal ischemia reperfusion injury induced-cognitive deficit in aged female Wistar rats.

Forty female Wistar rats between 200-250 kg were used in this study, they were weighed and randomly divided into four groups n=10. The animals pretreated with 100mg/kg for twenty-one days prior to surgical induction of ischemia for two hours followed by reperfusion which lasted for twenty-one days. After twenty-one days, behavioral assessment was done to investigate the behavioral functions. Animals were sacrificed and the brain was harvested, the hippocampus was isolated for biochemical and histological assessment. Data were analyzed using one-way ANOVA followed by Tukey's post hoc test with statistical significance set at $p < 0.05$.

The result shows that there was decrease in ability of the rats to recognize novel objects in IIRI groups but was improved curcumin treated groups. There was a reduction in the neurotransmitter level (AChE, Serotonin and Dopamine) in IIRI groups which was reversed in the CUR+IIRI groups ($p < 0.001$). Inflammatory markers (TNF- α , MPO) were also significantly reduced in CUR+IIRI group when compared to IIRI group ($p < 0.001$). Histological assessment shows that curcumin preserved hippocampal architecture, whereby reducing IIRI-induced cognitive deficit in aged female Wistar rats. This study showed that curcumin demonstrates significant neuroprotective effects against IIRI-induced Cognitive deficit.

Keywords: Intestinal ischemia/reperfusion injury, Aging, Behavioral, Neurotransmitters, Hippocampus, Curcumin.

1.0 Introduction

Cognitive deficit is characterized by deficiencies in executive function, language, memory, attention, and visuospatial abilities. It can greatly affect capacity to execute daily activities and independence. It can range in severity from mild to severe [1]. As people age, cognitive deficiencies become more common [2]. About 15-20% of elderly female individual of 65 years of age and above suffer from mild cognitive impairment (MCI), which is frequently seen as a prelude to dementia [3]. A variety of conditions, including neurodegenerative diseases, vascular disorders, traumatic brain damage, infections, metabolic disorders, and psychiatric disorders, can result in cognitive loss [4].

A well-known model of systemic inflammatory response syndrome is intestinal ischemia-reperfusion (I/R) injury, which is the temporary occlusion of bloodflow to the intestine followed by re-establishment of the blood flow to the intestine [5, 6], this condition could occur as a consequence of various conditions like cardiac insufficiency, necrotizing enterocolitis, trauma, atherosclerotic thrombus, embolism, or surgical complications, ischemia in the intestines is a relatively common occurrence in humans [7, 8]. It has been shown by previous studies that gut microbiome (GM) takes part in the pathogenesis of neurodegenerative disorders such as

cognitive deficit, this is possible through a bidirectional communication between the gut and the brain known as the microbiota-gut-brain axis (MGB)[9, 10]. Obstruction in the composition and activities of the GM have been reported not only in animal model but also in human patients indicated with AD[11]. GM dysbiosis has been implicated in increased intestinal permeability, blood-brain barrier (BBB) impairment, neuroinflammation and the development of hallmarks of AD [12]. Damage to the intestinal mucosal epithelium and villi, significant impairs local microvasculature, increased vascular and mucosal permeability resulting in edema, leukocyte plugging of capillaries, and systemic sequelae such blood acidosis, sepsis, multiple organ damage (MODs), and systemic inflammatory responses (SIRs) are among the consequences of an I/R injury [13]. Increased brain permeability brought on by inflammation enables proinflammatory mediators to penetrate brain and interact directly with neural cell extracellular receptors [14]. This may cause cerebral inflammation, which in turn alters the hippocampus, the structure of the brain involved in learning, memory, and cognitive functions, ultimately resulting in cognitive deficit [15]. Ischemic events, such as stroke, can cause severe harm to the hippocampal region, resulting in the death of neurons and a decline in memory function. These events also disrupt blood flow, which further exacerbates neurodegeneration in the hippocampal region [16, 17]. Due to the hippocampus's ongoing battle to play a vital role in cognition, persons who are impacted frequently experience difficulties remembering recent experiences or picking up new knowledge [19]. MODS and localized damage to a number of essential organs, such as the kidney and liver, have been directly linked to systemic inflammatory diseases that can induce encephalopathic symptoms [19].

Furthermore, ischemia damage from vascular diseases might impair neural transmission and brain integrity [20]. Other causes that lead to cognitive impairment include oxidative stress, neurotransmitter imbalances, genetic factors, and inflammatory responses [21, 22, 23].

The hippocampus can be severely damaged by oxidative stress, apoptosis, and inflammation, which can ultimately result in cognitive deficiencies [24]. Reactive oxygen species (ROS) generated due to excessive oxidative stress, which damages neurons, interferes with mitochondrial function, and reduces synaptic plasticity, a crucial component of memory formation [25]. In addition to decreasing cognitive capacities, oxidative damage can also cause apoptosis, or programmed cell death, which results in the loss of neurons and synaptic connections [26].

The brain's immune cells, known as microglia, are activated by chronic inflammation and generate inflammatory cytokines that damage neurons and compromise the blood-brain barrier [27]. According to Gomez-Arboleda *et al.* [28], this inflammation may cause excessive synaptic pruning as well as neurodegeneration. According to Escudero-Lourdes [29], oxidative stress, apoptosis, and inflammation increase the risk of neurodegenerative diseases, which includes cognitive impairment, by causing neuronal death, reduced synaptic function, and gradual cognitive decline.

Reduced physiological and biochemical organ functions are hallmarks of aging which is a biological transformation [30]. Additionally, it has a negative impact on cognitive processes, leading to a decline in coordination and locomotor activity as well as learning and memory impairment [31]. Numerous lines of evidence point to a progressive decline in learning and memory with age, and that this age-related deficit also affects activities involving spatial learning and memory. Numerous research [32, 33] have suggested that changes in the morphology and

function of the hippocampus formation are the cause of the age-related reduction in spatial learning ability.

Research has indicated that cognitive abilities in adult female rats are significantly reduced when compared to younger rats, as seen by observations made in a variety of maze models, including the Morris water maze, T-Maze, and Y-Maze. Laboratory rats can be used to estimate human life expectancy, with 13.8 rat days equal to one human year [34, 35]. Rats live an estimated 2-3.5 years [36, 35].

Asian medicine has traditionally employed curcumin, a yellow pigment derived from the rhizome of the plant *Curcuma longa*, as a traditional therapeutic treatment to treat a variety of illnesses [37, 38]. Previous research on humans and laboratory animals has shown that curcumin has antioxidant, anti-inflammatory, antiapoptotic, antiproliferative, anticancer, antidepressant, immunomodulatory, and neuroprotective properties [39, 40, 41, 42]. According to epidemiologic data, supplementing with curcumin appears to reverse several forms of cognitive impairment in rats and mice models. Regular consumption of curcumin also improves cognitive performance in healthy aged adults [43, 44, 45]. Studies have investigated the effect of curcumin on neurological disorder such as AD, however, there are limited study addressing cognitive deficit, a major clinical feature of AD in aged individual hence, the need for this study.

Methodology

3.1 Experimental Animals

Forty (40) female Wistar rats (200-250g) were purchased from Animal house of Ladoke Akintola University of Technology (LAUTECH), Ogbomoso, Oyo State, Nigeria. Ogbomoso, Oyo State, Nigeriabefore the onset of the study. They were acclimatized for two weeks and kept throughout the experiment in well aerated plastic cages in the animal house (temperature 28-31⁰C; photoperiod: 12-h natural light and 12-h dark; humidity:50-55%) of Faculty of Basic Medical Sciences (FBMS), LAUTECH, were fed with pelletized feed obtained from commercial dealer in Ogbomoso and watered *ad libitum*. The animal handling procedure was carried out in accordance with guidelines for the use and care of laboratory animals approved by LAUTECH's animal care and use research ethical committee.

3.2 Drugs and Reagents

All drugs and reagents used were of high analytical grade. D-galactose (Pan ReacAppliChem, Castellar del Vallès, Barcelona, Spain. Product code: A11311), curcumin (Cenik Chemicals, Sully Penarth, United Kingdom). Procaine penicillin (Anhui Chengushi Pharmaceutical CO., Ltd. China), Ketamine (Swiss Parenterals Ltd. India), Xylazine (BiovetaKomenskeho, Pharmaceutical, Czech Republic), normal saline, distilled water (Department of Pure and Applied Chemistry, LAUTECH, Oyo, Nigeria), buffered formalin (Department of Anatomy, FBMS, LAUTECH, Oyo, Nigeria), phosphate buffer saline (Department of Science Laboratory Technology, LAUTECH, Oyo, Nigeria).

3.3 Materials

Plastic cages, feeds, wood shavings, drinkers and feeders, weighing scale, surgical gloves, latex gloves, needle holder, universal bottles, plain bottles, EDTA bottles, razor blade, ethanol, sanitizer, methylated spirit, tissue paper, cotton wool, dissecting board, dissecting kits, 2.0

chromic sutures, sterile infusion set (tourniquet), paper tape, centrifuge, sensitive scale, incision set, ruler, normal saline, paper tapes, permanent markers, homogenizer, foil paper, Pasteur pipette, Syringes (insulin, 2ml and 5ml), Eppendorf bottles, methylated spirit, stop watch,

3.4 Experimental Design

Forty (40) female Wistar rats were divided into four groups of ten (10) animals each: **Group 1**; Curcumin (CUR), **Group 2**; Sham operated (SO), **Group 3**; Intestinal ischemia-reperfusion injury (IIRI) and **Group 4**; Curcumin and Intestinal ischemia-reperfusion injury (IIRI) (CUR + IIRI).

The treatments were given before the induction of IIRI. Intestinal ischemia was induced for 90 minutes while intestinal reperfusion was maintained for 72 hours[46].

The groups and their treatments are described below.

Group 1: Curcumin: Rats were treated with curcumin at the dosage of 100mg/kg [47].

Group 2: Sham Operated (SO); Rats were subjected to mid-line abdominal incision only involving mobilizations and replacement of intestine into the abdomen.

Group 3: Intestinal Ischemia Reperfusion injury (IIRI)

Group 4: Curcumin + Intestinal ischemia reperfusion injury (IIRI) (CUR + IIRI) group: Rats were treated with curcumin at the dosage of 100mg/kg orally for 21 days surgical induction of IIRI, after 21 days the animals were surgically induced with intestinal ischemia for 120 minutes and intestinal reperfusion was established after 90 minutes of ischemia, the animals were sacrificed 72 hours after reperfusion.

D-Galactose was administered intraperitoneally at 200 ml/kg for 8 weeks to induce aging in rats [48].

3.5 Surgical procedure for inducing intestinal ischemia reperfusion injury

- i. Animals were shaved at the abdominal region and cleaned with normal saline; then fasted for 12 hours before the surgical induction of Intestinal ischemia.
- ii. They were weighed and anaesthetized with Ketamine and Xylazine intraperitoneally (ketamine at 50mg/kg; xylazine at 10mg/kg) [49].
- iii. The animals were placed on a dissecting board and pinned down at the limbs.
- iv. A ventral midline laparotomy incision was made to open the abdominal cavity.
- v. Operating scissors was used to locate the small intestine and the superior mesenteric artery.
- vi. An infusion set of 5 cm was measured with a ruler and then cut to the appropriate size for measurement.
- vii. The intestine was lifted exposing the superior mesenteric artery: Some of the small intestine was measured from the proximal end of the ileocecal junction and 5cm to the distal end of the gut.
- viii. A small piece (3cm) was cut out from the infusion set (tourniquet), cut open, and wrapped on the measured part of the intestine and was sutured together lightly, taking care not to puncture the organ or any vessels.
- ix. Using the region wrapped with the infusion tube as a hand-hold, the gut was twisted clockwise 360 degrees.

- x. After twisting, the exposed gut was pushed back gently whilst ensuring that the twisted part is not loosened, and the abdominal region was closed with 2.0 chromic suture.

After 90 minutes ischemia, paleness of the intestine was observed.

The intestine is then untwisted to restore blood flow and animals were sacrificed after 72 hours[50].

3.6 Mode of Sacrifice, and Tissue Collection

3.6.1 Mode of Sacrifice

Seventy-two (72) hours after reperfusion, the animals were anesthetized using (ketamine at 50mg/kg; xylazine at 10mg/kg) [49].

Blood was collected through cardiac puncture with the use of 2ml syringe and needle and introduced into EDTA bottles.

3.6.2. Tissue Collection

- The hippocampus was harvested and cleared of adherent tissue.
- The harvested tissue was labelled appropriately and kept in universal bottles.
- The tissue was maintained at cold temperature after which it was homogenized

3.6.3. Preparation of Tissue homogenate

The hippocampal were harvested and cleared of adherent tissue. The harvested tissue was placed in a foil paper and weighed immediately with sensitive weighing scale (Lisay, China). The hippocampal tissue was washed in cold normal saline to remove blood stain. Some of the tissues were collected into sample bottles containing formaldehyde and labelled appropriately for histological examination. Other hippocampal and tissue were homogenized. For preparation of tissue homogenates, a specific weight of hippocampal tissue was homogenized in a universal bottle containing cold phosphate buffer ($p^H = 7.2$) in the ratio of 1:5. The homogenates were centrifuged at 3000 revolutions per minutes for 15 minutes with cold centrifuge. The supernatants were collected into Eppendorf bottles and refrigerated at $-4\text{ }^{\circ}\text{C}$ for further assays.

3.7 Behavioral evaluation (Novel objects recognition (NOR) test)

The novel object recognition test is a behavioral test for spatial working memory in rats. It consists of two sessions; first trial (T1) and the second trial (T2). The two trials were separated by an inter-trial interval of 2 h. In T1, each rat was put in the test box ($22.5 \times 24.5 \times 11.0\text{ cm}$), in which two identical objects (truncated cone: diameter, 4.5 and 6.0 cm; height, 3.5 cm) were placed at two adjacent corners, and the time spent exploring each object was measured for 5 min. During training, the animals explored between 40 and 60 % of the time on each object. Immediately after T1. In T2, a novel object replaced the object explored less by the mouse in T1, and the time spent exploring the familiar object (F) and the novel object (N) was recorded for 5 min. Objects were made of odorless plastic and similar in size. Between each trial, the objects and the field were cleaned. The total time spent sniffing or touching each object with the nose and/or forepaws were recorded. Recognition memory index was calculated by a discrimination index (%) using the method of Adeniyi *et al.* [51].

Memory Index (%) = $\frac{[(T_{\text{novel}} - T_{\text{familiar}})]}{(T_{\text{novel}} + T_{\text{familiar}})} \times 100$.

3.8 Biochemical evaluation

3.10.1 Determination of acetylcholinesterase (AChE) activity

The AChE enzymatic assay was determined using a modification of the spectrophotometric method of Ellman *et al.* [52] as previously described by Akinyemi *et al.* [53]. The reaction medium (2 mL final volume) contained 100 mmol/L of K⁺-phosphate buffer, pH 7.5, and 1 mmol/L of 5,5'-dithio-bis-nitrobenzoic acid. The method is based on the formation of the yellow anion, 5,5'-dithio-bis-acid-nitrobenzoic, measured by absorbance at 412 nm during 2-min incubation at 25 °C. The enzyme (40–50 µg of protein) was preincubated for 2 min. The reaction was initiated by adding 0.8 mmol/L of acetylthiocholine iodide. All samples were run in triplicate, and enzyme activity was expressed in micromole acetylthiocholine (ACh) hydrolysed per hour per milligram of protein. Serotone and dopamine were assayed using their respective colorimetric assay kit from bio vision and rat ELISA kit from Fine test with a catalog no: ERO 77. The manufacturers' procedures for assays were strictly followed.

Brain lipid peroxidation (MDA) level was estimated, brain superoxide dismutase activity level, brain reduced glutathione (GSH) level and brain catalase activity level. The extent of lipid peroxidation in the brain was determined quantitatively [54]. Briefly, the samples were mixed with 1 mL of 10 % trichloroacetic acid and 1 mL of 0.67 % thiobarbituric acid then heated in boiling water bath for 15 min after which butanol (2:1 v/v) was added to the solution. The amount of malondialdehyde (MDA) was measured by reaction with thiobarbituric (TBA) acid at 532 nm using UV Spectrophotometer. The values were calculated using the molar extinction coefficient of MDA-TBA adduct at 532 nm is 155 (mM⁻¹ cm⁻¹).

Superoxide Dismutase Activity (SOD) was assessed by Nitroblue Tetrazolium (NBT) method based on the principle that NBT undergoes photo-reduction (which is blue colored formazan) when exposed to light by superoxide radicals. It competes with the enzyme SOD for superoxide anions. With the presence of SOD in the reaction mixture, NBT produces a lesser quantity of coloured complex as compared to control.

Hippocampal tissue homogenate (500 µL) was mixed with chloroform (300 µL) and ethanol (500 µL). The mixture was centrifuged at 18,000 × g for 30 min. 50 µL of supernatant was mixed with 900 µL of SOD reagent (0.1 mmol/L xanthine, 0.1 mmol/L EDTA, 50 mg bovine serum albumin, 25 mmol/L NBT and 40 mmol/L Na₂CO₃) (pH 10.2). Further, twenty-five units of xanthine oxidase was added to the mixture and incubated for 20 min at 25 °C. The reaction was stopped by adding 1 mL of CuCl₂ (0.8 mmol/L) and absorbance was recorded at 560 nm [55].

The reduced glutathione (GSH) content in the mice brain was investigated spectrophotometrically at 412 nm. Briefly, in a test tube, the supernatant of the brain homogenate and trichloroacetic acid (10 % w/v) were mixed in 1:1 ratio and then the test tubes were centrifuged at 1000 × g for 10 min at 4 °C. The supernatant (0.5 mL) was mixed with 0.3 M disodium hydrogen phosphate (2 mL) and 0.25 mL of 0.001 M freshly prepared DTNB [5,5'-dithio-bis (2-nitrobenzoic acid) dissolved in 1 % w/v sodium citrate] was added and the absorbance was measured using a spectrophotometer at 412 nm. A standard curve was plotted using 10–100 µM of the reduced form of glutathione and the results were expressed as micromoles of reduced glutathione per mg of protein [56].

The activity of catalase enzyme was assessed through a spectrophotometer at 240 nm. Briefly, 1 mL of the brain homogenate was taken in a test tube and 1.9 mL of the phosphate buffer (50 mM, pH 7.4) was added. The reaction was initiated by the addition of 1 mL of 30 mM H₂O₂. The mixture of 2.9 mL of phosphate buffer and 1 mL of H₂O₂ without the brain homogenate was taken as blank. The decomposition of H₂O₂ resulted in the reduction of absorbance, which was recorded at 240 nm against the blank. The unit of catalase activity was expressed as the amount of enzyme that decomposes 1 μM of H₂O₂ per min at 25 °C using a molar coefficient of 43.6 M⁻¹ cm⁻¹ and the activity was expressed in terms of unit/mg proteins [56].

Hippocampal homogenates were used to assess superoxide dismutase (SOD) activity spectrophotometrically using the protocol of Obay et al. [57].

Measurement of nitric oxide (NO)

NO content in tissue homogenates was estimated in a medium containing 400 μL of 2 % vanadium chloride (VCl₃) in 5 % HCl, 200 μL of 0.1 % N-(1-naphthyl)ethylenediamine dihydrochloride, 200 μL of 2 % sulfanilamide (in 5 % HCl). After incubating at 37 °C for 60 min, nitrite levels, which corresponds to an estimate of levels of NO, were determined spectrophotometrically at 540 nm, based on the reduction of nitrate to nitrite by VCl₃. Hippocampal nitrite and nitrate levels were expressed as nanomole of NO/mg of protein.

Myeloperoxidase (MPO) activity was assessed as described by Pulli et al. [58]. TNF-α and Caspase 3 were also assessed.

Histological Analysis

The brains were stored in 10 % formaldehyde and fixed in methanol/chloroform/acetic acid solution (6:3:1) and dehydrated in ethanol. The dehydrated material was clarified with xylene and embedded in Paraplast. Then paraffin embedded coronal sections were cut (3 μm thickness) using a microtome and stained with hematoxylin (H) and eosin (E), after which it was mounted onto silane-coated slides for microscopic examination at 100x magnification.

3.11 Statistical analysis

Data were expressed as mean ± standard error of mean (Mean ± SEM). Analysis was performed with Graph Pad Prism, Version 7.0 (Graph Pad software, Inc., USA) was used to compare within group and Tukey's Post-test was used for multiple comparison. P-Values less than 0.05 were considered statistically significant.

3.0 Result

Results obtained from behavioral study (NOR) shows that there was a statistically significant decrease in both discrimination index and spatial short-term memory in intestinal ischemia reperfusion (IIRI) group when compared with Cur treated groups. In discrimination index, there is a significant decrease in IIRI when compared with CUR+IIRI (p<0.05). Spatial short-term memory shows that there was a statistically significant decrease in sham when compared with IIRI group (p<0.05), there was a statistically significant decrease in IIRI when compared with CUR+IIRI group (p<0.05) (Fig. 1A and B).

AChE, Serotonin and Dopamine were increased in CUR+IIRI groups when compared with IIRI group. There was a statistically significant increase in AChE level in sham group when compared with IIRI ($p < 0.05$), Also in CUR+IIRI group when compared with IIRI group ($p < 0.001$). There was a statistically significant increase in AChE level in sham group when compared with IIRI ($p < 0.05$). There was a statistically significant increase in Serotonin level in Sham when compared with IIRI group ($p < 0.001$) and CUR+IIRI when compared with IIRI group ($p < 0.001$). there was a statistically significant increase in dopamine level in CUR+IIRI when compared with IIRI ($P < 0.05$) (Fig. 2A, B and C).

The biochemical parameters obtained from the hippocampus includes CAT and SOD, the result shows that they were significantly decreased in IIRI group when compared with sham and CUR+IIRI group ($p < 0.001$) while MDA significantly increased in IIRI group when compared with CUR+IIRI group ($p < 0.001$). GSH shows no significant difference across the groups (Fig. 3A, B, C and D).

TNF- α , MPO and NO were significant increase in IIRI group when compared with CUR+IIRI groups. There was a statistically significant increase in TNF- α in IIRI when compared with CUR+IIRI ($p < 0.05$). MPO were decreased in sham and IIRI+CUR groups ($p < 0.05$; $p < 0.05$) when compared with IIRI ($p < 0.05$). NO shows that there was a statistically significant increase in IIRI when compared to sham and CUR+IIRI groups ($p < 0.001$; $p < 0.001$) (Fig. 4A, B and C).

Caspase 3 were increased sham groups when compared with IIRI group ($p < 0.01$). Also, there was a statistically significant increase in IIRI group when compared with CUR+IIRI group ($p < 0.05$) (Fig. 5A).

4.0 Discussion

The Novel Object Recognition (NOR) test is commonly used to assess cognitive functions, particularly recognition memory, which falls within the broader domain of cognitive abilities such as spatial and short-term memory. Intestinal ischemia-reperfusion injury (IIRI) is known to induce systemic inflammation and oxidative stress, which can extend to the brain, especially the hippocampus [59]. The hippocampus plays a critical role in memory and cognitive functions. Consequently, the damage caused by IIRI likely resulted in neuronal injury, synaptic dysfunction, and reduced neurogenesis, leading to impaired cognitive performance, as demonstrated by the low or negative discrimination index observed in the NOR test. The improved discrimination index in the CUR+IIRI group indicates that curcumin treatment alleviated the cognitive deficits caused by IIRI. Curcumin likely exerted neuroprotective effects on hippocampal neurons by counteracting the harmful consequences of inflammation and oxidative stress triggered by IIRI. Additionally, curcumin may have promoted synaptic plasticity and enhanced neurotransmitter systems involved in learning and memory, thus improving the processing and retention of new information [60].

Acetylcholinesterase (AChE), serotonin, and dopamine are key neurotransmitters involved in cognitive function, mood regulation, and memory [61]. Their balance and function are critical for maintaining normal cognitive processes, and disruptions in their levels led to significant impairments, as observed in intestinal ischemia-reperfusion injury (IIRI) [62]. AChE is responsible for breaking down acetylcholine, a neurotransmitter that facilitates learning and memory. Elevated AChE activity reduces acetylcholine levels, leading to impaired cholinergic transmission and cognitive dysfunction [63]. In the IIRI group, the decrease in AChE activity suggests that

cholinergic transmission was disrupted, likely due to oxidative stress and neuronal damage caused by IIRI. This impairment negatively affects cognitive functions such as learning and memory. However, curcumin, with its potent antioxidative and anti-inflammatory properties, likely helped restore neuronal integrity and normalize AChE activity [64]. By preserving acetylcholine levels and enhancing cholinergic signaling, curcumin supports cognitive function, particularly in memory and learning [65].

Serotonin, another crucial neurotransmitter, plays a significant role in mood regulation, cognition, and memory. A reduction in serotonin levels is often associated with mood disorders and cognitive deficits [66]. In the IIRI group, decreased serotonin levels can be attributed to oxidative stress and neuroinflammation, which damage serotonergic neurons and disrupt neurotransmitter synthesis and release [67]. This disruption contributes to both mood disturbances and cognitive decline. Curcumin's neuroprotective effects help counteract these detrimental processes, preserving serotonergic neurons and maintaining serotonin levels [68]. The increase in serotonin levels in the curcumin-treated IIRI group suggests that curcumin supports serotonergic function, which is essential for maintaining mood stability and cognitive performance.

Dopamine is similarly vital for cognitive processes, including motivation, reward, and attention [69]. Reduced dopamine levels are often linked to cognitive impairments and neurodegenerative conditions like Alzheimer's disease (AD) [70]. The drop in dopamine levels observed in the IIRI group is likely a result of oxidative stress and inflammation, which disrupt dopaminergic neurons and their signaling pathways [71]. This decrease can negatively impact motivation and other cognitive functions. Curcumin's antioxidative and anti-inflammatory properties help preserve dopaminergic neurons, maintaining dopamine levels and supporting cognitive health [72]. In the curcumin-treated group, the increase in dopamine levels highlights curcumin's role in protecting dopaminergic function, thereby aiding in cognitive processes such as attention and motivation. Curcumin exerts its neuroprotective effects by targeting multiple neurotransmitter systems, including acetylcholine, serotonin, and dopamine. Through its antioxidative and anti-inflammatory properties, curcumin helps preserve the integrity of cholinergic, serotonergic, and dopaminergic neurons, maintaining neurotransmitter balance and supporting cognitive functions such as learning, memory, and mood regulation. These effects suggest curcumin's potential therapeutic role in conditions like IIRI and neurodegenerative diseases, where cognitive function is compromised [73, 74].

Oxidative stress, driven by an imbalance between reactive oxygen species (ROS) and antioxidant defenses, is a key factor in the pathogenesis of numerous neurodegenerative diseases, including Alzheimer's Disease (AD), where cognitive decline is a primary symptom [75, 76]. In this study, intestinal ischemia-reperfusion injury (IIRI) was employed as a model to investigate oxidative stress and its effects on cognitive function, alongside the neuroprotective potential of curcumin, a polyphenol known for its strong antioxidant and anti-inflammatory properties [77]. The results demonstrated a significant increase in oxidative stress in the IIRI group, as evidenced by decreased levels of catalase (CAT) and superoxide dismutase (SOD), two key enzymatic antioxidants responsible for neutralizing harmful ROS. The decrease in these enzymes indicates that the antioxidant defense mechanisms were overwhelmed, leaving cells vulnerable to oxidative damage. This vulnerability was further supported by the elevated malondialdehyde (MDA) levels in the IIRI group, which reflected increased lipid peroxidation, a hallmark of oxidative damage to cellular

membranes. Curcumin treatment in the CUR+IIRI group led to a noticeable reversal of these oxidative stress markers. CAT and SOD levels increased, suggesting that curcumin mitigated oxidative damage by reducing ROS levels, thereby allowing these antioxidant enzymes to recover and function effectively. This protective effect of curcumin was further underscored by the significant reduction in MDA levels, indicating a lower rate of lipid peroxidation and, thus, less cellular membrane damage [78].

The stabilization of oxidative stress markers with curcumin treatment shows its potential to preserve neuronal health by preventing the cascade of oxidative damage that contributes to neuroinflammation and cognitive decline. Interestingly, while other markers of oxidative stress showed significant changes, reduced glutathione (GSH) levels remained relatively unchanged between the groups, suggesting that curcumin's antioxidant effects may target specific pathways rather than inducing a broad shift in overall antioxidant capacity. By reducing oxidative stress and lipid peroxidation, curcumin not only protects neuronal integrity but also helps prevent the progression of neuroinflammation, a key process involved in diseases like AD. These findings suggest that curcumin's antioxidant and anti-inflammatory properties make it a promising therapeutic candidate for mitigating cognitive decline associated with oxidative stress-related neurodegenerative diseases.

Tumor Necrosis Factor- α (TNF- α), myeloperoxidase (MPO), and nitric oxide (NO) levels indicate the relationship between neuroinflammation, oxidative stress, and cognitive function. These parameters are interconnected, as both TNF- α and MPO are key drivers of inflammatory and oxidative damage, while NO, though essential for normal signaling, exacerbates neurotoxicity when overproduced [79]. The increased levels of TNF- α , MPO, and NO in the intestinal ischemia-reperfusion injury (IIRI) group reflect a heightened inflammatory and oxidative response in the hippocampus, a brain region essential for learning and memory. TNF- α , a pro-inflammatory cytokine, amplifies neuroinflammation, contributing to neuronal damage and cognitive deficits, as seen in neurodegenerative conditions like Alzheimer's Disease (AD) [80, 81]. MPO, an enzyme that generates highly reactive oxidative species during inflammation, further worsens this damage by promoting oxidative stress [82]. NO, while necessary in small amounts for physiological processes, becomes harmful in excess, contributing to oxidative damage and neurotoxicity [83]. The combined elevation of these markers in the IIRI group underscores the widespread damage inflicted on the brain by inflammatory and oxidative processes. Curcumin's effects on these parameters in the CUR+IIRI group demonstrate its ability to disrupt this cascade of inflammation and oxidative stress. By reducing TNF- α levels, curcumin alleviates the inflammatory response, thereby limiting the harmful effects of chronic neuroinflammation on neuronal function and structure. The reduction in MPO levels indicates a decrease in oxidative stress, suggesting that curcumin protects neurons from oxidative damage that can impair cognitive functions. Finally, curcumin's ability to lower NO levels suggests its effectiveness in reducing neurotoxicity, further preserving neuronal health and preventing cognitive decline.

Taken together, the reduction of TNF- α , MPO, and NO in the CUR+IIRI group reflects curcumin's broad protective effects. Its antioxidative and anti-inflammatory properties act synergistically to mitigate the damaging effects of IIRI-induced neuroinflammation and oxidative stress, preserving hippocampal neurons and maintaining cognitive function. This multifaceted action makes curcumin

a promising therapeutic candidate for neurodegenerative diseases like AD, where inflammation and oxidative damage play central roles in disease progression and cognitive decline.

The blood-brain barrier (BBB) is one of the physiological sanctuaries of the body which protects the brain from harmful substances, when compromised, can lead to several pathological conditions [84]. Excessive nitric oxide (NO) can impair the BBB by forming peroxynitrite (ONOO⁻), a potent oxidant that damages endothelial cells and increases BBB permeability [85]. This damage allows inflammatory cytokines and oxidative agents to enter the brain, worsening neuroinflammation and oxidative stress, which contribute to cognitive deficits seen in conditions like Alzheimer's disease (AD) [86].

Intestinal ischemia-reperfusion injury (IIRI), due to inflammation and oxidative stress responses promotes disruption of BBB integrity. Elevated NO levels in the IIRI group indicate BBB disruption, which exacerbates neuroinflammation and neuronal damage. This observation is consistent with findings that traumatic injury or ischemia-reperfusion injury (IRI) can compromise the BBB and disrupt normal neuronal function, potentially leading to neurodegenerative diseases [87, 88]. Curcumin, known for its antioxidative and anti-inflammatory properties, helps restore BBB integrity by reducing NO levels and, consequently, the formation of peroxynitrite. This action protects BBB endothelial cells from oxidative damage, preventing the influx of harmful substances into the brain [89]. Studies have reported that curcumin can prevent BBB disruption through multiple mechanisms, such as inhibiting iNOS expression, reducing water content in the brain, preventing the absorbance of Evans blue dye after ischemia, and blocking NF- κ B activation [90, 91]. The ability of curcumin to inhibit NF- κ B activation, a key regulator of inflammatory responses, reducing the production of pro-inflammatory cytokines and oxidative stress markers may be attributed to the reason for observed reduction in inflammation in the curcumin treated group. By downregulating NF- κ B, curcumin maintains BBB integrity and protects against cognitive deficits.

Caspase-3 is a pivotal enzyme in the execution of apoptosis, responsible for cleaving various cellular components to facilitate cell death [93]. Increased caspase-3 expression is a hallmark of apoptotic processes and signifies significant neuronal loss [94]. Systemic inflammation and oxidative stress induced by IIRI results to neuronal apoptosis in the brain [95]. The elevated caspase-3 expression observed in the IIRI group highlights enhanced apoptotic activity in the hippocampus, a region critical for memory and learning. This increased apoptosis in the hippocampus can lead to substantial cognitive deficits, which is one of the characteristics of neurodegenerative diseases such as Alzheimer's Disease (AD) [96].

Curcumin, known for its strong antioxidative properties, helps neutralize reactive oxygen species (ROS) and mitigate oxidative stress [97]. By reducing oxidative damage, curcumin protects neurons from apoptosis [83]. Additionally, curcumin exhibits anti-inflammatory effects by lowering levels of pro-inflammatory cytokines such as TNF- α [98]. This reduction in inflammation decreases the signaling pathways that activate apoptotic processes, leading to lower caspase-3 expression [99]. Curcumin also modulates cell survival pathways, notably by activating the PI3K/Akt pathway, which enhances cell survival and inhibits apoptosis, thus decreasing caspase-3 activity [100]. Its ability to inhibit NF- κ B activation, a key regulator of inflammatory responses, further reduces the production of pro-inflammatory cytokines and subsequently diminishes the activation of apoptotic signaling cascades, including those involving caspase-3 [101]. Additionally, curcumin's

enhancement of survival pathways contributes to reduced caspase-3 expression by promoting cell survival and inhibiting apoptotic mechanisms [102, 103]. These may be the possible mechanisms by which curcumin suppress caspase 3 level in this study.

Histological examination of the hippocampus in the IIRI group reveals significant histopathological alterations, including the presence of apoptotic neurons and dystrophic changes. These findings indicate that IIRI induces considerable neuronal damage, likely due to oxidative stress and inflammation, leading to cell death and a loss of functional neurons in the hippocampus, which is crucial for cognitive function. These observations are consistent with the study of Oyeleke *et al.* [104], which also reported neurodegeneration due to amyloid beta injection. In disparity, the histological evaluation of the CUR+IIRI group shows reduced neuronal apoptosis and lack of dystrophic changes, suggesting that curcumin's anti-inflammatory and antioxidant properties mitigate the damage caused by oxidative stress and inflammation. This supports the protective role of curcumin in preserving neuronal integrity and cognitive function, consistent with findings by Yang *et al.* [59], who observed similar neuroprotective effects with melatonin in an IIRI model.

5. Conclusion

Intestinal ischemia-reperfusion injury (IIRI) leads to significant cognitive impairment as observed in the novel object recognition test, disrupted neurotransmitter balance, depleted antioxidant level, increased inflammation, apoptosis and damage to the hippocampus via activation of oxidative stress, inflammation and apoptotic pathways in the hippocampus. Curcumin, with its neuroprotective, antioxidant and anti-inflammatory properties, effectively mitigated IIRI induced cognitive impairment in female aged Wistar rats.

Limitation of the study

This study does not investigate social interaction and mood disorder which are also features of cognitive deficit. Therefore, further studies should investigate social cues in relation to IIRI-induced cognitive deficit.

Ethical Approval

Ethical approval was obtained from Faculty of Basic Medical Sciences, LAUTECH Ogbomosho with the approval number: ERCFBMSLAUTECH:056/08/2024.

Disclaimer (Artificial intelligence)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

References

1. Kirova, A. M., Bays, R. B., and Lagalwar, S. (2015). Working memory and executive function decline across normal aging, mild cognitive impairment, and Alzheimer's disease. *BioMed research international*, (1), 748212.

2. Murman, D. L. (2015). The impact of age on cognition. *In Seminars in hearing* (Vol. 36, No. 03, pp. 111-121). Thieme Medical Publishers.
3. Anderson, N. D. (2019). State of the science on mild cognitive impairment (MCI). *CNS spectrums*, 24(1), 78-87.
4. Brett, B. L., Gardner, R. C., Godbout, J., Dams-O'Connor, K., and Keene, C. D. (2022). Traumatic brain injury and risk of neurodegenerative disorder. *Biological psychiatry*, 91(5), 498-507.
5. Arumugam TV, Shiels IA, Woodruff TM, Granger DN, Taylor SM (2004): The role of the complement system in ischemia-reperfusion injury. *Shock* 21:401-409.
6. Arumugam TV, Magnus T, Woodruff TM, Proctor LM, Shiels IA, Taylor SM (2006). Complement mediators in ischemia-reperfusion injury. *Clin Chim Acta* 374:33-45.
7. Edwards M, Sidebotham D, Smith M, Leemput JV, Anderson B (2005). Diagnosis and outcome from suspected mesenteric ischaemia following cardiac surgery. *Anaesth Intensive Care* 33:210-217.
8. Oldenburg WA, Lau LL, Rodenberg TJ, Edmonds HJ, Burger CD (2004). Acute mesenteric ischemia-a clinical review. *Arch Intern Med* 164:1054-1062.
9. Krishaa, L., Ng, T. K. S., Wee, H. N., and Ching, J. (2023). Gut-brain axis through the lens of gut microbiota and their relationships with Alzheimer's disease pathology: review and recommendations. *Mechanisms of Ageing and Development*, 211, 111787.
10. Wei, W., Wang, S., Xu, C., Zhou, X., Lian, X., He, L., and Li, K. (2022). Gut microbiota, pathogenic proteins and neurodegenerative diseases. *Frontiers in Microbiology*, 13, 959856.
11. Bäuerl, C., Collado, M. C., Diaz Cuevas, A., Viña, J., and Pérez Martínez, G. (2018). Shifts in gut microbiota composition in an APP/PSS1 transgenic mouse model of Alzheimer's disease during lifespan. *Letters in applied microbiology*, 66(6), 464-471.
12. Giovannini, M. G., Lana, D., Traini, C., and Vannucchi, M. G. (2021). The microbiota–gut–brain axis and alzheimer disease. From dysbiosis to neurodegeneration: Focus on the central nervous system glial cells. *Journal of Clinical Medicine*, 10(11), 2358.
13. Hsieh, Y. H., McCartney, K., Moore, T. A., Thundyil, J., Gelderblom, M., Manzanero, S., and Arumugam, T. V. (2011). Intestinal ischemia-reperfusion injury leads to inflammatory changes in the brain. *Shock*, 36(4), 424-430.
14. Huang, J., Huang, N., Xu, S., Luo, Y., Li, Y., Jin, H., ... and Jin, F. (2021). Signaling mechanisms underlying inhibition of neuroinflammation by resveratrol in neurodegenerative diseases. *The Journal of nutritional biochemistry*, 88, 108552.
15. Skelly, D. T., Griffin, É. W., Murray, C. L., Harney, S., O'Boyle, C., Hennessy, E., ... and Cunningham, C. (2019). Acute transient cognitive dysfunction and acute brain injury

induced by systemic inflammation occur by dissociable IL-1-dependent mechanisms. *Molecular psychiatry*, 24(10), 1533-1548.

16. Farooqui, A. A. (2010). Neurochemical aspects of neurotraumatic and neurodegenerative diseases. *Springer Science and Business Media*.
17. Dhungana, H. (2014). Modelling of ischemic stroke: focus on co-morbidities and therapeutic intervention (Doctoral dissertation, Itä-Suomenyliopisto).
18. Lane, R. D., Ryan, L., Nadel, L., and Greenberg, L. (2015). Memory reconsolidation, emotional arousal, and the process of change in psychotherapy: New insights from brain science. *Behavioral and brain sciences*, 38, e1.
19. Zhou, J. (2023). Multiple Organ Dysfunction Syndrome. In *Explosive Blast Injuries: Principles and Practices* (pp. 265-279). *Singapore: Springer Nature Singapore*.
20. Levit, A. (2018). White Matter Inflammation And Executive Dysfunction: Implications For Alzheimer Disease And Vascular Cognitive Impairment (Doctoral dissertation, The University of Western Ontario (Canada)).
21. Rekatsina, M., Paladini, A., Piroli, A., Zis, P., Pergolizzi, J. V., and Varrassi, G. (2020). Pathophysiology and therapeutic perspectives of oxidative stress and neurodegenerative diseases: a narrative review. *Advances in therapy*, 37, 113-139.
22. Teleanu, D.M., Niculescu, A.G., Lungu, I.I., Radu, C.I., Vladăcenco, O., Roza, E., Costăchescu, B., Grumezescu, A.M. and Teleanu, R.I., (2022). An overview of oxidative stress, neuroinflammation, and neurodegenerative diseases. *International journal of molecular sciences*, 23(11), p.5938.
23. Leyane, T. S., Jere, S. W., and Houreld, N. N. (2022). Oxidative stress in ageing and chronic degenerative pathologies: molecular mechanisms involved in counteracting oxidative stress and chronic inflammation. *International journal of molecular sciences*, 23(13), 7273.
24. Gu, M., Mei, X.L. and Zhao, Y.N., (2021). Sepsis and cerebral dysfunction: BBB damage, neuroinflammation, oxidative stress, apoptosis and autophagy as key mediators and the potential therapeutic approaches. *Neurotoxicity research*, 39, pp.489-503.
25. Beckhauser, T. F., Francis-Oliveira, J., and De Pasquale, R. (2016). Reactive oxygen species: Physiological and physiopathological effects on synaptic plasticity: Supplementary issue: Brain plasticity and repair. *Journal of experimental neuroscience*, 10, JEN-S39887.
26. Reis, P. A., and Castro-Faria-Neto, H. C. (2022). Systemic response to infection induces long-term cognitive decline: neuroinflammation and oxidative stress as therapeutical targets. *Frontiers in Neuroscience*, 15, 742158.
27. Gullotta, G. S., Costantino, G., Sortino, M. A., and Spampinato, S. F. (2023). Microglia and the blood–brain barrier: An external player in acute and chronic neuroinflammatory conditions. *International Journal of Molecular Sciences*, 24(11), 9144.

28. Gomez-Arboledas, A., Acharya, M. M., and Tenner, A. J. (2021). The role of complement in synaptic pruning and neurodegeneration. *ImmunoTargets and Therapy*, 373-386.
29. Escudero-Lourdes, C. (2016). Toxicity mechanisms of arsenic that are shared with neurodegenerative diseases and cognitive impairment: Role of oxidative stress and inflammatory responses. *Neurotoxicology*, 53, 223-235.
30. Dharmarajan, T. S. (2021). Physiology of aging. *Geriatric gastroenterology*, 101-153.
31. Nakanishi, K., Sakakima, H., Norimatsu, K., Otsuka, S., Takada, S., Tani, A., and Kikuchi, K. (2021). Effect of low-intensity motor balance and coordination exercise on cognitive functions, hippocampal A β deposition, neuronal loss, neuroinflammation, and oxidative stress in a mouse model of Alzheimer's disease. *Experimental neurology*, 337, 113590.
32. Belviranl, M., Okudan, N., Atalık, K. E. N., and Öz, M. (2013). Curcumin improves spatial memory and decreases oxidative damage in aged female rats. *Biogerontology*, 14, 187-196.
33. Foster, T. C. (2012). Dissecting the age-related decline on spatial learning and memory tasks in rodent models: N-methyl-D-aspartate receptors and voltage-dependent Ca²⁺ channels in senescent synaptic plasticity. *Progress in neurobiology*, 96(3), 283-303.
34. Quinn R (2005). Comparing rat's to human's age: How old is my rat in people years? *Nutrition*.21:775-7.
35. Sengupta P (2012). A Scientific Review of Age Determination for a Laboratory Rat: How old is it in comparison with Human age? *Biomed Int*.2:81-9.
36. Pass D and Freeth G (1993). The rat. *Anzccart News*. 6:1-4.
37. Jovanovic SV, Boone CW, Steenken S, Trinoga M, Kaskey RB (2001). How curcumin works preferentially with water soluble antioxidants. *J Am Chem Soc* 123:3064-3068
38. Aggarwal BB, and Harikumar KB (2009) Potential therapeutic effects of curcumin, the anti-inflammatory agent, against neurodegenerative, cardiovascular, pulmonary, metabolic, autoimmune and neoplastic diseases. *Int J Biochem Cell Biol* 41:40-59
39. Xu Y, Ku BS, Yao HY, Lin YH, Ma X, Zhang YH, and Li XJ (2005). Antidepressant effects of curcumin in the forced swim test and olfactory bulbectomy models of depression in rats. *PharmacolBiochemBehav* 82:200-206
40. Bala K, Tripathy BC, Sharma D (2006) Neuroprotective and anti-ageing effects of curcumin in aged rat brain regions. *Biogerontology* 7:81-89
41. Jagetia GC, and Aggarwal BB (2007) "Spicing up" of the immune system by curcumin. *J Clin Immunol* 27:19-35
42. Zhao J, Zhao Y, Zheng W, Lu Y, Feng G, Yu S (2008) Neuroprotective effect of curcumin on transient focal cerebral ischemia in rats. *Brain Res* 1229:224-232

43. Ng TP, Chiam PC, Lee T, Chua HC, Lim L, Kua EH (2006) Curry consumption and cognitive function in the elderly. *Am J Epidemiol* 164:898–906
44. Reeta KH, Mehla J, and Gupta YK (2009) Curcumin is protective against phenytoin-induced cognitive impairment and oxidative stress in rats. *Brain Res* 1301:52–60
45. Tian M, Wang L, Yu G, Liu B, Li Y (2012) Curcumin preserves cognitive function and improve serum HDL in chronic cerebral hypoperfusion aging-rats. *Mol Neurodegener* 7:S3
46. Zhou, J., Huang, W.Q., Li, C., Wu, G.Y., Li, Y.S., Wen, S.H., Lei, W.L. and Liu, K.X., (2012). Intestinal ischemia/reperfusion enhances microglial activation and induces cerebral injury and memory dysfunction in rats. *Critical care medicine*, 40(8), pp.2438-2448.
47. Tanwar, V., Sachdeva, J., Kishore, K., Mittal, R., Nag, T.C., Ray, R., Kumari, S. and Arya, D.S., (2010). Dose-dependent actions of curcumin in experimentally induced myocardial necrosis: a biochemical, histopathological, and electron microscopic evidence. *Cell Biochemistry and Function: Cellular biochemistry and its modulation by active agents or disease*, 28(1), pp.74-82.
48. Wei, H., Li, L., Song, Q., Ai, H., Chu, J., and Li, W. (2005). Behavioural study of the D-galactose induced aging model in C57BL/6J mice. *Behavioural brain research*, 157(2), 245-251.
49. Lorenzini F, Tambara Filho R, Gomes RPX, Martino-Andrad AJ, Erdmann TR, Matias JE (2012). Long-term effects of the testicular torsion on the spermatogenesis of the contralateral testis and the preventive value of the twisted testis orchiepididymectomy. *Acta Cirurgica Brasileira*. 27:388-395.
50. Alabi, B., Iwalewa, O., Omobowale, T., Adedapo, A., Hammed, O., Ajike, R., and Afolabi, O. (2023). Cysteamine attenuate intestinal reperfusion injury induced by occlusion of mesenteric artery by enhancing intracellular thiol activities. *Drug Research*, 73(03), 137-145.
51. Adeniyi, I. A., Oregbesan, P. O., Adesanya, A., Olubori, M. A., Olayinka, G. S., Ajayi, A. M., and Onasanwo, S. A. (2024). Olaxsubscorpioidea prevented scopolamine-induced memory impairment through the prevention of oxido-inflammatory damage and modulation of cholinergic transmission. *Journal of Ethnopharmacology*, 318, 116995.
52. Ellman GL, Courtney KD, Andres V, Jr Featherstone RM (1961) A new and rapid colorimetric determination of acetylcholinesterase activity. *BiochemPharmacol* 7:88–95
53. Akinyemi AJ, Thome GR, Morsch VM, Stefanello N, da Costa P, Cardoso A, Goularte JF, Bello-Klein A, Akindahunsi AA, Oboh G, Schetinger MRC (2016) Effect of dietary supplementation of ginger and turmeric rhizomes on ectonucleotidases, adenosine de_aminase and acetylcholinesterase activities in synaptosomes from the cerebral cortex of hypertensive rats. *J Appl Biomed* 14:59–70

54. Wills, E. D (1966). Mechanism of lipid peroxide formation in animal tissues. *Biochem. J.*, 99, 667-676.
55. Pal, R., Nath, R., and Gill, K. D. (1993). Lipid Peroxidation and Antioxidant Defense Enzymes in Various Regions of Adult Rat Brain after Co-Exposure to Cadmium and Ethanol. *Pharmacology and Toxicology*, 73(4), 209–214.
56. Yadav, M., Parle, M., Sharma, N., Jindal, D.K., Bhidhasra, A., Dhingra, M.S., Kumar, A. and Dhingra, S., (2018). Protective effects of Spinacia oleracea seeds extract in an experimental model of schizophrenia: possible behavior, biochemical, neurochemical and cellular alterations. *Biomedicine & Pharmacotherapy*, 105, pp.1015-1025.
57. Obay, B. D., Taşdemir, E., Tümer, C., Bilgin, H. M., and Atmaca, M. (2008). Dose dependent effects of ghrelin on pentylene-tetrazole-induced oxidative stress in a rat seizure model. *Peptides*, 29(3), 448-455.
58. Pulli, B., Ali, M., Forghani, R., Schob, S., Hsieh, K.L., Wojtkiewicz, G., Linnoila, J.J. and Chen, J.W., 2013. Measuring myeloperoxidase activity in biological samples. *PLoS one*, 8(7), p.e67976.
59. Yang, B., Zhang, L.-Y., Chen, Y., Bai, Y.-P., Jia, J., Feng, J.-G., Zhou, J. (2020). Melatonin alleviates intestinal injury, neuroinflammation and cognitive dysfunction caused by intestinal ischemia/reperfusion. *International Immunopharmacology*, 85, 106596.
60. Nam, S.M., Choi, J.H., Yoo, D.Y., Kim, W., Jung, H.Y., Kim, J.W., Yoo, M., Lee, S., Kim, C.J., Yoon, Y.S. and Hwang, I.K., (2014). Effects of curcumin (*Curcuma longa*) on learning and spatial memory as well as cell proliferation and neuroblast differentiation in adult and aged mice by upregulating brain-derived neurotrophic factor and CREB signaling. *Journal of medicinal food*, 17(6), pp.641-649.
61. Walczak-Nowicka, Ł. J., and Herbet, M. (2021). Acetylcholinesterase inhibitors in the treatment of neurodegenerative diseases and the role of acetylcholinesterase in their pathogenesis. *International journal of molecular sciences*, 22(17), 9290.
62. Ucar, B. I., Ucar, G., Saha, S., Buttari, B., Profumo, E., and Saso, L. (2021). Pharmacological protection against ischemia-reperfusion injury by regulating the Nrf2-Keap1-ARE signaling pathway. *Antioxidants*, 10(6), 823.
63. Ozdemir, Z., Ozelik, A.B. and Uysal, M., (2019). Approaches based on cholinergic hypothesis and cholinesterase inhibitors in the treatment of Alzheimer's disease. *Frontiers in Clinical Drug Research-Alzheimer Disorders*, 8, pp.154-190.
64. Ashafaq, M., Hussain, S., Alshahrani, S., Siddiqui, R., Alam, M.I., Elhassan Taha, M.M., Almoshari, Y., Alqahtani, S.S., Jali, A.M. and Aljohani, H.M., (2023). Neuroprotective Effects of Nano-Curcumin against Cypermethrin Associated Oxidative Stress and Up-Regulation of Apoptotic and Inflammatory Gene Expression in Rat Brains. *Antioxidants*, 12(3), p.644.

65. Aggarwal BB, Sundaram C, Malani N, and Ichikawa H (2007). Curcumin: the Indian solid gold. *Adv Exp Med Biol.* 595:1–75.
66. Jenkins, T.A., Nguyen, J.C., Polglaze, K.E. and Bertrand, P.P., (2016). Influence of tryptophan and serotonin on mood and cognition with a possible role of the gut-brain axis. *Nutrients*, 8(1), p.56.
67. Muraleedharan, A. and Ray, S.K., (2024). Epigallocatechin-3-Gallate and Genistein for Decreasing Gut Dysbiosis, Inhibiting Inflammasomes, and Aiding Autophagy in Alzheimer's Disease. *Brain Sciences*, 14(1), p.96.
68. Winiarska-Mieczan, A., Kwiecień, M., Jachimowicz-Rogowska, K., Donaldson, J., Tomaszewska, E. and Baranowska-Wójcik, E., (2023). Anti-Inflammatory, Antioxidant, and Neuroprotective Effects of Polyphenols Polyphenols as an Element of Diet Therapy in Depressive Disorders. *International Journal of Molecular Sciences*, 24(3), p.2258.
69. Aarts, E., van Holstein, M., Hoogman, M., Onnink, M., Kan, C., Franke, B., Buitelaar, J. and Cools, R., (2015). Reward modulation of cognitive function in adult attention-deficit/hyperactivity disorder: a pilot study on the role of striatal dopamine. *Behavioural pharmacology*, 26(1 and 2-Special Issue), pp.227-240.
70. Klein, M.O., Battagello, D.S., Cardoso, A.R., Hauser, D.N., Bittencourt, J.C. and Correa, R.G., (2019). Dopamine: functions, signaling, and association with neurological diseases. *Cellular and molecular neurobiology*, 39(1), pp.31-59.
71. Sivandzade, F., Prasad, S., Bhalerao, A. and Cucullo, L., (2019). NRF2 and NF-κB interplay in cerebrovascular and neurodegenerative disorders: molecular mechanisms and possible therapeutic approaches. *Redox biology*, 21, p.101059.
72. Bhowmick, S., Sarkar, M., Hussain, J., Hassan, M., Basunia, M., Nahar, T., Rahman, A., Uddin, B. and Hossain, S., (2021). Curcuma longa extract ameliorates motor and cognitive deficits of 6-hydroxydopamine-infused Parkinson's disease model rats. *Advances in Traditional Medicine*, pp.1-15.
73. Sharma, N., Sharma, S., and Nehru, B. (2017). Curcumin protects dopaminergic neurons against inflammation-mediated damage and improves motor dysfunction induced by single intranigral lipopolysaccharide injection. *Inflammopharmacology*, 25, 351-368.
74. Khayatan, D., Razavi, S.M., Arab, Z.N., Hosseini, Y., Niknejad, A., Momtaz, S., Abdolghaffari, A.H., Sathyapalan, T., Jamialahmadi, T., Kesharwani, P. and Sahebkar, A., (2024). Superoxide dismutase: a key target for the neuroprotective effects of curcumin. *Molecular and Cellular Biochemistry*, 479(3), pp.693-705.
75. Bai, R., Guo, J., Ye, X. Y., Xie, Y., and Xie, T. (2022). Oxidative stress: The core pathogenesis and mechanism of Alzheimer's disease. *Ageing research reviews*, 77, 101619.

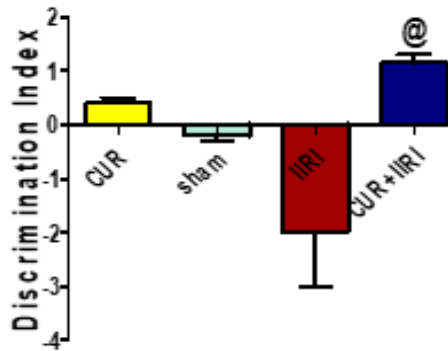
76. Ekundayo, B.E., Obafemi, T.O., Adewale, O.B., Obafemi, B.A., Oyinloye, B.E. and Ekundayo, S.K., (2024). Oxidative Stress, Endoplasmic Reticulum Stress and Apoptosis in the Pathology of Alzheimer's Disease. *Cell Biochemistry and Biophysics*, pp.1-21.
77. Tsai Y-M, Chien C-F, Lin L-C, and Tsai T-H (2011). Curcumin and its nano-formulation: the kinetics of tissue distribution and bloodbrain barrier penetration. *Int J Pharm.* 416:331–338.
78. Khater, S.I., Dowidar, M.F., Abdel-Aziz, A.E., Khamis, T., Dahran, N., Alqahtani, L.S., Metwally, M.M., Al-Hady Abd-Elrahman, A.S., Alsieni, M., Alosaimi, M.E. and Abduljabbar, M.H., (2022). β -Cell autophagy pathway and endoplasmic reticulum stress regulating-role of liposomal curcumin in experimental diabetes mellitus: a molecular and morphometric study. *Antioxidants*, 11(12), p.2400.
79. Tewari, D., Sah, A.N., Bawari, S., Nabavi, S.F., Dehpour, A.R., Shirooie, S., Braidy, N., Fiebich, B.L., Vacca, R.A. and Nabavi, S.M., (2021). Role of nitric oxide in neurodegeneration: function, regulation, and inhibition. *Current neuropharmacology*, 19(2), pp.114-126.
80. Montgomery, S. L., and Bowers, W. J. (2012). Tumor necrosis factor-alpha and the roles it plays in homeostatic and degenerative processes within the central nervous system. *Journal of neuroimmune pharmacology*, 7(1), 42-59.
81. Onyango, I. G., Jauregui, G. V., Čarná, M., Bennett Jr, J. P., and Stokin, G. B. (2021). Neuroinflammation in Alzheimer's disease. *Biomedicines*, 9(5), 524.
82. Chen, S., Chen, H., Du, Q., and Shen, J. (2020). Targeting myeloperoxidase (MPO) mediated oxidative stress and inflammation for reducing brain ischemia injury: Potential application of natural compounds. *Frontiers in physiology*, 11, 433.
83. Tiwari, V., and Chopra, K. (2012). Attenuation of oxidative stress, neuroinflammation, and apoptosis by curcumin prevents cognitive deficits in rats postnatally exposed to ethanol. *Psychopharmacology*, 224, 519-535.
84. Furtado, D., Björnalm, M., Ayton, S., Bush, A. I., Kempe, K., and Caruso, F. (2018). Overcoming the blood–brain barrier: the role of nanomaterials in treating neurological diseases. *Advanced materials*, 30(46), 1801362.
85. Sanchez-Cano, F., Hernández-Kelly, L. C., and Ortega, A. (2021). The blood–brain barrier: much more than a selective access to the brain. *Neurotoxicity Research*, 39(6), 2154-2174.
86. Shandilya, S., Kumar, S., Jha, N. K., Kesari, K. K., and Ruokolainen, J. (2022). Interplay of gut microbiota and oxidative stress: Perspective on neurodegeneration and neuroprotection. *Journal of Advanced Research*, 38, 223-244.
87. Hausburg, M.A., Banton, K.L., Roman, P.E., Salgado, F., Baek, P., Waxman, M.J., Tanner II, A., Yoder, J. and Bar-Or, D., (2020). Effects of propofol on ischemia-reperfusion and traumatic brain injury. *Journal of Critical Care*, 56, pp.281-287.

88. Khan, M., Dhammu, T. S., Sakakima, H., Shunmugavel, A., Gilg, A. G., Singh, A. K., and Singh, I. (2012). The inhibitory effect of S-nitrosoglutathione on blood–brain barrier disruption and peroxynitrite formation in a rat model of experimental stroke. *Journal of neurochemistry*, 123, 86-97.
89. Pan MH, Lin-Shiau SY, and Lin JK (2000) Comparative studies on the suppression of nitric oxide synthase by curcumin and its hydrogenated metabolites through down-regulation of IκB kinase and NFκB activation in macrophages. *BiochemPharmacol* 60(11):1665–1676.
90. Ferri KF, and Kroemer G (2001) Organelle-specific initiation of cell death pathways. *Nat Cell Biol* 3(11):E255.
91. Jiang C, Li G, Huang P, Liu Z, and Zhao B (2017). The gut microbiota and Alzheimer's disease. *J Alzheimers Dis*, 58: 1–15
92. Zhu, H., Zhang, H., Hou, B., Xu, B., Ji, L., and Wu, Y. (2022). Curcumin Regulates Gut Microbiota and Exerts a Neuroprotective Effect in the MPTP Model of Parkinson's Disease. *Evidence-Based Complementary and Alternative Medicine*.
93. Lossi, L., Castagna, C., and Merighi, A. (2018). Caspase-3 mediated cell death in the normal development of the mammalian cerebellum. *International journal of molecular sciences*, 19(12), 3999.
94. Snigdha, S., Smith, E. D., Prieto, G. A., and Cotman, C. W. (2012). Caspase-3 activation as a bifurcation point between plasticity and cell death. *Neuroscience bulletin*, 28, 14-24.
95. Cakir, E., Cakir, U., Tayman, C., Turkmenoglu, T. T., Gonel, A., and Turan, I. O. (2020). Favorable effects of astaxanthin on brain damage due to ischemia-reperfusion injury. *Combinatorial Chemistry & High Throughput Screening*, 23(3), 214-224.
96. Wang, S., Sun-Waterhouse, D., Waterhouse, G. I. N., Zheng, L., Su, G., and Zhao, M. (2021). Effects of food-derived bioactive peptides on cognitive deficits and memory decline in neurodegenerative diseases: A review. *Trends in Food Science and Technology*, 116, 712-732.
97. Singh, P., Singh, S., Garg, G., Singh, A. K., and Rizvi, S. I. (2020). Curcumin has protective effects on ROS production and redox imbalance in an experimental oxidative-stressed model of rat. *Journal of Biologically Active Products from Nature*, 10(6), 484-494.
98. Makuch, S., Więcek, K., and Woźniak, M. (2021). The immunomodulatory and anti-inflammatory effect of curcumin on immune cell populations, cytokines, and in vivo models of rheumatoid arthritis. *Pharmaceuticals*, 14(4), 309.
99. Arjumand, W., Seth, A., and Sultana, S. (2011). Rutin attenuates cisplatin induced renal inflammation and apoptosis by reducing NFκB, TNF-α and caspase-3 expression in wistar rats. *Food and chemical toxicology*, 49(9), 2013-2021.
100. Wang, M., Jiang, S., Zhou, L., Yu, F., Ding, H., Li, P., Zhou, M. and Wang, K., (2019). Potential mechanisms of action of curcumin for cancer prevention: focus on cellular

- signaling pathways and miRNAs. *International journal of biological sciences*, 15(6), p.1200.
101. Patel, S. S., Acharya, A., Ray, R. S., Agrawal, R., Raghuwanshi, R., and Jain, P. (2020). Cellular and molecular mechanisms of curcumin in prevention and treatment of disease. *Critical reviews in food science and nutrition*, 60(6), 887-939.
 102. Fiorillo, C., Becatti, M., Pensalfini, A., Cecchi, C., Lanzilao, L., Donzelli, G., Nassi, N., Giannini, L., Borchini, E. and Nassi, P., (2008). Curcumin protects cardiac cells against ischemia-reperfusion injury: effects on oxidative stress, NF- κ B, and JNK pathways. *Free Radical Biology and Medicine*, 45(6), pp.839-846.
 103. Jin, H., Qiao, F., Wang, Y., Xu, Y., and Shang, Y. (2015). Curcumin inhibits cell proliferation and induces apoptosis of human non-small cell lung cancer cells through the upregulation of miR-192-5p and suppression of PI3K/Akt signaling pathway. *Oncology reports*, 34(5), 2782-2789.
 104. Oyeleke, M. B., Oni, H. T., Arokoyo, O. L., and Owoyele, B. V. (2022). Therapeutic effects of crude extracts of *Bacopa floribunda* on beta-amyloid 1-42-induced Alzheimer's disease via suppression of dyslipidemia, systemic inflammation and oxidative stress in male Wistar Rats. *Heliyon*, 8(4).

1. Behavioral Test (Novel Object Recognition (NOR) Test)

Fig. 1A



1B

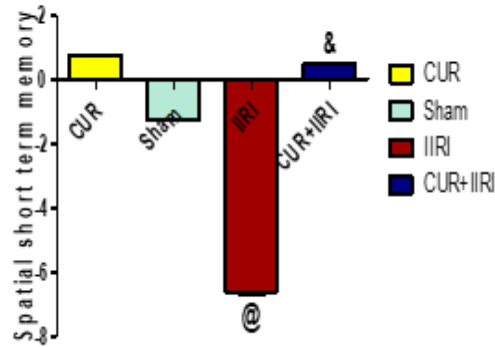


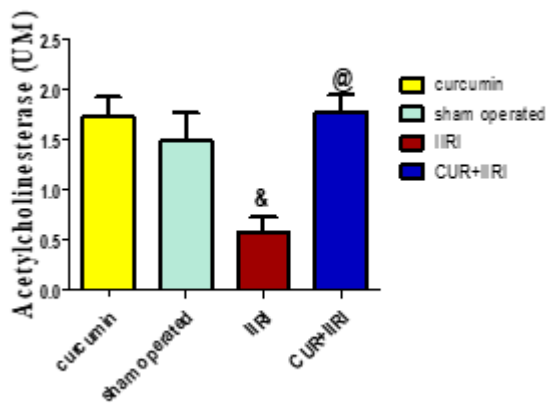
Fig. 1A and B: Effects of curcumin on intestinal ischemia reperfusion injury induced-cognitive deficit on discrimination index and spatial short term memory in aged female Wistar rats.

@ represents significance at (P<0.05) vs IIRI group.

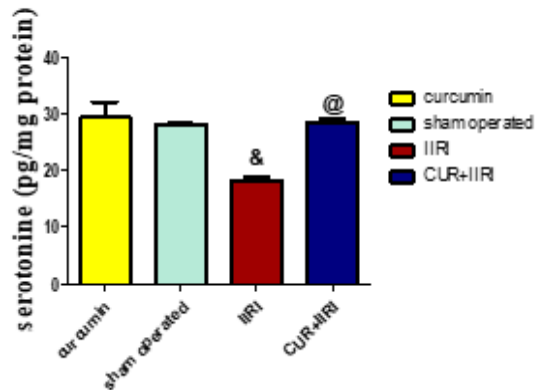
& represents significance at (P<0.001) vs IIRI group. @ represents significance at (p<0.001) vs sham group.

2. Brain Neurotransmitters

2A



B



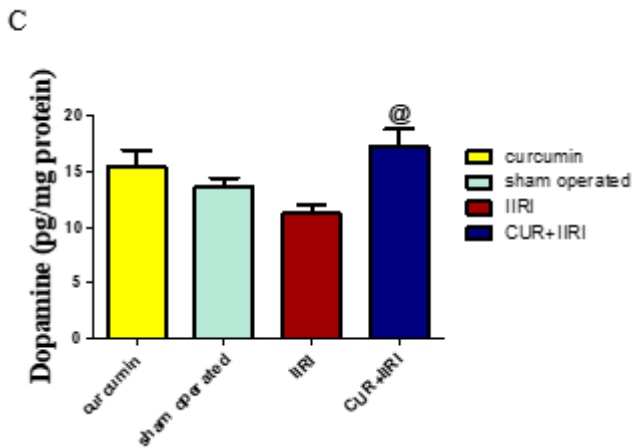


Fig 2. Fig. 2A, B and C: Effects of curcumin on intestinal ischemia reperfusion injury induced-cognitive deficit on acetylcholinesterase, serotonin and dopamine in aged female Wistar rats.

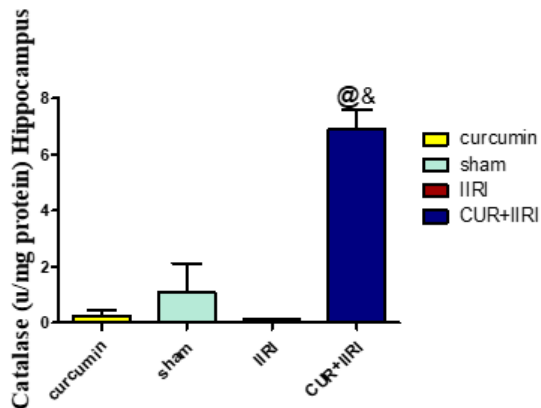
[@]represents significance at ($P < 0.05$) vs IIRI group. [&]represents significance at ($p < 0.01$) vs sham group.

[@]represents significance at ($P < 0.001$) vs sham group. [&]represents significance at ($p < 0.001$) vs IIRI group.

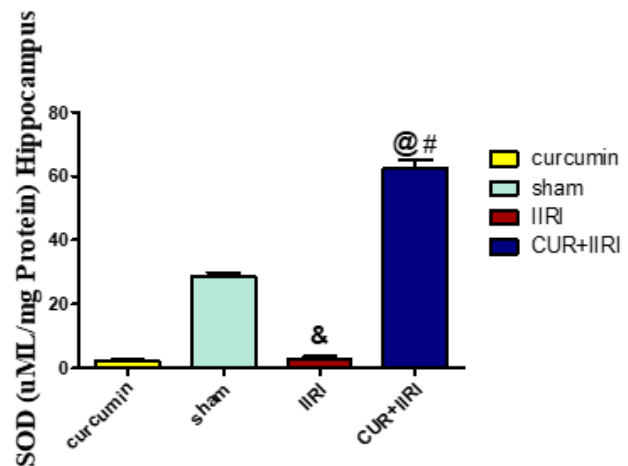
[@]represents significance at ($P < 0.05$) vs IIRI group.

3. Antioxidants parameters

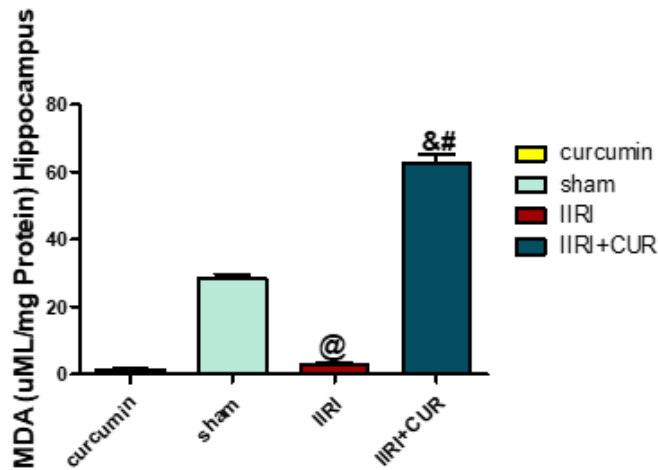
A



B



C



D

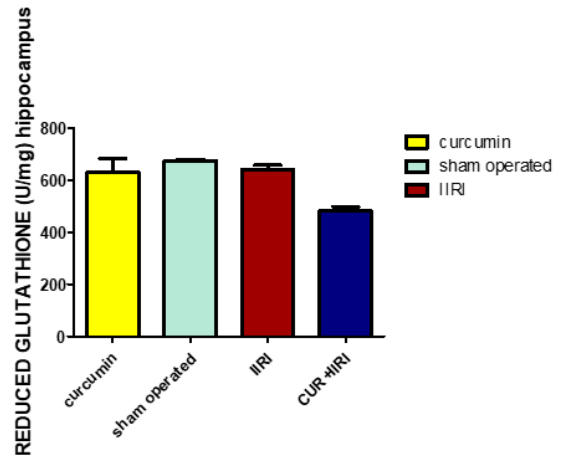


Fig. 3A, B, C and D: Effects of curcumin on intestinal ischemia reperfusion injury induced-cognitive deficit on CAT, SOD, MDA and GSH in aged female Wistar rats.

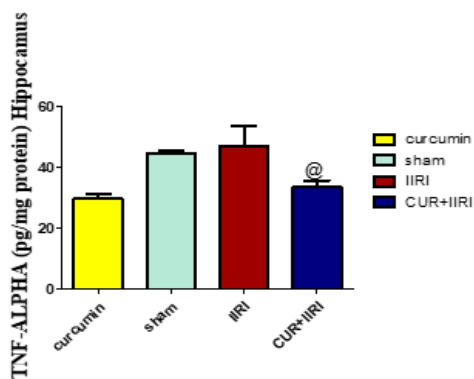
@ represents significance at (P<0.001) vs IIRI group. & represents significance at (p<0.001) vs sham group.

@ represents significance at (P<0.001) vs IIRI group. # represents significance at (p<0.01) vs sham group. & represents significance at (p<0.001) vs sham.

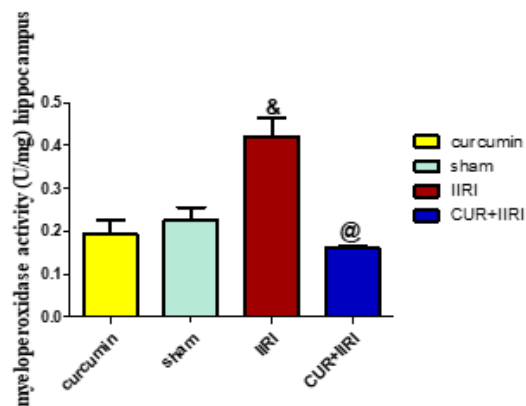
@ represents significance at (P<0.001) vs sham group. & represents significance at (p<0.001) vs IIRI group.

4. Inflammatory Markers

A



B



C

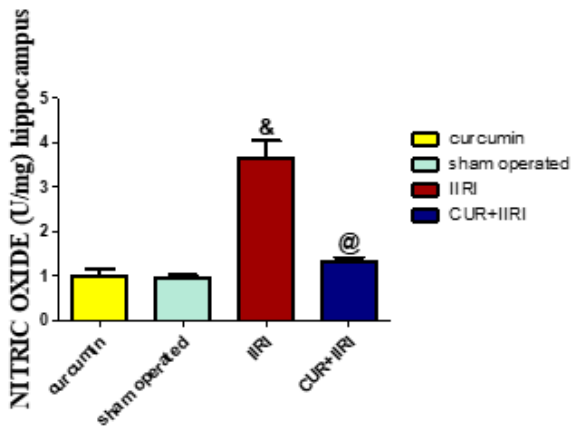


Fig. 4A, B, and C: Effects of curcumin on intestinal ischemia reperfusion injury induced-cognitive deficit on TNF- α , MPO and NO in aged female Wistar rats.

@ represents significance at (P<0.05) vs IIRI group.

@ represents significance at (P<0.001) vs IIRI group. & represents significance at (p<0.001) vs sham group.

@ represents significance at (P<0.001) vs IIRI group. & represents significance at (p<0.001) vs sham group.

5. Apoptotic Markers

A

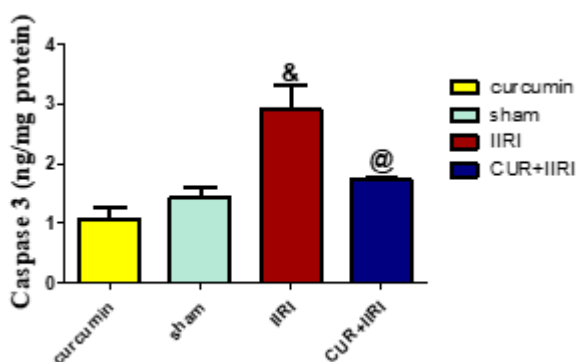
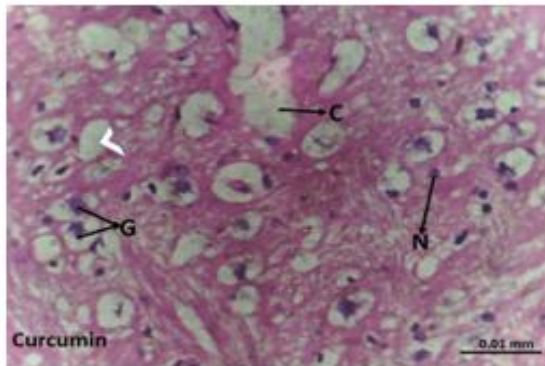


Fig. 5A: Effects of curcumin on intestinal ischemia reperfusion injury induced-cognitive deficit on caspase 3 in aged female Wistar rats.

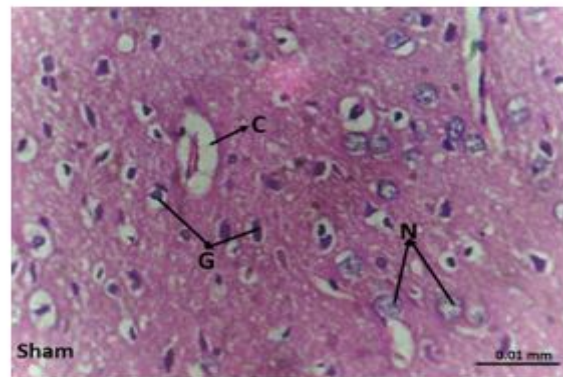
& represents significance at (p<0.001) vs sham group.

@ represents significance at (P<0.001) vs IIRI group.

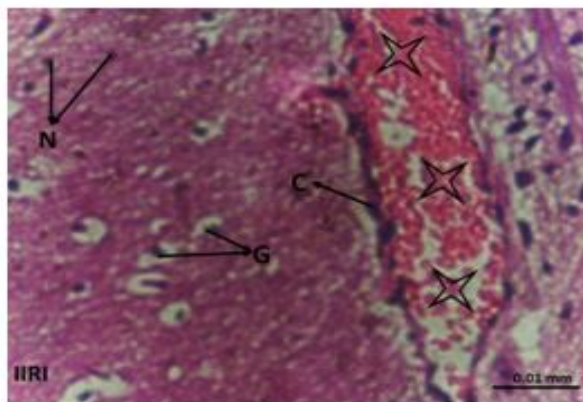
6. Histology (H&E)



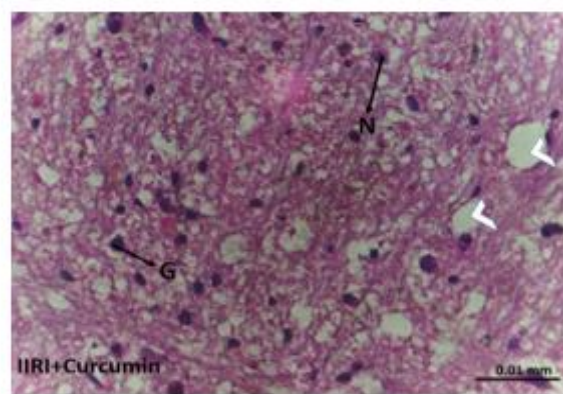
Curcumin (X100)



Sham (100)



IIRI (100)



CUR+IIRI (100)

Fig. 6: Effects of curcumin on intestinal ischemia reperfusion injury induced-cognitive deficit on histology of the hippocampus in aged female Wistar rats.

N represents Apoptotic neurons; G represents Glial cells and C represents capillary.