Review Article

An overview on Amaranthuscruentus L.:Alternative source of crop, its economical uses and associated beneficial microbes

ABSTRACT

Amaranthus cruentus L.is a medicinally important locally available tropical plant belonging to the family Amaranthaceae. In present scenario, this plant is designated as 'alternative source of crop' or 'Pseudocereal' having various nutritional properties (Protein, carbohydrate, fatty acid, Vitamins, Minerals). Moreover, this plant has various medicinal properties also i.e. antioxidant, antihypertensive, antidiabetic, immune stimulation, anthelmintic, hepatoprotective, antimalarial etc. This plant has Worldwide distribution and short growing period of time. It is also regarded as Fodder crop and other industrial uses. The microbes associated with this plant help to protect the host plants from both pathogenic microorganisms and adverse climatic conditions by secreting different types of bioactive secondary metabolites i.e. phenolics, flavonoids, terpenoids, terpenes, tannins, alkaloids, quinones, anthocyanin, saponin, steroids and promote growth and development of the plant. Some unique features of this plant are also descried here. This review intends to expand the understanding of taxonomic characteristics of this plant, economic uses, associated beneficial microbes and their application for sustainability of this alternative source of crop. The cultivation of A. cruentus possess a good attraction among both the researchers and farmers and cultivation procedure is more easy than other species. Some species of this genus are not cultivated yet, they are under process. Additionally, this species is more adorably cultivated due to the high nutritional value of its seeds.

Keywords: Amaranthus cruentus, alternative source of crop, secondary metabolites, microbes.

1.INTRODUCTION

Amaranthus mainly consists of 400 species, which are under cultivation. But, the total number of species under this genus are needed to be determined. The genus includes leafy vegetable amaranth, grain amaranth, ornamental, and weedy amaranth. Three major seed amaranth types: A. caudatus L., A. hypochondriacus L., and A. cruentus L., with A. tricolor grouped as an ornamental. A. blitum (slender amaranth), A. lividus, A. dubius (spleen amaranth), A. spinosus, A. thunbergia, and A. graecizans are produced for their leaves (leafy amaranth). Typical types common in Sub-Sahara are A. graezianus & thumbergii (Mukuwapasi et al., 2024).

Amaranthuscruentus is a tropical, endemic, invasive, short lived perennial herbaceous plant in Africa and South America (Gresta et al., 2020). Nevertheless, now this plant is distributed all over the world and contains approximately 70 species (Gresta et al., 2020). It is cultivated mainly in tropical and temperate region (Letchamo et al., 2017). This species has greater variability (Ma et al., 2021; Gins et al., 2024). Phenotypic plasticity, Introgression and hybridization are profoundly observed in this species (Gresta et al., 2020; Prajitha&Thoppil, 2018; Jimohetal., 2022; Amosova et al., 2024). To protect from pathogenic microorganism, this plant has some advanced physiological features (Jimohetal., 2022). In Africa, this plant is recognized as source of food and constructive and traditional medicine (Letchamo et al., 2017). In present scenario, food scarcity is a major problem and food security is necessary to protect the future generations. So, this plant is used as 'alternative source of crop' and also considered as 'Pseudocereals' (Fletcher, 2015; Mburu, 2011; Gresta et al., 2020). Moreover, this plant is cultivated for using both the Grains and leaves in Africa and Asia (Hoidal et al., 2019). The cultivation procedure is followed by firstly, 2-4 inches shoot with 2-4 leaves cut and then they are deeply buried into the soil and regularly watered. When the root is optimum for its growth and when the plants are 14-20 days old, the leaves are harvested (Achigan et al., 2014). It has various medicinal principles i.e. antioxidant, anti-diabetic, anti-inflammatory properties, antihypertensive properties, stimulation of immune system, anthelmintic properties (Torane et al., 2017) etc. Presence of 3.1% phenolics and 11.6% flavonoid contribute to the antioxidant and anti-inflammatory property to scavenge the free radicles and protect from Reactive Oxygenic Species (ROS) (Ali, 2021).3.6% alkaloids present in it possessing antimicrobial property and antidiabetic property is shown due to the presence of 1.3% saponin. (Ali, 2021). This plant harbor endophytic and others microflora. Endophytic

microbes secrete active secondary metabolites i.e. phenolics, flavonoids, terpenoids, terpenes, tannins, alkaloids, quinones, anthocyanin, saponin, steroid etc. These metabolites are potential agents for antimicrobial, anticancerous and many other therapeutic properties which are beneficial to protect the host plant from both biotic and abiotic stresses (Mousavi&Karami,2022). This plant is an example of C4 photosynthetic ones (Achigan et al., 2014, Gins et al., 2024) and reported as highwater using efficiency (WUE) (Wolosik&Markowska,2019) and grow in any types of soil i.e. acidic, alkaline, saline or nutrient-poor soil (Park et al., 2014). This species is highly resistant to drought (Gresta et al., 2020; Achigan et al., 2014) and salinity(Gins et al., 2024; Emam et al., 2024) heat, (Jimohetal., 2022). and UV stress radiation also(Castrillón&Frier, and;Omami&Hammes, 2006). It has very good nutritional properties as it contains vitamins, proteins (Lysine, Tryptophan), unsaturated fatty acid, minerals, tocopherols and tocotrienol etc. and essential antioxidant properties (Wolosik&Markowska,2019). Its short growing period give expectancy and provide a source of employment to the farmers (Makinde&Ojeniyi, 2010). According to genomic studies, A. cruentus contains 17 chromosomes (Sammour&Mitra, 2012). The species are diploid (2n=32) sometimes chromosome no. varies (2n=34). So, the chromosome no. n=17 sometimes n=16 (Hricovaetal., 2016, Ma et al., 2021). Previous report shows that it is hybridized easily (Lanta&Ondrej,2003). In South Africa, it is considered as potential alternative cereal having minerals, phytochemicals (Manyelo et al., 2022). Several species of Amaranthus have a role in CO₂ sequestration (Jimoh et al., 2022; Malik et al., 2023). This is short day plant and flowering takes place in 44 days and harvested in 102 days (Ogwu,2020). Microbes present in the plant significantly help the host to increase growth, protect from both biotic and abiotic stresses and also stimulate the medicinal activities without causing any disturbance or damage. In this review, taxonomic characteristics, secondary metabolites, economic importance of this plant and lastly the microbes (PGPR, PGPF, fungal and bacterial Endophytes and Mycorrhizae) associated with this plant and how they promote the plant growth is discussed here. In this review, we discussed mainly the beneficial features and also the exceptional features of this plant A. cruentus (Red Amaranth). It distributed all over the world and can grow any type of soils. This C4 photosynthetic plant can tolerate adverse climatic conditions. Short growing period are very advantageous for agriculture and farmers are also benefitted. As India and most of the countries are suffered from Malnutrition, this Red Amaranth provide high amount of nutrition (amino acid, carbohydrate, fatty acid, vitamins, minerals, fibres, antioxidant). Financial expenditure of the maintenance of this plant is comparatively low.

2.TAXONMIC DESCRIPTION OF AMARANTHUS CRUENTUS L.

2.1Taxonomic classification: (Wolosik&Markowska,2019)

Kingdom: Plantae

Subkingdom: Tracheobionta
Subdivision: Spermatophyta
Division: Magnoliophyta

Class: Magnoliopsida

Subclass: Caryophyllidae Order: Caryophyllales

Family: Amaranthaceae
Genus: Amaranthus

Species: Amaranthuscruentus L.

2.2Habit:

This plant is herbaceous, tropical and belongs to the family Amaranthaceae. The common name of the plant is red or blood amaranth. In Bengali, it is popularly known as 'Lal note sak'. It reproduces by seeds and have very short (4-6 weeks) growth period (Wolosik&Markowska,2019).

2.3 Phenological Characteristics:

One dominant large tap root is present which is soft, 6-7 cm long (Wolosik& Markowska, 2019; Mukuwapasi et al., 2024).

Stem is straight, branched, thick, ribbed, 60-65 cm in height, reddish in colour, surface minutely haired (Wolosik and Markowska, 2019).

Leaves are simple, reddish, 5-7.5 cm length and 2.1-4.2 cm breadth, margin entire, spiral arrangement, shape ovoid, exstipulate, leaf surface hairy. Petiolate, petiole 0.5 to 1 cm long, leaf apex retuse(Wolosik and Markowska, 2019; Mukuwapasiet al., 2024).

Inflorescence is numerous concentrated spikes, axillary arrangement, ended with racemes and spikes. More than 50 cm long, reddish or pinkish colour. Branches are 40-45 cm long, perpendicular with numerous laterals. Monoecious bearing both male and female flower (Wolosik and Markowska, 2019).

Flowers are numerous, unisexual, greenish and form a terminal spike (Wolosik and Markowska, 2019).

This plant produces thousands of seeds which are round or lenticular, dark brown or shiny, Campylotropus ovule (Wolosik and Markowska, 2019). Endosperm present in the micropylar region (Wolosik and Markowska, 2019)

Dissected parts of the plant under the simple microscope are represented in figure 1.

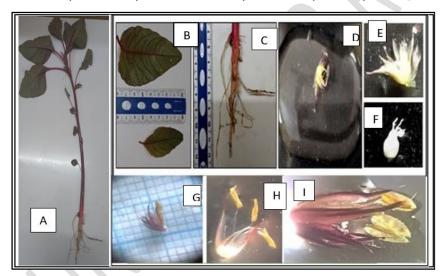


Figure 1. Morphological characters of Amaranthus cruentus (where A, B, C, D, E, F, G, H,I represent full plant, dorsal and ventral surface of the leaf, root morphology, Dissection of female flower, male flower)

3.ECONOMICAL USES

3.1Food: Amaranthuscruentus is a leafy vegetable (Toraneetal., 2017;Mukuwapasiet al., 2024)and useful food supplement for human. This vegetable is similar to the well-known Barley, Wheat and Bazra crop for its nutritional properties and contain high amount of amino acids. So, this plant is considered as 'Alternative Source of Crop' or 'Pseudocereal' (Fletcher, 2015; Mburu etal., 2011) and it protects the disease Malnutrition or Undernutrition. It has essential nutritional properties i.e. amino acid glutamine (Motta etal., 2019) proteins (Lysine and Tryptophan), vitamins, essential minerals, saturated and unsaturated fatty acid (Grestaetal., 2020), tocopherol, tocotrienol etc. and also contain high amount of fiber(Ali, 2021). Powdery dry leaves are used in sauces during the dry weather (Aderibigbeetal., 2022; Toraneetal., 2017). In Nigeria, leaves are mixed with the condiments to

prepare the soups (Mepba&Banigo, 2007; Ali, 2021). Boiled leaves are mixed with groundnut flower and consumed as relish (Ali ,2021). Leaves are consumed as spinach or green vegetable (Dhellotetal., 2006). Mixture of boiled leaves and groundnut sauce are prepared for salad in West Africa and Mozambique (KauffmanandWeber,1990). Seeds are also used as nutritious food which enriched in minerals, vitamins and proteins (Emam et al., 2024;Mukuwapasi et al., 2024). In America, this plant is cultivated as Grain seed or grain crop. Seeds are boiled and then eaten as rice. Laddoos are made from Grain Amaranth in India. Flour is made from the seeds which is gluten free, so it is fed to the Cealic patient or Gluten allergic patient comfortably and popularly used in baking Industry and producing snack bars (Mukuwapasi et al., 2024, Wolosik and Markowsa, 2019). The steps are followed to prepare flour from seeds are Flaking, Extruded and finally Grinding.

- **3.2Fodder**: This plant is used not only for food but also as fodder. It has been planted as leafy vegetables for cattle, rabbit, pigs and chicken food across many countries. Dietary fiber present in the plant have positive impact on animal gut by improving behavior on digestive tract (Manyeloetal., 2022; Alegbejo 2013). A. cruentus plant have high energy source and have not any negative effect on cattles i.e., Ships, Rabbits and Pigs. It is a substitute of Oil cake which is very essential for animal food. But, oil cake is too much costly. The clustered inflorescence of this plant have potential nutritional properties which acts as a substitute of Oil cake in Rabbits. In case of pig, this Amaranth grain is also helpful because it fulfils the dietary criteria but not fatter the pigs. It has no negative effect on growth of lambs also (Grestaet al., 2020).
- **3.3Cosmetics Industry:** Amaranth oil contains high amount of unsaturated fatty acid, tocopherols, Squalene which act as hair and skin conditioner (Baraniak&Kania-Dobrowolska2022). This seed oil also cures aging effect, wounds, skin diseases and moistening the skin. It is efficiently used as strong antioxidant and photoprotector in herbal medicine industry (Baraniak&Kania-Dobrowolska,2022).

It is also used as lubricant in computer industry (Achiganetal., 2014).

Oil extraction: Amaranthseeds were grinded for 30 seconds in a micro mill and oil extractedwith hexane in a Soxhlet apparatus for 7 hour (Leon*et al.*, 2001). Antioxidant extraction:Powdered seed samples and blended grains (1 g) were extracted with40 mL 8:1:1 (*v/v*) methanol–0.16 molar hydrochloric acid–water for 2 h. Theextracts were then separated by decantation and the residues were extracted with 40 mL 70% acetone again for 2 h. The extracts were combined, separatedby decantation, centrifuged, and stored in dark condition in a refrigerator at –20°C (Pasko et al., 2008).

- **3.4Dye**: In some countries, this plant is cultivated or grown only for making different dyes (Toraneetal., 2017) Dried plant is burnt for the preparation of potash in Benin countries (Toraneetal., 2017). Plants are used to extract red dye or coloring corn-based foods in South Western United states (Allemann, 2016). Red dye obtained from this plant is utilized to colour textile materials, medicines, food product and beverages (Ogwu, 2020).
- **3.5Ornamental value**: A. cruentus is cultivated as an ornamental plant in some countries (Toraneetal., 2017) specially in Nigeria (Ogwu, 2020; Mukuwapasi et al., 2024) valued for its feather-like flowering plumules and also have aesthetic value also (Hricovaetal., 2016). And sometimes it used as 'potherb' (Das & das, 2016). The A. cruentus isgreatly valued as ornamental because of its bright colour flowers(Amosovaetal., 2024).
- **3.6Medicinal properties**: It has various medicinal properties i.e. antioxidant (Osei et al., 2023), anti-inflammatory properties, antihypertensive properties stimulation of immune system, cardioprotective, antidiabetic, antimalarial, hepatoprotective (Peter & Gandhi,2017), anthelmintic (Torane et al., 2017), antiandrogenic (Alegbejo, 2013) etc. Bioactive secondary metabolites and protein hydrosylates present in the plant are potential anticancerous, antimicrobial agents (Ramkisson et al., 2020). Presence of high amount of fiber reduces cholesterol levels as well as risk of cardiovascular diseases (Jimoh et al., 2022) and inhibits the intake of starchy food decreases the chances of diabetes (Adeleke &Babalola, 2021). Prevention of diuretic and hypertension problem is revealed by potassium content (George, 2015). Oils or fats decreases the lipid content of blood and chances of coronary diseases have been reduced (Chatepa& Masamba,2020)

Traditional Methods: According to report of Azab 2020, the cooked leaves of the plant have laxative properties. Seeds have some medicinal properties also i.e. hypocholesterolemic effect. For infants,

treatment of constipation, roots of this plant is boiled with honey and feed it (Aderibigbe et al., 2022). Its water extract is used for the treatment of limb pain and as expellant of tapeworm also for wound dressing and tumors (Aderibigbe et al., 2022). This leafy vegetable is also important for lactating women to treat kidney complaints, constipation, anemia etc. (Torane et al., 2017; Aderibigbe et al., 2022; Achigan et al., 2014). Decoction of leaves is utilized as sore throat, ulcers and extraction of inflorescence swallowing by women for relieving from periodic pain (Ali et al., 2021).

Industrial Methods: Ethanolic leaf extract have protective effect such as restoring RBC, WBC and hemoglobin levels in blood (Azab, 2020). So, it is used in Anemia treatment. A. cruentus extract administrated to rats receive Cyclophosphamide which is an immunosuppressant to treat nephritis and arthritis and also used in Chemotherapy in combination with other drugs (Pandey et al., 2016). Ethanolic leaf extract of mixture of Amaranthus cruentus and Cleome gynandra have blood glucose lowering effect (Siwale ,2018).

4.BIOACTIVE SECONDARY METABOLITES PRESENT IN THE PLANT

A number of different bioactive secondary metabolite present in the plant is graphically represented in Figure 2 (Peter&Gandhi,2017). which helps to promote plant growth and protect from pathogens. These metabolites also possess antioxidant, biochemical and pharmacological properties and beneficial for human health (Omale & Okafor,2008).

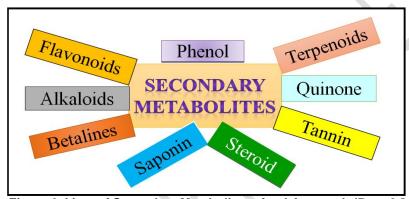


Figure 2. Lists of Secondary Metabolites of red Amaranth (Peter&Gandhi, 2017)

- **4.1Betalines**: Betacyanin (red violet pigment) and Betaxanthin (yellow pigment) present in different parts (root, stem, leaf, flower) of the plant (Castrillón&Frier, 2016), water soluble vacuolar pigment used in Folk medicine and also as anticancerous, anti-inflammatory and cardiovascular agent (Nana etal., 2012). The Betacyanin and Betaxanthin content were 7.74 and 19.34 mg/100 g dry weight respectively (Nana et al., 2012). Extraction and testing method of Betalines followed by the method Nana et al., 2012. 5gram fresh plant material was homogenized in mortarpestle and extracted with 25 ml distilled water containing 250 milimolar ascorbate for 15 minutes. Then centrifuged at 6000 rpm for 15 minute and stored the supernatant at 4°C. Betalines content was done by spectrophotometric method 536 nm for betacyanins and 475 nm for betaxanthins with molar extinction coefficient.
- **4.2Phenolics**: Anthocyanin produce excessive radicles which surpass the physiological antioxidant properties (Alvarez etal.,2010). Salicylic, cinnamic, ferulic, p-coumaric, gallic, sinapic acids are also present (Peter&Gandhi,2017;Castrillón&Frier, 2016). Premature grain of this plant contains a huge amount of phenolic acids i.e., ferulic acid, caffeic acid, galic acid, p-coumaric acid and anthocyanin) than mature grain (Manyeloetal., 2022, Osei et al., 2023). It has been reported that phenolics have beneficial role in antioxidative property serving as the free radical scavengers and lower the heart diseases and inflammation (Omale&Okafor,2008; Iwuagwuetal., 2019). Biosynthetic pathway of phenolics is diagrammatically represented in figure 3(Buchanon, 2015).

Extraction procedure is done according to the method of Torane et al., 2017.First, plant material was collected and then air dried. Afterthat,10g plant material was extracted with 50 ml of solvent which have different polarity. Then they are kept at room temperature for 24hrs.To obtain the crude extract,

the solvent was recuperated under low pressure. Quantity of each phenolics is represented in Table 1.

Table1. Quantity of each phenolics in premature and mature grain (Manyelo et al.,2022)

Phenolics	Premature Grain (mg/kg)	Mature Grain(mg/kg)
Ferulic acid	310	345.20
Gallic acid	41	43.48
Caffeic acid	6.5	9.93
p-Coumaric acid	1.2	2.94
Anthocyanin	35.2	35.92

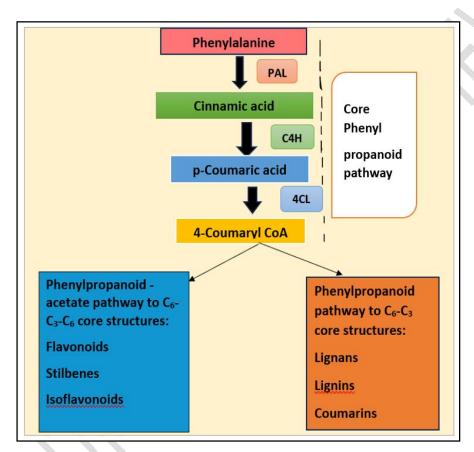


Figure 3. Overview of the biosynthetic pathways contribute to the production of several major phenolic compounds in plants. The synthesis process starts with Phenylalanine and diverge into phenylpropanoid – acetate and phenylpropanoid pathways. First three steps are common to both pathways. The core phenylpropanoid pathway ends with the formation of 4-Coumaryl CoA where PAL- Phenylalanine ammonia lyase, C4H- Cinnamate-4- hydroxylase, 4CL-Coumarate CoA ligase. adapted from Buchanon (2015).

- **4.3Flavonoids**: Rutin is one of most important flavonoids reported to have various beneficial effects on human and protective effect on memory disfunction (Koda etal., 2009). Quercetin is also reported. It serves as antimicrobial agent and showing potential antioxidant property (Castrillón&Frier, 2016) and reduce the oxidation level of triglyceride, cholesterol, low density lipoprotein (Erdtmanetal., 2007).
- **4.4Terpenoids**: Terpenoid showing antimicrobial, antifungal, antiparasitic, antiviral, anticancerous, antispasmodic, antiallergic, anti-inflammatory, immunomodulatory properties (Rabi&Bishayee,2009).

- **4.5Alkaloids**: It is one of the important metabolites in living organisms help in development of the plant and served as useful repellant for any pathogen or parasite and possesses antimicrobial properties (Usunomena&Ngozi,2016;Cherian&Sheela,2016).
- **4.6Quinones**: According to the result of phytochemical screening, ethanolic extract contains flavonoid, phenoilcs and quinones which are used in glucose tolerance test i.e., antihyperglycemia (Nofianti,2015).
- **4.7Saponin**: It limits the growth of cancerous cells and has beneficial effects on antiinflammation (Prohp&Onoagbe,2012; Rao&Sung,1995) and stimulate release of insulin and lower blood glucose level (Siwale, 2018).
- **4.8Steroid**: Steroid is present at vegetative and flowering stage which protect from harmful effects of microbial pathogens and also have anti-inflammatory and anti-aging properties (Okunlolaetal., 2017). Amaranth grain and leaf extract contain high amount of Squalene (He&Corke, 2003).
- **4.9Tannin**: It has been reported to possess potential antiviral (Cheng& Lin, 2002) and anticancerous activity (Narayanan etal., 1999).

The Extraction procedure is discussed here. Collected plant material was air dried and grinded and then filtrate to obtain fine powder. 30 g dried plant powder was soaked in 300 ml of ethanol andnhexane respectively and kept this mixture for 2 days. Then the mixtures were filtered by using the Whatman filter paper. The filtered material

was dried in the vacuum by utilizing the rotary evaporator. Dried-out residue was used as extract (Edo and Onoharigo,2024).

This procedure confirms the presence of alkaloids, tannins, steroids, terpenoids and quinones in ethanol and n-hexane extract.

Saponin extraction begins with the dried plant extract mixed with methanol or water and then used organic solvent chloroform to remove the lipid substance. Soxhlet extraction done, after evaporation and most often dissolution in water and phase separation into n-butanol. Saponins with higher polarity could found in aqueous phase (Mroczek, 2015).

The quantity of each secondary metabolites is represented in Table 2.

Table 2. Quantity of each secondary metabolites (Ali,2021)

SecondaryMetabolite	Quantity (%)
Phenolics	3.10
Flavonoids	11.60
Terpenoids	1.8
Alkaloids	3.6
Saponin	1.3
Steroid	4.3
Tannin	2.40

5.BENEFICIAL MICROBIAL ASSOCIATION

5.1Endophytes: Numerous studies have shown that symbiotic (mostly Endophytic) fungi enhance abiotic stress tolerance mainly water deficit i.e. Drought and protect the host plant from critical environmental condition and plant survive (Rodriguezetal., 2009; Singhetal., 2011). Endophytic microbes reside within living plant tissues and can interact with roots, stem and leaves without causing symptoms of diseases in their host plants (Schulz& Boyle, 2006). Commonly found fungal endophytes belong to different classes of Ascomycota, mostly Dothidiomycetes, Leotiomycetes, and Sordariomycetes, and in some hosts Basidiomycetous fungi are also observed (Rodriguez et al., 2009). Endophytic fungi form a symbiont with the host plant and can protect from insect herbivore by changing the chemical composition of the plant and form a chain between the parasitism and mutualism (Rodriguez et al., 2009). The endophyte infected plants express a range of adaptations to biotic stress (Morelietal., 2020) and abiotic stresses mainly drought tolerance. Some previous reports demonstrated that many grass species showed vegetative growth improvement in the presence of their fungal symbionts and these endophytic fungi enlarge plant fitness by the discharge of plant

growth advancing secondary metabolites (Priti etal., 2009; Hardoimetal., 2015). Endophytes have both the horizontal and vertical mode of transmission (Frank etal., 2017). Endophytic microbes are mainly bacteria and fungi and some reports revealed that it may be algae also (Rit et al., 2023). Fungal endophytes mainly belong to the Ascomycetes, Zygomycetes and Deuteromycetes group and Basidiomycetous fungi is rarely reported (Rajamanikyametal., 2017; Stoneetal., 2000). Endophytes showing mutualistic behaviour i.e., it receives nutrition and shelter from host plant and give the competitive ability to the host (Rajamanikyametal.,2017). After isolation, pure culture and characterization of each specific endophyte, Maximum individual possessing antibiotics, anticancerous, antimycotics and immunosuppressant property (Alvin etal., 2014). Treatment of various types of endophytes upon Amaranthus plant showing enhancement of growth due to discharge of different plant growth regulator i.e. IAA, GA etc and increased SAR by decreasing diseases by direct antagonistic behaviour (Uppala etal., 2010). Fungi Isolated from Phyllosphere and Rhizosphere of A. cruentus (Cladosporium, Talaromyces, Gibberella, Penecillum, Alternaria, Trichoderma, Rhizopus, Fusarium) increase the soil quality of the plant and plants health and produce antibiotic compound against pathogens (Puszetal., 2015). According to Uppala etal., 2010, several bacterial endophytes promotes biometrics of Amaranth. Some endophytic bacteria isolated from plant roots and promote plant growth, survival and protect from biotic stress (Enebe&Babalola, 2018). Endophyte protects the host plant from oxidative damage by producing ROS scavenging enzymes i.e. Superoxide dismutase (SOD), Glutathione peroxidase (GR) etc. (Choudhury etal., 2023). It was reported in Red Amaranth that the application of an endophytic strain Sphingomonaspanaciterraenot only increases the morphological parameter i.e., root length, shoot length, leaf number etc. of the host plant but also increase the chlorophyll and mineral content of the plant(Sultana et al., 2024).

Endophytic fungi are isolated(Shilpa&Shikha, 2015) from different partsof the plant *A. cruentus* is represented in figure 4.

Collection of plant and washed under running tap water and segmented into suitable sizes

P

Surface sterilization with 70% ethanol for 30 seconds, 2% Sodium hypochlorite for 30 seconds, 0.1%mercuric chloride for 10-20 seconds, 5% Azithromycin (antibiotic) solution for 30 seconds

卆

Segmented plant parts are washed in distilled water and placed on potato dextrose agar (PDA) plates

Figure 4. Protocol of Isolation of endophytic fungi from A. cruentus (Shilpa and Shikha, 2015)





The plates containing pure culture of the isolates are represented in figure 5.

Figure 5. Fungal isolates after pure culture

According to standard protocol, leaf tissues of red Amaranth were dissolved in organic solution and used to detect endophytes under microscope after staining with Trypan blue (Vahabi et al., 2011).

5.2 PGPR and PGPF: Plant Growth Promoting Rhizobacteria (PGPR) are a group of bacteria that enhance phytoremediation (Muthukumar et al., 2017). PGPR are actually the beneficial, non pathogenic rhizobacteria which colonize the root surfaces but not inside the plant tissues and promote the growth and development of the host plant (Rit et al., 2023, Choudhury et al., 2022). It colonizes plant root surface efficiently and promote plant growth.PGPR may facilitate plant growth either by promoting the acquisition of nutritional resources such as Nitrogen, Phosphorus or Iron; preventing or decreasing the damage to plants by pathogenic organisms (mainly fungi and bacteria) or by directly stimulating plant growth by either providing plant hormones such as Auxin, Cytokinin or GA or lowering plant ethylene levels through the action of the enzymes 1 amino-cyclopropane 1 carboxylic acid deaminase (ACC)(del Carmen Orozco-Mosqueda et al., 2020, Choudhury et al., 2022, Rit et al., 2023) . Other mechanisms involved i.e. solubilization of minerals, production of siderophores, increases in leaf area, chlorophyll and soluble protein content. These can also produce antioxidant, enzymes to protect plants from environmental stresses that lead to generation of ROS which cause cell damage (Choudhury et al., 2022). Rhizobacteria may reduce the level of disease by synthesizing hydrolytic enzymes, antibiotics, introduction of systemic acquired resistance (SAR) and competition for nutrients and niches (Sindhu & Sharma, 2019). PGPR significantly decrease the use chemical fertilizer, pesticides, insecticide and herbicides in modern agricultural practices(Choudhury et al., 2022).

PGPR *Pseudomaonas sp.*, isolated from rhizosphere soil improve plant growth and crop production and survival of *Amaranthus* species and protect from pathogen attack(Enebe& Babalola, 2018;Ikhajiagbe et al., 2021). They have the ability to produce siderophore, hydrogen cyanide (HCN) and phosphate solubilization property (Sagar et al., 2018).

PGPF are mainly rhizospheric fungi which promote plant growth and protected from diseases by colonizing the root. But, endophytic fungi (inside the plant tissues), phyllospheric fungi are also included in PGPF (Rit et al., 2023;EI Maraghy et al., 2021).Plant Growth Promoting Fungi (PGPF) maintain plant growth through the generation of a number of significant enzymes like ACCD (1-aminocyclopropane-1-carboxylate deaminase), urease, catalase etc, phosphate solubilization, siderophores and IAA production, antagonism to pathogens and resistant to antibiotics and take a crucial role in plant growth. PGPF are non-pathogenic, soil borne fungi enhance plant growth and survival (HyAKuMAcHi,1994; Debashis et al., 2020). It also enhances soil fertility, productivity, nutrient uptake and Defence mechanism (Yapa et al., 2022). It triggers induced systemic resistance (ISR) against pathogenic microorganism by modifying cell wall accumulating callose, phenol, lignin (Yapa et al., 2022). Different PGPR and PGPF strains when combined and applied to soil of

Amaranth, then morphological parameters i.e., shoot height, root height, fresh weight all are increased and induced SAR (Debashis et al., 2020).

5.3 VAM: Mycorrhiza (VAM) is the symbiotic association between higher plants and fungi(Giovannini etal., 2020). The fungus grows internally inside the root cortical cells where they form specialized structures called vesicles and arbuscules(Holland& Roth, 2023). Vesicles are swollen, sac like, unbranched, modified structures of the fungal hyphae. The major function is the storage of reserve food whereas arbuscules are branched, finger like projected hyphal modifications of the endomycorrhiza and the major function is the absorption of food nutrients and water from the host cytoplasm (Holland & Roth, 2023). VAM efficiently extend plant roots making the uptake of water much more efficient, better mineral nutrition especially Phosphate, alteration in root architecture, modification of some physiological and enzymatic activities, induction of the plant hormones ABA (Fengetal., 2020). VAM inoculation increases stomatal conductance under drought condition. The VAM fungi are obligatory biotrophs in nature (Giovannini etal., 2020) and also act as bioprotectants against pathogen and toxic stresses. Application of VAM as fertilizer to the Amaranth plant significantly increase the shoot diameter, root length, leaf number etc. (Dada etal., 2017). According to the study of Safeena&Sasmina, 2018, AM (Glomus fasiculatum) were inoculated into the pesticide treated soil of A. cruentus, the result revealed that growth of the plant is increased and pesticide toxicity in the soil is decreased and soil-microbe growth are not inhibited. Report of Kalaikandhanetal.,2012, revealed that when VAM (*Glomus hoi*) is inoculated in soil of *A. cruentus*, heavy metal concentration of soil is reduced because of the VAM fungi having capacity to sequester it in their mycelia and make the soil less toxic. n semiarid and arid regions of India, occurrence of VAM was reported in some members of Amaranthaceae mainly Amaranthus. Vesicles, arbuscules and intracellular hyphae was observed in root cortical zone of the host plant (Neeraj et al., 1991)

7.CONCLUSION

This plant has huge role in food, fodder, dye, cosmetic and medicinal industry. Secondary metabolites extracted from this plant possesses important role in pharmacy Food quality is more important to prevent malnutrition than food quantity. Major food crops are deficient in proper nutrient. By consuming these crops, many peoples are susceptible to diseases. This Red Amaranth is considered as 'Alternative source of Crop' or 'Pseudocereal'. It contains high percentage of nutritional compounds, bioactive secondary metabolites and medicinal properties. This plant is not only used for human food but also very efficient Fodder crop. This leafy vegetable is rich source of dietary fibre which helps in digestion of animals. In some countries, it, regarded as 'Ornamental' and also used in dye and cosmetic industry. Bioactive secondary metabolites (Phenolics, Flavonoids, Terpenoids, Saponins, Alkaloids, Quinone, Tannin) present in this plant prevent the entry of pathogenic microorganisms and have several medicinal activities i.e. antioxidant, anti-inflammatory, immunomodulatory etc. Microbes are a noble and consistent source of unique natural medicines with a high level of biodiversity and may also yield several pharmaceutically important compounds. This symbiotic association at the molecular and genetic levels will be helpful for enhancing secondary metabolite production and can be an active area for future investigations. In recent years, there have been a large number of studies, both in the laboratory and in the field, that have provided information about the use of plant- associated microbes to the benefit of plant growth and development. Bacteria, fungi, mycorrhiza provide resistance to plants against various environmental (abiotic) stresses. Fungi isolated from this plant secret antibiotic compounds and protect plant from abiotic stresses (Pusz et al., 2015). PGPF are non- pathogenic soil borne fungi enhance nutrient uptake and soil quality of the plant have different antagonistic mechanisms.VAM have the capacity to decrease the heavy metal toxicity and devasting effect of pesticides of the soil. In the recent time, we are hopefully determined that the purposeful use of microorganisms will be commenced on to replace the use of energy-intensive and often pollution causing chemicals in agricultural practices. Application of microbes and microbial inoculants as biofertilizers and biocontrol agent is an integral component in organic farming practices. Lastly, improved understanding of 'mycorrhizosphere' interactions is crucial in order to preserve crop yield stability under an increasingly unstable climate change. To think about the food scarcity of our near future, we should protect this plant as a Crop for alternative source and try to increase the production of the plant by using the associated microbes and secondary metabolites extracted from this plant.

Acknowledgement

Authors are like to acknowledge Department of Botany, University of Burdwan for providing the research facilities. For financial support, Chandrama Mukherjee and Shinjan Dey are thankful to the State Funded Fellowship, Govt. of West Bengal. Debapriya Choudhury and Sayani Datta are thankful to UGC, Govt. for India for funding.

Competing interests

Authors declared that no competing interests exist.

Author Contribution:

Main concept was given by Sikha Dutta. Literature review was performed by Chandrama Mukherjee. Editing of figures, arrangement of references and formatting of total manuscript was done by Debapriya Choudhury. Arrangement of references were done by Shinjan Dey and Sayani Datta. Whole manuscript was discussed by all the authors. Finally, all authors read and approved the manuscript.

Disclaimer (Artificial intelligence)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

REFERENCES

Achigan-Dako, E. G., Sogbohossou, O. E., & Maundu, P. (2014). Current knowledge on Amaranthus spp.: research avenues for improved nutritional value and yield in leafy amaranths in sub-Saharan Africa. *Euphytica*, *197*, 303-317. DOI 10.1007/s10681-014-1081-9

Adeleke, B. S., & Babalola, O. O. (2021). Pharmacological potential of fungal endophytes associated with medicinal plants: A review. *Journal of Fungi*, 7(2), 147. https://doi.org/10.3390/jof7020147

Aderibigbe, O. R., Ezekiel, O. O., Owolade, S. O., Korese, J. K., Sturm, B., & Hensel, O. (2022). Exploring the potentials of underutilized grain amaranth (Amaranthus spp.) along the value chain for food and nutrition security: A review. *Critical reviews in food science and nutrition*, 62(3), 656-669. https://doi.org/10.1080/10408398.2020.1825323

Alegbejo, J. O. (2013). Nutritional value and utilization of Amaranthus (Amaranthus spp.)—a review. *Bayero journal of pure and applied sciences*, *6*(1), 136-143. https://doi.org/10.4314/bajopas.v6i1.27

Ali, M., Dahiya, S. S., Sultana, S., & Munjal, K. (2021). Fatty acid constituents from the leaves of Amaranthus hybridus L. *European Journal of Pharmaceutical and Medical Research*, 8(9), 613-617.

Ali, M. (2021). Determination of Proximate, Phytochemicals and Minerals Composition of Amaranthus spp. DOI: 10.36346/sarjps.2021.v03i05.002

Allemann, I. (2016). Influence of abiotic stress on allelopathic properties of Amaranthus cruentus L. http://hdl.handle.net/11660/5825

Alvarez-Jubete, L., Wijngaard, H., Arendt, E. K., & Gallagher, E. (2010). Polyphenol composition and in vitro antioxidant activity of amaranth, quinoa buckwheat and wheat as affected by sprouting and baking. *Food chemistry*, 119(2), 770-778. https://doi.org/10.1016/j.foodchem.2009.07.032

- Alvin, A., Miller, K. I., & Neilan, B. A. (2014). Exploring the potential of endophytes from medicinal plants as sources of antimycobacterial compounds. *Microbiological research*, *169*(7-8), 483-495. https://doi.org/10.1016/j.micres.2013.12.009
- Amosova, A. V., Yurkevich, O. Y., Semenov, A. R., Samatadze, T. E., Sokolova, D. V., Artemyeva, A. M., ... &Muravenko, O. V. (2024). Genome Studies in Amaranthus cruentus L. and A. hypochondriacus L. Based on Repeatomic and Cytogenetic Data. *International Journal of Molecular Sciences*, 25(24), 13575. https://doi.org/10.3390/
- Azab, A. (2020). Amaranthaceae plants of Israel and palestine: Medicinal activities and unique compounds. *EUROPEAN CHEMICAL BULLETIN*, *9*(12), 366-400. http://dx.doi.org/10.17628/ecb.2020.9.366-400
- Baraniak, J., & Kania-Dobrowolska, M. (2022). The dual nature of amaranth—Functional food and potential medicine. *Foods*, *11*(4), 618. https://doi.org/10.3390/foods11040618
- Buchanan, B. B., Gruissem, W., & Jones, R. L. (2015). Biochemistry and molecular biology of plants (2nd ed) John wiley& sons.
- Castrillón-Arbeláez, P. A., & Frier, J. P. D. (2016). Secondary metabolism in Amaranthus spp.—a genomic approach to understand its diversity and responsiveness to stress in marginally studied crops with high agronomic potential. *Abiotic and Biotic Stress in Plants. Recent Advances and Future Perspectives. India: In Tech*, 185-227. https://doi.org/10.5772/61820
- Chatepa, L. E. C., & Masamba, K. G. (2020). Proximate and phytochemical composition of selected indigenous leafy vegetables consumed in Malawi. *African Journal of Food Science*, *14*(9), 265-273. http://dx.doi.org/10.5897/AJFS2020.1979
- Cheng, H. Y., Lin, C. C., & Lin, T. C. (2002). Antiherpes simplex virus type 2 activity of casuarinin from the bark of Terminalia arjuna Linn. *Antiviral research*, *55*(3), 447-455. https://doi.org/10.1016/S0166-3542(02)00077-3
- Cherian, P., & Sheela, D. (2016). Antimicrobial activity of Amaranth Alkaloid against pathogenic microbes. *International Journal of Herbal Medicine*, 4(5), 70-72.
- Choudhury, D., Tarafdar, S., Parvin, N., Rit, R., Roy, S., & Dutta, S. (2023). Endophytic microbes and their diverse beneficial aspects in various sectors: A critical insight. *Plant Science Today*, *10*(1), 96-107. https://doi.org/10.14719/pst.1877
- Choudhury, D., Tarafdar, S., & Dutta, S. (2022). Plant growth promoting rhizobacteria (PGPR) and their eco-friendly strategies for plant growth regulation: A review. *Plant Science Today*, *9*(3), 524-537. https://doi.org/10.14719/ pst.1604
- Dada, O. A., Imade, F., & Anifowose, E. M. (2017). Growth and proximate composition of Amaranthus cruentus L. on poor soil amended with compost and arbuscular mycorrhiza fungi. *International Journal of Recycling of Organic Waste in Agriculture*, *6*, 195-202. DOI 10.1007/s40093-017-0167-5
- Das, S., & Das, S. (2016). Amaranths: the crop of great prospect. *Amaranthus: A promising crop of future*, 13-48. https://doi.org/10.1007/978-981-10-1469-7_3
- Debasis, M., Ganeshamurthy, A. N., Komal, S., Radha, T. K., & Rupa, T. R. (2020). Plant growth promotion and biocontrol activity of some typical harsh environment rhizo-microbes and their effects on Amaranthus cruentus plants. *Res. J. Biotechnol. Vol.*, 15, 1.
- del Carmen Orozco-Mosqueda, M., Glick, B. R., & Santoyo, G. (2020). ACC deaminase in plant growth-promoting bacteria (PGPB): An efficient mechanism to counter salt stress in crops. *Microbiological Research*, 235, 126439.https://doi.org/10.1016/j.micres.2020.126439
- Dhellot, J. R., Matouba, E., Maloumbi, M. G., Nzikou, J. M., Ngoma, D. S., Linder,

- M., ... & Parmentier, M. (2006). Extraction, chemical composition and nutrional characterization of vegetable oils: Case of Amaranthus hybridus (var 1 and 2) of Congo Brazzaville. *African journal of biotechnology*, 5(11).http://www.academicjournals.org/AJB
- Edo, G. I., &Onoharigho, F. O. (2024). A comparative study of biological and chemical composition of African spinach (Amaranthus cruentus) extract: An approach in drug formulation, food and nutrition. *Advances in Traditional Medicine*, *24*(2), 519-529. https://doi.org/10.1007/s13596-023-00718-x
- Emam, T. M., Hosni, A. E. M., Ismail, A., El-Kinany, R. G., Hewidy, M., Saudy, H. S., ... & El-Sayed, S. M. (2024). Physiological and Molecular Responses of Red Amaranth (Amaranthus cruentus L.) and Green Amaranth (Amaranthus hypochondriacus L.) to Salt Stress. *Journal of Soil Science and Plant Nutrition*, 1-12.https://doi.org/10.1007/s42729-024-02125-w
- Enebe, M. C., & Babalola, O. O. (2018). The influence of plant growth-promoting rhizobacteria in plant tolerance to abiotic stress: a survival strategy. *Applied microbiology and biotechnology*, 102, 7821-7835. https://doi.org/10.1007/s00253-018-9214-z
- Erdman Jr, J. W., Balentine, D., Arab, L., Beecher, G., Dwyer, J. T., Folts, J., ... & Burrowes, J. (2007). Flavonoids and Heart Health: Proceedings of the ILSI North America Flavonoids Workshop, May 31–June 1, 2005, Washington, DC1. *The Journal of nutrition*, 137(3), 718S-737S.https://doi.org/10.1093/jn/137.3.718S
- Feng, Z., Liu, X., Zhu, H., & Yao, Q. (2020). Responses of arbuscular mycorrhizal symbiosis to abiotic stress: a lipid-centric perspective. *Frontiers in Plant Science*, *11*, 578919. https://doi.org/10.3389/fpls.2020.578919
- Fletcher, R. J. (2015). Pseudocereals: overview.<u>https://doi.org/10.1016/B978-0-12-394437-5.00039-5</u>
- Frank, A. C., Saldierna Guzmán, J. P., & Shay, J. E. (2017). Transmission of bacterial endophytes. *Microorganisms*, *5*(4), 70.https://doi.org/10.3390/microorganisms5040070
- George, G. M. (2015). Postharvest handling for extending shelf life of Amaranthus (Amaranthus tricolor L.) (Doctoral dissertation, Department of Processing Technology, College of Agriculture, Vellayani).http://krishikosh.egranth.ac.in/handle/1/5810029695
- Gins, E. M., Baikov, A. A., Khasanova, S. D., Goryunova, S. V., Gins, V. K., Gins, M. S., &Motyleva, S. M. (2024). AMARANTHUS SPECIES ASSESSMENT FOR MORPHOLOGICAL AND BIOCHEMICAL PARAMETERS. SABRAO Journal of Breeding & Genetics. 56(4).http://doi.org/10.54910/sabrao2024.56.4.6http://sabraojournal.org/
- Giovannini, L., Palla, M., Agnolucci, M., Avio, L., Sbrana, C., Turrini, A., & Giovannetti, M. (2020). Arbuscular mycorrhizal fungi and associated microbiota as plant biostimulants: research strategies for the selection of the best performing inocula. *Agronomy*, 10(1), 106. https://doi.org/10.3390/agronomy10010106
- Gresta, F., Meineri, G., Oteri, M., Santonoceto, C., Lo Presti, V., Costale, A., & Chiofalo, B. (2020). Productive and qualitative traits of Amaranthus cruentus L.: An unconventional healthy ingredient in animal feed. *Animals*, *10*(8), 1428.https://doi.org/10.3390/ani10081428
- Hardoim, P. R., Van Overbeek, L. S., Berg, G., Pirttilä, A. M., Compant, S., Campisano, A., ... &Sessitsch, A. (2015). The hidden world within plants:

- ecological and evolutionary considerations for defining functioning of microbial endophytes. *Microbiology and molecular biology reviews*, 79(3), 293-320.https://doi.org/10.1128/mmbr.00050-14
- He, H. P., & Corke, H. (2003). Oil and squalene in amaranthus grain and leaf. *Journal of Agricultural and Food Chemistry*, *51*(27), 7913-7920.https://doi.org/10.1021/jf030489q
- Hoidal, N., Díaz Gallardo, M., Jacobsen, S. E., &Alandia, G. (2019). Amaranth as a dual-use crop for leafy greens and seeds: Stable responses to leaf harvest across genotypes and environments. *Frontiers in Plant Science*, *10*, 817.https://doi.org/10.3389/fpls.2019.00817
- Holland, S., & Roth, R. (2023). Extracellular vesicles in the arbuscular mycorrhizal symbiosis: current understanding and future perspectives. *Molecular Plant-Microbe Interactions*, *36*(4), 235-244. https://doi.org/10.1094/MPMI-09-22-0189-FI
- Hricova, A., Fejer, J., Libiakova, G., Szabova, M., Gazo, J., &Gajdosova, A. (2016). Characterization of phenotypic and nutritional properties ofvaluable Amaranthus cruentus L. mutants. *Turkish Journal of Agriculture and Forestry*, *40*(5), 761-771. 10.3906/tar-1511-31
- HyAKuMAcHi, M. (1994). Plant-growth-promoting fungi from turfgrass rhizosphere with potential for disease suppression. *Soil Microorganisms*, *44*, 53-68. https://doi.org/10.18946/jssm.44.0 53
- Ikhajiagbe, B., Ogwu, M. C., Fawehinmi, F. O., & Adekunle, I. J. (2021). Comparative growth responses of Amaranthus [L.] species in humus and ferruginous ultisols using plant growth promoting Rhizobacteria (Pseudomonas species). South African Journal of Botany, 137, 10-18.https://doi.org/10.1016/j.sajb.2020.09.029
- Iwuagwu, M. O., Ogbonna, N. C., & Okechukwu, U. H. International Journal of Plant Science and Horticulture. https://doi.org/10.36811/ijpsh.2019.110008
- Jimoh, M. O., Okaiyeto, K., Oguntibeju, O. O., & Laubscher, C. P. (2022). A systematic review on Amaranthus-related research. *Horticulturae*, *8*(3), 239. https://doi.org/10.3390/horticulturae8030239
- Kalaikandhan, R., Vijayarengan, P., Thamizhiniyan, P., & Sivasankar, R. (2012). Impact of the pesticides monocrotophos and quinalphos on the morphological features of red amaranth under arbuscular mycorrhizal fungus inoculation. *World J Agric Sci*, 8(2), 212-217.
- Kauffman, C. S., & Weber, L. E. (1990). Grain amaranth. *Advances in new crops. Timber Press, Portland, OR*, 127-139.
- Koda, T., Kuroda, Y., & Imai, H. (2009). Rutin supplementation in the diet has protective effects against toxicant-induced hippocampal injury by suppression of microglial activation and pro-inflammatory cytokines: protective effect of rutin against toxicant-induced hippocampal injury. *Cellular and molecular neurobiology*, 29, 523-531. https://doi.org/10.1007/s10571-008-9344-4
- Lanta, V., Havranek, P., & Ondrej, V. (2003). Morphometry analysis and seed germination of Amaranthus cruentus, A. retroflexus and their hybrid (A. x turicensis). *Plant Soil and Environment, 49*(8), 364-369.
- León-Camacho, M., García-González, D. L., & Aparicio, R. (2001). A detailed and comprehensive study of amaranth (Amaranthus cruentus L.) oil fatty profile. *European Food Research and Technology*, *213*, 349-355.DOI 10.1007/s002170100340

- Letchamo, W., Hartman, T., Gosslin, A., Mamedov, N. A., & Craker, L. (2017). The accumulation of phenolic compounds in genetically selected Amaranthus hybridus is influenced by endophytic natural growth regulator. *International Journal of Secondary Metabolite*, *5*(1), 12-19.http://dergipark.gov.tr/ijsm
- Ma, X., Vaistij, F. E., Li, Y., Jansen van Rensburg, W. S., Harvey, S., Bairu, M. W., ... & Denby, K. J. (2021). A chromosome-level Amaranthus cruentus genome assembly highlights gene family evolution and biosynthetic gene clusters that may underpin the nutritional value of this traditional crop. *The Plant Journal*, *107*(2), 613-628.https://doi.org/10.1111/tpj.15298
- Makinde, E. A., Ayeni, L. S., &Ojeniyi, S. O. (2010). Morphological characteristics of amaranthuscruentus L. as influenced by Kola Pod Husk, organomineral and NPK Fertilizers in Southwestern Nigeria. *New York Science Journal*, *3*(5), 130-134.http://www.sciencepub.net/newyork
- Malik, M., Sindhu, R., Dhull, S. B., Bou-Mitri, C., Singh, Y., Panwar, S., &Khatkar, B. S. (2023). Nutritional composition, functionality, and processing technologies for amaranth. *Journal of Food Processing and Preservation*, 2023(1), 1753029.https://doi.org/10.1155/2023/1753029
- Manyelo, T. G., Sebola, N. A., Hassan, Z. M., Ng'ambi, J. W., Weeks, W. J., &Mabelebele, M. (2022). Chemical composition and metabolomic analysis of Amaranthus cruentus grains harvested at different stages. *Molecules*, 27(3), 623. https://doi.org/10.3390/molecules27030623
- Mburu, M. W., Gikonyo, N. K., Kenji, G. M., &Mwasaru, A. M. (2011). Properties of a complementary food based on amaranth grain (Amaranthus cruentus) grown in Kenya. *Journal of Agriculture and Food Technology*, 1(9), 153-178.
- Mepba, H. D., Eboh, L., &Banigo, D. E. B. (2007). Effects of processing treatments on the nutritive composition and consumer acceptance of some Nigerian edible leafy vegetables. *African Journal of Food, Agriculture, Nutrition and Development*, 7(1). http://ajfand.net/AJFAND/copyrightstatement.html
- Morelli, M., Bahar, O., Papadopoulou, K. K., Hopkins, D. L., & Obradović, A. (2020). Role of endophytes in plant health and defense against pathogens. Frontiers in Plant Science, 11, 1312.https://doi.org/10.3389/fpls.2020.01312
- Motta, C., Castanheira, I., Gonzales, G. B., Delgado, I., Torres, D., Santos, M., & Matos, A. S. (2019). Impact of cooking methods and malting on amino acids content in amaranth, buckwheat and quinoa. *Journal of Food Composition and Analysis*, 76, 58-65.https://doi.org/10.1016/j.jfca.2018.10.001
- Mousavi, S. S., & Karami, A. (2022). Application of Endophyte microbes for production of secondary metabolites. *Application of Microbes in Environmental and Microbial Biotechnology*, 1-37.https://doi.org/10.1007/978-981-16-2225-0_1
- Mukuwapasi, B., Mavengahama, S., & Gerrano, A. S. (2024). Grain amaranth: A versatile untapped climate-smart crop for enhancing food and nutritional security. *Discover Agriculture*, 2(1), 44. https://doi.org/10.1007/s44279-024-00057-8
- Muthukumar, A., Udhayakumar, R., & Naveenkumar, R. (2017). Role of bacterial endophytes in plant disease control. *Endophytes: Crop Productivity and Protection: Volume 2*, 133-161.https://doi.org/10.1007/978-3-319-66544-3_7
- Nana, F. W., Hilou, A., Millogo, J. F., & Nacoulma, O. G. (2012). Phytochemical composition, antioxidant and xanthine oxidase inhibitory activities of Amaranthus

- cruentus L. and Amaranthus hybridus L. extracts. *Pharmaceuticals*, *5*(6), 613-628.https://doi.org/10.3390/ph5060613
- Narayanan, B. A., Geoffroy, O., Willingham, M. C., Re, G. G., & Nixon, D. W. (1999). p53/p21 (WAF1/CIP1) expression and its possible role in G1 arrest and apoptosis in ellagic acid treated cancer cells. *Cancer letters*, *136*(2), 215-221.https://doi.org/10.1016/S0304-3835(98)00323-1
- Neeraj, Shanker, A., Mathew, J., & Varma, A. (1991). Occurrence of vesicular-arbuscular mycorrhizae with Amaranthaceae in soils of the Indian semi-arid region. *Biology and Fertility of soils*, *11*, 140-144.https://doi.org/10.1007/BF00336379
- Nofianti, T. (2015). ANTIHYPERGLICEMIA ACTIVITY OF WATER FRACTION, ETHYL ACETATE FRACTION, AND N-HEXANE FRACTION OF SPINACH LEAVES EXTRACT (Amaranthus Cruentus L) ON MALE MICE SWISS. *JURNAL FARMASIGALENIKA*, 2(02).https://journal.bku.ac.id/jfg/index.php/jfg/article/view/36
- Ogwu, M. C. (2020). *Value of Amaranthus [L.] species in Nigeria* (pp. 1-21). London: IntechOpen. https://dx.doi.org/10.5772/intechopen.86990
- Okunlola, G. O., Jimoh, M. A., Olatunji, O. A., &Olowolaju, E. D. (2017). Comparative study of the phytochemical contents of Cochorusolitorius and Amaranthus hybridus at different stages of growth. *Annales of West University of Timisoara*. Series of Biology, 20(1), 43.
- Omale, J., & Okafor, P. N. (2008). Comparative antioxidant capacity, membrane stabilization, polyphenol composition and cytotoxicity of the leaf and stem of Cissusmultistriata. *African Journal of Biotechnology*, 7(17). http://www.academicjournals.org/AJB
- Omami, E. N., & Hammes, P. S. (2006). Interactive effects of salinity and water stress on growth, leaf water relations, and gas exchange in amaranth (Amaranthus spp.). New Zealand Journal of Crop and Horticultural Science, 34(1), 33-44. https://doi.org/10.1080/01140671.2006.9514385
- Osei-Owusu, J., Kokro, K. B., Ofori, A., Apau, J., Dofuor, A. K., Vigbedor, B. Y., ... & Okyere, H. (2023). Evaluation of phytochemical, proximate, antioxidant, and antinutrient properties of Corchorus olitorius, Solanum macrocarpon and Amaranthus cruentus in Ghana. *International Journal of Biochemistry and Molecular Biology*, *14*(2), 17.
- Pandey, S., Ganeshpurkar, A., Bansal, D., & Dubey, N. (2016). Hematopoietic effect of Amaranthus cruentus extract on phenylhydrazine-induced toxicity in rats. *Journal of Dietary Supplements*, *13*(6), 607-615. https://doi.org/10.3109/19390211.2016.1155685
- Park, Y. J., Nishikawa, T., Matsushima, K., Minami, M., & Nemoto, K. (2014). A rapid and reliable PCR-restriction fragment length polymorphism (RFLP) marker for the identification of Amaranthus cruentus species. *Breeding science*, *64*(4), 422-426. https://doi.org/10.1270/jsbbs.64.422
- Paśko, P., Sajewicz, M., Gorinstein, S., & Zachwieja, Z. (2008). Analysis of selected phenolic acids and flavonoids in Amaranthus cruentus and Chenopodium quinoa seeds and sprouts by HPLC. *Acta chromatographica*, *20*(4), 661-672.10.1556/AChrom.20.2008.4.11

- Peter, K., & Gandhi, P. (2017). Rediscovering the therapeutic potential of Amaranthus species: A review. *Egyptian journal of basic and applied sciences*, *4*(3), 196-205. https://doi.org/10.1016/j.ejbas.2017.05.001
- Prajitha, V., &Thoppil, J. E. (2018). Cytogenetic characterization of Amaranthus caudatus L. and Amaranthus hybridus subsp. cruentus (L.) Thell. *Cytotechnology*, *70*, 95-101. https://doi.org/10.1007/s10616-017-0100-9 Priti, V., Ramesha, B. T., Singh, S., Ravikanth, G., Ganeshaiah, K. N., Suryanarayanan, T. S., & Uma Shaanker, R. (2009). How promising are endophytic fungi as alternative sources of plant secondary metabolites?. *Current Science*, *97*(4), 477-478.
- Prohp, T. P., &Onoagbe, I. O. (2012). Determination of phytochemical composition of the stem bark of Triplochitonscleroxylon K. Schum.(sterculiaceae).
- Pusz, W., Plaskowska, E., Yildirim, I., & Weber, R. (2015). Fungi occurring on the plants of the genus Amaranthus L. *Turkish Journal of Botany*, *39*(1), 147-161. Doi:10.3906/bot-1403-106
- Rabi, T., &Bishayee, A. (2009). Terpenoids and breast cancer chemoprevention. *Breast cancer research and treatment*, 115, 223-239. https://doi.org/10.1007/s10549-008-0118-y
- Rajamanikyam, M., Vadlapudi, V., & Upadhyayula, S. M. (2017). Endophytic fungi as novel resources of natural therapeutics. *Brazilian Archives of Biology and Technology*, *60*, e17160542. https://doi.org/10.1590/1678-4324-2017160542
- Ramkisson, S., Dwarka, D., Venter, S., & Mellem, J. J. (2020). In vitro anticancer and antioxidant potential of Amaranthus cruentus protein and its hydrolysates. *Food Science and Technology*, *40*(suppl 2), 634-639. https://doi.org/10.1590/fst.36219
- Rao, A. V., & Sung, M. K. (1995). Saponins as anticarcinogens. *The Journal of nutrition*, 125, 717S-724S. https://doi.org/10.1093/jn/125.suppl_3.717S
- Rit, R., Choudhury, D., Mukherjee, C., & Dutta, S. (2023). Investigation on microfloral association in the roots of Macrotyloma uniflorum (Lam.) Verdc., a medicinally important tropical pulse-crop and their possible applications for crop improvement: A review. *Plant Science Today*, *10*(3), 300-311. https://www.horizonepublishing.com/journals/index.php/PST/index
- Rodriguez, R. J., Woodward, C., Kim, Y. O., & Redman, R. S. (2009). Habitat-adapted symbiosis as a defense against abiotic and biotic stresses. In *Defensive mutualism in microbial symbiosis* (pp. 353-364). CRC Press.
- Safeena, M. I. S., &Sasmina, T. (2018). Effect of biofertilizer (with mycorrhiza inoculum), cowdung, nitrogen and phosphorous on growth and yield of three varieties of amaranth (Amaranthus tricolor L.). http://ir.lib.seu.ac.lk/handle/123456789/3534
- Sagar, A., Dhusiya, K., Shukla, P. K., Singh, A., Lawrence, R., & Ramteke, P. W. (2018). Comparative analysis of production of hydrogen cyanide (HCN) with production of siderophore (SD) and phosphate solubilization (PS) activity in plant growth promoting bacteria (PGPB). *Vegetos*, *31*(2), 130-135.Doi:10.5958/2229-
- Sammour, R. H., Radwan, S. A., & Mira, M. (2012). Genetic diversity in genus Amaranthus: From morphology to genomic DNA. *Res. Rev. Biosci, 6,* 351-360.
- Schulz, B., & Boyle, C. (2006). What are endophytes?. In *Microbial root endophytes* (pp. 1-13). Berlin, Heidelberg: Springer Berlin Heidelberg. https://doi.org/10.1007/3-540-33526-9 1

Sharma, S., & Roy, S. (2015). Isolation and identification of a novel endophyte from a plant Amaranthus spinosus. *International Journal of Current Microbiology and Applied Sciences*, *4*(2), 785-798.http://www.ijcmas.com/

Sindhu, S. S., & Sharma, R. (2019). Amelioration of biotic stress by application of rhizobacteria for agriculture sustainability. *Plant Growth Promoting Rhizobacteria for Sustainable Stress Management: Volume 2: Rhizobacteria in Biotic Stress Management*, 111-168.https://doi.org/10.1007/978-981-13-6986-5_5

Singh, L. P., Gill, S. S., & Tuteja, N. (2011). Unraveling the role of fungal symbionts in plant abiotic stress tolerance. *Plant signaling&behavior*, *6*(2), 175-191.https://doi.org/10.4161/psb.6.2.14146

Siwale, D. R. (2018). Blood glucose lowering effects of leaf extracts of cleome gynandra, amaranthuscruentus and their mixture in normoglycaemic and hyperglycaemic rats (Doctoral dissertation, The University of Zambia).

Stone, J. K., Bacon, C. W., & White Jr, J. F. (2000). An overview of endophytic microbes: endophytism defined. *Microbial endophytes*, 17-44.

Sultana, R., Islam, S. M. N., Shuvo, S. B., Ehsan, G. M. A., Saha, P., Khan, M. M. R., & Rumman, N. (2024). Endophytic bacterium Sphingomonaspanaciterrae NB5 influences soil properties and improves growth, nutrient contents, and yield of red amaranth (Amaranthus tricolor L.). *Current Plant Biology*, *39*, 100372.https://doi.org/10.1016/j.cpb.2024.100372

Torane, R., Gaikwad, S., Khatiwora, E., & Adsul, V. (2017). Comparative estimation of phenol and flavonoid content of medicinally important plant–Amaranthus curentus. *Int J Chem Tech Res*, *10*(4), 306-310.

Uppala, S., Beena, S., Chapala, M., & Bowen, K. L. (2010). Amaranth endophytes and their role in plant growth promotion. *Plant growth promotion by Rhizobacteria for sustainable agriculture. Scientific Publishers, Jodhpur*, 531-537. DOI: 10.13140/RG.2.1.3291.3441

Usunomena, U., & Ngozi, O. P. (2016). Phytochemical analysis and proximate composition of Vernonia amygdalina. *Int. J. Sci. World, 4*(1), 11-14.doi: 10.14419/ijsw.v4i1.5845

Vahabi, K., Johnson, J. M., Drzewiecki, C., &Oelmüller, R. (2011). Fungal staining tools to study the interaction between the beneficial endophyte Piriformosporaindica with Arabidopsis thaliana roots. *J Endocyt Cell Res*, *21*, 77-88.

Wolosik, K., & Markowska, A. (2019). Amaranthus Cruentus taxonomy, botanical description, and review of its seed chemical composition. *Natural Product Communications*, *14*(5), 1934578X19844141. https://doi.org/10.1177/1934578X19844141

Yapa, N., Lakmali, D., Zoysa, D., Silva, K. S., Manawadu, C., Herath, B. M., ... &Bamunuarachchige, C. (2022). Biofertilizers: An Emerging Trend

Sustainability.http://repo.lib.jfn.ac.lk/ujrr/handle/123456789/8984

